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INFLUÊNCIA DE ESTRATÉGIAS DE BIOMODIFICAÇÃO DENTINÁRIA NAS
PROPRIEDADES FÍSICO-QUÍMICAS DE SISTEMAS ADESIVOS

FORTALEZA

2015

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Tese apresentada ao Programa de Pós-Graduação em Odontologia da Faculdade de Farmácia, Odontologia e Enfermagem da Universidade Federal do Ceará, como um dos requisitos para a obtenção do título de Doutor em Odontologia.

Área de Concentração: Clínica Odontológica

Orientador: Prof. Dr. Sérgio Lima Santiago

Co-orientadora: Profa. Dra. Monica Yamauti

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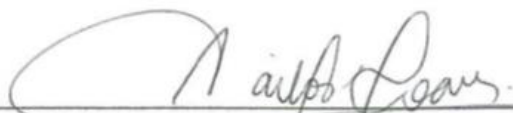
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O presente trabalho não é simplesmente uma coleção de dados científicos, trata-se também de uma declaração de amor aos meus pais, Neri e Inar, e aos meus irmãos, Eugenie, Niskiêr e Pascoal.

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“Pessoas quietas possuem mentes barulhentas.”

(Stephen Hawking)

RESUMO

Proporcionar maior durabilidade aos procedimentos adesivos quando executados sobre a dentina é considerado um desafio. Portanto, o uso de agentes bioativos pode ser uma estratégia promissora para preservar a camada híbrida. Dessa forma, a presente tese foi constituída por cinco capítulos que objetivaram, respectivamente: 1) Avaliar o efeito de soluções de desinfecção cavitária bioativas na resistência de união de sistema adesivo autocondicionante à dentina após termociclagem; 2) Avaliar a efetividade da biomodificação dentinária com epigallocatequina-3-galato (EGCG) na resistência de união de sistema adesivo autocondicionante ao longo do tempo; 3) Avaliar a influência da incorporação de EGCG nas propriedades físico-químicas de um sistema adesivo autocondicionante; 4) Promover o desenvolvimento e a caracterização de partículas poliméricas para liberação controlada de epigallocatequina-3-galato, usando dois tipos de ácido polilático glicólico (PLGA); e 5) Avaliar o efeito da incorporação de micropartículas de PLGA carregadas com EGCG nas propriedades físico-químicas de sistema adesivo convencional de 2 passos. Como abordagens metodológicas foram realizados 5 estudos *in vitro*. No Capítulo 1, a superfície dentinária de 18 dentes foram tratadas com água destilada, fluoreto de sódio (NaF) ou clorexidina (CHX). Espécimes em forma de palito foram confeccionados e submetidos ao teste de resistência de união (μ TBS), após 24 horas e 60.000 ciclos térmicos. No Capítulo 2, a superfície dentinária de 27 dentes foram tratadas com água destilada, EGCG ou CHX. Espécimes em forma de palito foram confeccionados e submetidos ao teste de μ TBS, após 24 h, 6 meses e 12 meses de armazenamento. Para o Capítulo 3, o EGCG foi incorporado diretamente, em concentrações de 0,01% e 0,1%, a um sistema adesivo autocondicionante de 1 passo e, em seguida, foram realizados os testes de sorção (S) e solubilidade (SL), grau de conversão (GC) e resistência flexural (RF). No Capítulo 4, micropartículas foram desenvolvidas a partir de uma técnica de atomização por secagem e, em seguida, foram determinadas o tamanho e a morfologia das partículas, o rendimento, a eficácia de encapsulação e carregamento de EGCG e a liberação da catequina pelas micropartículas. Por fim, no Capítulo 5, além da incorporação direta de EGCG a 0,01% e 0,1%, foram incorporadas também micropartículas de dois tipos de PLGA (PLGA 50:50 e PLGA 75:25) carregadas com EGCG, na concentração de 0,5%, 1% e 2%. Inicialmente foi realizado o ensaio de liberação de EGCG e, em seguida, foram avaliados o GC, a RF, o módulo de elasticidade (ME), a S, a SL e a μ TBS. Os dados obtidos foram submetidos a teste de Análise de Variância e eventuais diferenças estatísticas foram analisadas pelo teste de Holm-Sidak. O nível de significância adotado foi de 5%. Os resultados mostraram que o NaF manteve

a resistência de união após a termociclagem ($p=0,336$) (Capítulo 1). Os grupos tratados com EGCG e CHX não interferiram na resistência de união após 24 h ($p>0,05$), e mantiveram a resistência de união após 6 e 12 meses ($p<0,05$) (Capítulo 2). Não houve diferença estatística entre os valores de WS, GC e RF quando os grupos foram comparados ($p>0,05$). Contudo, a incorporação de EGCG reduziu significativamente a SL ($p<0,05$) (Capítulo 3). Não houve diferença estatística entre os grupos em relação aos valores de GC, ME, S, SL e μ TBS imediata. ($p>0,05$). Os valores de RF foram significativamente elevados pela incorporação de micropartículas poliméricas carregadas com EGCG na concentração de 1% ($p<0,05$). As micropartículas confeccionadas com PLGA 50:50 e PLGA 75:25 apresentaram um padrão de liberação pulsátil (Capítulo 4). A incorporação de micropartículas poliméricas carregadas com EGCG ao sistema adesivo obteve a maior taxa de liberação de EGCG entre todos os grupos ($p<0,05$) (Capítulo 5). Portanto, conclui-se que as estratégias de biomodificação dentinária além de não prejudicarem as propriedades físicas dos sistemas adesivos estudados, podem também manter as interfaces de união mais estáveis ao longo do tempo.

Palavras-chave: Propriedades físicas; Dentina; Catequina; Compostos de flúor; Clorexidina; Sistemas de liberação de medicamento

ABSTRACT

Provide greater durability to the bonding procedures when performed on the dentin is considered a challenge. Therefore, the use of bioactive agents may be a promising strategy to preserve the hybrid layer. Thus, this thesis consisted of five chapters that aimed, respectively: 1) To evaluate the effect of sodium fluoride on the resin-dentin bond strength of a self-etch adhesive after thermal cycles; 2) To evaluate the effectiveness of dentin biomodification with epigallocatechin-3-gallate (EGCG) on the resin-dentin bonds overtime; 3) To evaluate the influence of EGCG incorporation on the physicochemical properties of a methacrylate-based dental adhesive; 4) To develop and characterize of poly (lactide-co-glycolide) acid (PLGA) microparticles for controlled release of epigallocatechin-3-gallate (EGCG), using two types of PLGA; 5) To evaluate the EGCG-load PLGA microparticles incorporation on the physicochemical properties of a two-step etch-and-rinse adhesive system, and the release rate of EGCG. As methodological approaches were performed 5 *in vitro* studies. In the Article 1, dentin surface of 18 teeth were treated with distilled water, 2% chlorhexidine digluconate solution (CHX) or 1.23% sodium fluoride solution (NaF). Bonded sticks were obtained and submitted to bond strength test (μ TBS), after 24 h and 60.000 thermal cycles. In Article 2, dentin surface of 27 teeth were treated with distilled water, or EGCG, or CHX. Bonded sticks were obtained and submitted to bond strength test (μ TBS), after 24 h, 6 and 12 months of storage. To Article 3, EGCG was added to one-step self-etch adhesive, except in control group, to obtain concentrations of 0.01% w/w and 0.1% w/w of EGCG-doped adhesives. Then, water sorption (WS), solubility (SL), degree of conversion (DC) and flexural strength (FS) tests were performed. To Article 4, microparticles were developed from a spray drying technique and then were determined size and particle morphology, yield, efficacy of encapsulation, drug loading and and EGCG release. Finally, in Article 5, beyond the concentrations of 0.01% w/w and 0.1% w/w of EGCG-doped adhesives, were also incorporated EGCG-load polymeric microparticles at 1%. Then, DC, FS, elastic modulus (E), WS, SL, DC, μ TBS and EGCG release. Data were submitted to ANOVA and any statistical differences were analyzed by Holm-Sidak test. The significance level was 5%. The results showed that the NaF maintained bond strength after thermocycling ($p = 0.336$) (Chapter 1). Groups treated with EGCG and CHX did not affect the bond strength after 24 h ($p > 0.05$), and maintained the bond strength after 6 and 12 months ($p < 0.05$) (Chapter 2). There was no statistical difference between the WS, GC and RF values when the groups were compared ($p > 0.05$). However, the incorporation of EGCG reduced SL values ($p < 0.05$) (Chapter 3). There was no significant difference in the DC, E, WS SL and μ TBS values among all groups ($p > 0.05$). FS values were significantly increased by

incorporating polymer microparticles loaded with EGCG at 1% ($p < 0.05$). The microparticles made from PLGA 50:50 and 75:25 PLGA showed a pattern of pulsatile release (Chapter 4). Polymeric microparticles had the highest EGCG release rates, when compared with other groups ($p < 0.05$) (Chapter 5). Therefore, it is concluded that the dentin biomodification strategies besides not impair the physical properties of the adhesive systems may also maintain more stable bonding interface over time.

Key words: Physical properties; Dentin; Catechin; Fluorine compounds; Chlorhexidine; Drug delivery system

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Introdução

1 INTRODUÇÃO

Os sistemas adesivos evoluíram consideravelmente, desde a sua introdução há mais de 50 anos, e influenciaram indubitavelmente os rumos da Odontologia restauradora atual (CARDOSO *et al.*, 2011). Todavia, proporcionar maior durabilidade aos procedimentos adesivos, principalmente quando executados sobre a dentina, ainda é considerado um desafio (DE MUNCK *et al.*, 2005, VAN MEERBEEK *et al.*, 2011).

A dentina é um tecido mineralizado complexo disposto em uma estrutura tridimensional repleta de túbulos, que se estendem da polpa à junção amelo-dentinária (GARBEROGLIO; BRÄNNSTRÖM, 1976; BEDRAN-RUSSO *et al.*, 2014). Aproximadamente, 50% do volume da dentina é composto por minerais, o restante é constituído por colágeno do Tipo I e proteínas não colágenas (30%), e água (20%) (MARSHALL, 1993; MARSHALL *et al.*, 1997; TJÄDERHANE *et al.*, 2012; TJÄDERHANE *et al.*, 2013a). A execução de procedimentos restauradores adesivos sobre o substrato dentinário é considerada crítica devido à sua heterogeneidade e complexa morfologia (PASHLEY; CARVALHO, 1997).

Durante a realização do procedimento adesivo, a superfície e a subsuperfície da dentina são desmineralizadas pelo condicionamento ácido, dos sistemas adesivos convencionais ou pelos monômeros resinosos acídicos, dos sistemas adesivos autocondicionantes (TJÄDERHANE *et al.*, 2013b). Contudo, os monômeros resinosos não são capazes de envolver totalmente as fibrilas colágenas, deixando-as expostas, principalmente, na base da camada híbrida. As áreas incompletamente infiltradas por monômeros dentro da camada híbrida podem ser consideradas o ponto de partida para o processo de degradação dos componentes resinosos e das fibrilas colágenas que compõem a camada híbrida (TAY & PASHLEY, 2003; PASHLEY *et al.*, 2004).

A biodegradação das fibrilas colágenas que não foram completamente envolvidas ocorre através da ação de proteases presentes na matriz extracelular dentinária conhecidas como metaloproteinases da matriz (MMPs) (PASHLEY *et al.*, 2004; HEBLING *et al.*, 2005; OSORIO *et al.*, 2013; TOLEDANO *et al.*, 2013, MAZZONI *et al.*, 2013; MAZZONI *et al.*, 2015) e cisteínas catepsinas (NASCIMENTO *et al.*, 2011; TJÄDERHANE *et al.*, 2013; BEDRAN-RUSSO *et al.*, 2014; VIDAL *et al.*, 2014a). As MMPs são endopeptidases, zinco e cálcio dependentes, que contribuem para a organização e para a mineralização da matriz dentinária (TJÄDERHANE *et al.*, 1998; VAN STRIJP *et al.*, 2003). Essas enzimas são depositadas na dentina de forma inativa, e são conhecidas nessa etapa como zimogênio (VISSE; NAGASE, 2003). A latência das MMPs é decorrente da existência de um propeptídeo contendo

um resíduo de cisteína conservado, unido ao íon zinco do seu sítio catalítico (NAGASE; WOESSNER, 1999). O rompimento da ligação zinco-cisteína pode ocorrer devido a fatores químicos, como o baixo pH dentinário decorrente da aplicação de ácido fosfórico ou de monômeros acídicos (NISHITANI *et al.*, 2006; OSORIO *et al.*, 2011).

Por outro lado, as cisteínas catépsinas (CTs), enzimas pertencentes à família das papaínas, são expressas por odontoblastos humanos e por outras células da polpa, e podem facilmente alcançar os túbulos dentinários e porções mais profundas da dentina (TERSARIOL *et al.*, 2010; TURK *et al.*, 2012). O colágeno íntegro pode ser degradado pela CT-K, em regiões específicas da sua tripla hélice (BRÖMME; OKAMOTO, 1995; HOU *et al.*, 2002), enquanto, o colágeno desnaturado é degradado por vários outros tipos de CTs (LI *et al.*, 2002). Essas enzimas são encontradas tanto em dentina sadia como cariada, contudo, a atividade das CTs é aumentada de forma significativa em dentina profunda e cariada, especialmente em pacientes jovens (NASCIMENTO *et al.*, 2011; VIDAL *et al.*, 2014b). Levando em consideração que a maioria das restaurações é executada em dentina profunda e cariada, é possível que níveis elevados de atividade de CTs também estejam presentes nos túbulos dentinários, durante a execução do procedimento adesivo, o que fatalmente levaria à degradação gradual das fibrilas colágenas da camada híbrida (TJÄDERHANE *et al.*, 2013b). Desta forma, novas abordagens devem ser desenvolvidas para preservar a camada híbrida e garantir a longevidade dos procedimentos adesivos em dentina.

A biomodificação dentinária é uma estratégia biomimética que objetiva melhorar as propriedades biomecânicas e bioquímicas do tecido dentário duro (BEDRAN-RUSSO *et al.*, 2014). A aplicação de agentes bioativos, sintéticos ou naturais, durante os procedimentos adesivos pode modificar a estrutura do colágeno Tipo I, através da formação de ligações cruzadas, aumentando assim a resistência à tração e o módulo de elasticidade do colágeno (AL-AMMAR; DRUMMOND; BEDRAN-RUSSO, 2009). Por outro lado, a maioria das estratégias de biomodificação atuam reduzindo a atividade proteolítica das enzimas que degradam a matriz extracelular, promovendo a remineralização da dentina e prevenindo a ocorrência de lesões cáries (BEDRAN-RUSSO *et al.*, 2014).

Os fluoretos são agentes bioativos amplamente utilizados na prática odontológica devido a sua comprovada ação anticárie (TENUTA; CURI, 2010; TENUTA *et al.*, 2008). Contudo, nos últimos anos o fluoreto de sódio tem chamado a atenção por apresentar também a capacidade de prevenir a degradação do colágeno da camada híbrida através da inibição da atividade da MMP-2 e da MMP-9 (MEI *et al.*, 2012; KATO *et al.*, 2014). Adicionalmente, as soluções fluoretadas podem melhorar as propriedades biomecânicas da dentina através do seu

efeito remineralizador (ITOTA *et al.*, 2002). Desta forma, o uso do fluoreto de sódio pode resultar na estabilização das interfaces adesivas (NERI *et al.*, 2011).

Outro agente bioativo sintético que tem ganhado notoriedade perante a comunidade científica é o digluconato de clorexidina (BRESCHI *et al.*, 2008; BRESCHI *et al.*, 2009). Além de ser um excelente agente antimicrobiano, a clorexidina também é um potente inibidor de MMP-2, MMP-8 e MMP-9 (GENDRON *et al.*, 1999; ZHOU *et al.*, 2011; TOLEDANO *et al.*, 2012), e CT-B, CT-L e CT-K (SCAFFA *et al.*, 2012). O digluconato de clorexidina pode atuar de duas maneiras para inativar as MMPs: através da quelação com os íons zinco ou mediante a interação com grupos sulfidril e/ou cisteínas presentes nos sítios catalíticos das MMPs ativas (GENDRON *et al.*, 1999). O emprego do digluconato de clorexidina tem apresentado resultados promissores em estudos *in vivo* e *in vitro*, confirmando a manutenção da resistência de união ao longo do tempo (CARRILHO *et al.*, 2007a; CARRILHO *et al.*, 2007b; LOGUERCIO *et al.*, 2009; CAMPOS *et al.*, 2009; STANISLAWCZUK *et al.*, 2009; BRESCHI *et al.*, 2010; SANTIAGO *et al.*, 2013). Todavia, por apresentar reações inflamatórias e necrose tecidual (DE SOUZA *et al.*, 2007; FARIA *et al.*, 2007), tem-se procurado substâncias mais biocompatíveis e provenientes de produtos naturais (SANTIAGO *et al.*, 2013).

O epigallocatequina-3-galato (EGCG) é o principal polifenol encontrado no chá verde (*Camellia sinensis*), correspondendo a mais de 65% das catequinas encontradas nesse chá (DEMEULE *et al.*, 2000). Esse polifenol despertou grande interesse científico devido ao seu potencial antimutagênico e anticancerígeno (CAO & CAO, 1999; GARBISA *et al.*, 2001; SEN *et al.*, 2009). O epigallocatequina-3-galato apresentou baixa toxicidade e propriedades anti-inflamatórias quando em contato com células pulpares e possui comprovada capacidade de inibir a ação de MMP-2 e MMP-9 (DEMEULE *et al.*, 2000; GARBISA *et al.*, 2001; DELL'AICA *et al.*, 2007; VIDAL *et al.*, 2014), e CTs (DEVIKA; PRINCE, 2008, VIDAL *et al.*, 2014). Estudos prévios demonstraram que o epigallocatequina-3-galato se liga ao colágeno através de pontes de hidrogênio, e promove o aumento do número de ligações cruzadas inter e intra-fibrilares, prevenindo, portanto, o livre acesso das collagenases aos sítios específicos de degradação na cadeia de colágeno (JACKSON *et al.*, 2010; VIDAL *et al.*, 2014; BEDRAN-RUSSO *et al.*, 2014).

O epigallocatequina-3-galato tem sido utilizado na Odontologia adesiva em diferentes estratégias (pré-tratamento dentinário e incorporado ao sistema adesivo) e em diferentes concentrações (0,01%, 0,02%, 0,03%, 0,1%, 0,5%), resultando na manutenção da resistência de união após 6 meses de armazenamento dos espécimes (DU *et al.*, 2012, SANTIAGO *et al.*, 2013). Embora a incorporação de epigallocatequina-3-galato aos sistemas adesivos tenha

demonstrado resultados promissores, há uma preocupação em relação à sua biodisponibilidade dentro da camada híbrida. Pallan *et al.* (2012) observaram uma alta taxa de liberação de EGCG, nas primeiras 24 horas de armazenamento em água destilada, seguida de uma redução significativa na taxa de liberação até o 28º dia. Portanto, o tempo de permanência do EGCG na camada híbrida provavelmente foi curto devido solubilidade da molécula em água (PALLAN *et al.*, 2012). Esse inconveniente pode ser contornado a partir de métodos de liberação controlada de fármacos (LIANG *et al.*, 2005; GAIGNAUX *et al.*, 2012).

A liberação lenta e duradoura do epigalocatequina-3-galato pode ser obtida a partir do seu encapsulamento em micropartículas de ácido poli-láctico-co-glicólico (PLGA) (ANDERSON & SHIVE, 1997; MATSUMOTO *et al.*, 2005). O PLGA é considerado o material padrão-ouro na liberação controlada de fármacos, devido a sua incomparável biodegradação e biocompatibilidade (JAIN, 2000; TAMBER *et al.*, 2005). A liberação dos fármacos ocorre a partir do contacto das micropartículas com os fluidos orgânicos aquosos (GAIGNAUX *et al.*, 2012). A água penetra nas micropartículas, promove a degradação da cadeia do polímero e a dissolução do fármaco. Uma vez dissolvido, o fármaco começa a difundir-se para o fluido circundante através da rede de polímero degradado (LI; ROUAND; PONCELET, 2008). Apesar dos benefícios comprovados da liberação controlada de fármacos nas ciências médicas, pouco se sabe a respeito da sua utilização em Odontologia adesiva.

Nesse contexto, o uso de agentes bioativos pode significar um importante avanço na tentativa de se obter maior estabilidade dos componentes da camada híbrida, tornando os procedimentos adesivos menos susceptíveis à degradação.

Proposição

2 PROPOSIÇÃO

A presente tese será apresentada em capítulos, tendo como objetivos:

Capítulo 1: Avaliar a influência do fluoreto de sódio na resistência de união de sistema adesivo autocondicionante à dentina após termociclagem.

Capítulo 2: Avaliar a efetividade da biomodificação dentinária com epigallocatequina-3-galato na resistência de união de sistema adesivo autocondicionante ao longo do tempo.

Capítulo 3: Avaliar a influência da incorporação de epigallocatequina-3-galato nas propriedades físico-químicas de um sistema adesivo autocondicionante.

Capítulo 4: Promover o desenvolvimento e a caracterização de partículas poliméricas para liberação controlada de epigallocatequina-3-galato, usando dois tipos de PLGAs.

Capítulo 5: Avaliar o efeito da incorporação de micropartículas de ácido polilático coglicólico carregadas com epigallocatequina-3-galato nas propriedades físico-químicas de sistema adesivo convencional de 2 passos, e a taxa de liberação de epigallocatequina-3-galato.

Capitulos

3 CAPÍTULOS

REGIMENTO INTERNO

A presente tese está baseada no Artigo 46 do Regimento Interno do Programa de Pós-graduação em Odontologia da Universidade Federal do Ceará que regulamenta o formato alternativo para dissertações de Mestrado e teses de Doutorado e permite a inserção de artigos científicos de autoria ou coautoria do candidato. Por se tratar de pesquisas envolvendo seres humanos, os projetos de pesquisa referentes aos trabalhos desenvolvidos nos capítulos 1, 2 e 5 foram submetidos à apreciação do Comitê de Ética em Pesquisas da Universidade Federal do Ceará, tendo sido aprovados (Anexos B, D e G). Desta forma, a tese é composta por cinco artigos, sendo um artigo publicado e quatro a serem submetidos para publicação em periódicos científicos, conforme descrito abaixo:

Capítulo 1: Does sodium fluoride maintain resin–dentin bond strength after thermal stressing? O presente artigo será submetido à publicação na revista “*European Journal of Oral Sciences*”.

Capítulo 2: Influence of dentin biomodification with epigallocatechin-3-gallate on the bond strength of self-etch adhesive: twelve-month results. O presente artigo será submetido à publicação na revista “*Journal of Biomedical Materials Research Part B: Applied Biomaterials*”.

Capítulo 3: Physicochemical properties of a methacrylate-based dental adhesive incorporated with epigallocatechin-3-gallate. O presente artigo foi submetido e publicado na revista “*Brazilian Dental Journal*”.

Capítulo 4: Development and characterization of PLGA microparticles for controlled release of epigallocatechin-3-gallate. O presente artigo será submetido à publicação na revista “*International Journal of Pharmaceutics*”.

Capítulo 5: Physicochemical properties and pattern of drug release of etch-and-rinse adhesive system incorporated with catechin-loaded polymeric microparticles. O presente artigo será submetido à publicação na revista “*Journal of Dentistry*”.

Capítulo 1

3.1 CAPÍTULO 1

Does Sodium fluoride maintain resin–dentin bond strength after thermal stressing?

Running title: Sodium fluoride maintain MTBS

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ABSTRACT

This study evaluated the effect of sodium fluoride on the resin-dentin bond strength of a self-etch adhesive after thermal cycles. Eighteen human third molars were prepared to expose a flat dentin surface and were divided into 3 groups ($n=6$) according to the cavity cleaning solutions, as follows: distilled water, 2% chlorhexidine digluconate solution (CHX) or 1.23% sodium fluoride solution (NaF). Solutions were rubbed for 60 seconds on dentin surfaces, followed by bonding with Clearfil SE Bond, and 5-mm-thick resin crown build-up. Bonded teeth were stored in distilled water for 24 hours and then longitudinally sectioned to obtain bonded sticks. Half of the specimens were immediately tested in tension at 0.5 mm/min, while the remaining specimens were tested after 60.000 thermal cycles. Data were analyzed using Two-way ANOVA and the Holm-Sidak method. There was no significant difference between the groups after 24 hours ($p>0.05$). Thermocycling resulted in significant bond strength reduction for distilled water and CHX ($p<0.05$). When 24 hours values were compared to thermocycling, NaF maintained its bond strength ($p>0.05$), while significant reductions in bond strength were observed for distilled water and CHX ($p<0.05$). Pretreatment with NaF preserved the bonding of Clearfil SE Bond to dentin after 60.000 thermal cycles.

Key words: Dentin-Bonding Agents; Sodium Fluoride; Self-etch adhesive; Dentin bond strength.

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INTRODUCTION

Self-etch adhesives contain acidic monomers that simultaneously dissolve and infiltrate the smear layer, smear plugs and hydroxyapatite to generate a hybrid layer (1). However, the use of self-etching adhesive systems on smear layers has been considered a controversial matter (2). Self-etching primers may not be able to penetrate through thick smear layers (3). The acidity of the primer could also be buffered by the mineral components of the smear layer to the extent that the potential for primer penetration into the underlying sound dentin might be reduced, resulting in gaps in adhesive restorations (3). Thus, the pretreatment of the smear layer can be an alternative for improving the longevity of the restorative procedures (4,5).

The use of cavity cleaning solutions prior to restorative procedures may totally or partially remove the smear layer (6). Chlorhexidine digluconate is a non-demineralizing agent that acts via the mechanical action of washing and scrubbing, which may promote changes in the smear layer without exposing opened dentinal tubules (6). It has been widely used as an antimicrobial agent due to its broad spectrum of action against Gram-negative and Gram-positive bacteria, particularly *Streptococcus mutans* (7). Additionally, chlorhexidine has shown to preserve dentin bond strength via the inhibition of matrix metalloproteinases (MMPs) and cysteine cathepsins (8-11).

On the other hand, when dentin is cleaned with 1.23% acidic sodium fluoride solution, the smear layer and smear plug are completely removed, exposing dentinal tubules (6). Thus, the removal of smear layer provide direct contact of sel-etch primers with dentin, and can influence the formation of compact hybrid layer (3). Recently, sodium fluoride has demonstrated its efficacy on the inhibition of MMP action, which suggests its potential to preserve the resin-dentin interface (12, 13). Although fluoride solutions could present beneficial effects in preserving bonded interfaces, little is known about the use of 1.23% sodium fluoride solution, as a cavity cleanser associated with self-etch adhesives.

The aim of this study was to evaluate the influence of sodium fluoride on the resin-dentin bond strength of a self-etching adhesive after thermal cycles. The tested null hypotheses were that 1) there will be no differences in immediate bond strength caused by different pretreatments and 2) there will be no differences in the bond strength between the 24-h period and after thermocycling for all groups.

MATERIALS AND METHODS

Eighteen unerupted third molars were collected after patients' informed consents had been obtained under a protocol (#35/12) that was reviewed and approved by the local Ethics Committee. Selected teeth were stored in 0.01% thymol solution and used within one month after extraction. Occlusal enamel was removed using a #120-grit silicon carbide (SiC) paper mounted to an electric polishing machine (Aropol 2V; Arotec, São Paulo, SP, Brazil) to expose a flat coronal dentinal surface. The dentinal surface was prepared with #600-grit SiC paper under copious water for 60 s to standardize the smear layer.

The teeth were randomly allocated using Excel software (Excel 2003, Microsoft Corporation, Redmond, WA, USA) into 3 groups ($n=6$) according to the pre-treatment solution used prior to adhesive application. Dentinal surfaces of all teeth were dried with oil/water-free air for 10 s and treated with 10 μ L of distilled water (pH 7.40), 2% chlorhexidine digluconate solution (pH 7.55) (CHX) (FGM, Joinville, SC, Brazil - batch #210211) or 1.23% sodium fluoride solution (pH 3.60) (NaF). The solutions were rubbed for 60 s, and the excess of each solution was removed with absorbent paper.

The two-step self-etch adhesive system, Clearfil SE Bond (Kuraray Medical, Tokyo, Japan), was applied according to the manufacturer's instructions (Table 1) and light-cured with Variable Intensity Polymerizer, VIP Junior (500 mW/cm², Bisco Inc., Schaumburg, IL, USA). Five 1-mm-thick resin composite increments were built up (Filtek Z250; 3M ESPE, St. Paul, MN, USA - batch #1117600319). Each increment was light cured for 20 s. The bonded teeth were stored in distilled water at 37°C for 24 h.

After storage, the prepared teeth were longitudinally sectioned in both “x” and “y” directions across the bonded interface using a diamond saw in a Labcut 1010 (Extex, Enfield, CT, USA) under water cooling to obtain sticks with cross-sectional area of approximately 1.0 mm². The cross-sectional area of each stick was measured with a digital caliper (Absolute Digimatic, Mitutoyo, Tokyo, Japan) to the nearest 0.01 mm and recorded for the subsequent calculation of bond strength values. Half of the sectioned sticks for each tooth were randomly selected using the Excel software (Excel 2003, Redmond, WA, USA) and immediately tested under tension. The remaining sticks were submitted at 60,000 thermal cycles (THE-1100; SD Mechatronik GMBH, Feldkirchen-Westerham, Germany) in distilled water at $5 \pm 2^\circ\text{C}$ and $55 \pm 2^\circ\text{C}$ baths, with a dwell time of 60 s in each bath. Thermocycling took approximately 3 months (approximately 20,000 cycles per month).

For the microtensile test, each bonded stick was attached with cyanoacrylate glue (Super Bonder Gel, Loctite, São Paulo, SP, Brazil) to a modified Geraldeli testing apparatus (Odeme Biotechnology, Joaçaba, SC, Brazil) and subjected to a tensile force at 0.5 mm/min in a universal testing machine (Instron 3345; Instron Inc., Canton, MA, USA). The load at fracture was used to calculate bond strength (MPa).

The failure mode was evaluated using a stereoscope at 80X magnification (StereoZoom® Leica S8 APO, Leica Microsystems, Wetzlar, Germany) and classified as cohesive when the fracture occurred exclusively within the dentin (CD) or resin composite (CR); adhesive (A) when the failure was at the dentin/resin interface or mixed (M) when two modes of failure (adhesive and cohesive) occurred simultaneously.

A Shapiro-Wilk test was applied to all groups to analyze the normal distribution of errors and the Bartlett test was applied to test for homoscedasticity. After a normal distribution was confirmed, data were analyzed with a two-way ANOVA (pretreatments and aging) and the Holm-Sidak method was used for *post hoc* comparisons. Statistical procedures were performed with the Sigmastat 3.5 for Windows (Systat Software Inc., San Jose, CA, USA) statistical program software. The level of significance was set at $p < 0.05$. Teeth were used as a statistical unit and the number of prematurely debonded specimens was recorded, although this was not included in the analysis.

RESULTS

Bond strength results are shown in Table 2. The results were affected by treatment ($p=0.020$; $F=4.615$) and aging ($p<0.001$; $F=13.161$). Interactions were not statistically significant ($p=0.815$; $F=0.206$).

After 24 h of storage, no significant difference was observed among the bond strength values of all groups ($p>0.05$). After thermocycling, NaF had the highest bond strength values, when compared with CHX ($p=0.025$) and distilled water ($p=0.048$). When 24-h values were compared to thermocycling, NaF maintained its bond strength ($p=0.129$), whereas significant reductions in bond strength were observed for distilled water ($p=0.029$) and CHX ($p=0.025$) (Table 2).

Table 3 summarizes the percentage of failure modes of the debonded specimens. Most failures were mixed in all tested groups for all tested conditions. There was an increase in the number of premature failures for all groups, mainly to distilled water and CHX after thermocycling.

DISCUSSION

The smear layer represent a great challenge for the interaction of adhesive systems and prepared tooth surfaces (14). Once that the thickness and structure of smear layer may interfere in the demineralizing potential of adhesives systems, thus decreasing the resin-dentin bond strength (15-17). However, in the present study the presence of a smear layer did not seem to affect the immediate bond strength of self-etch adhesive to dentin (Table 2), corroborating with others studies (18-20). This fact could be explained due to the use of 600-grit SiC paper to standardize smear layer, resulting in the formation of thin smear layer (thickness $\approx 1 \mu\text{m}$) (21, 22). When Clearfill SE Bond was applied on thin smear layer besides to demineralize and completely infiltrate in the smear layer (hybridized smear layer) also forms a true hybrid layer with the superficial peritubular dentin (3).

Previous studies have reported that the application of chlorhexidine prior to the self-etch adhesive had no adverse effects on immediate adhesive bonds in dentin (23-27). The present study results corroborate the results from previous ones. Nevertheless, other researchers have suggested that such use of chlorhexidine prior to the application of self-etch adhesives should be avoided because of potential interactions among chlorhexidine and the adhesive components (28-30). Chlorhexidine may decrease wettability and the level of dentin conditioning of self-etching adhesives, which could interfere with the stability of the hybrid layer (28).

The results of the current study show that the application of NaF did not affect the immediate resin-dentin bond strength in the same way that the distilled water and CHX. Thereby, failed to reject the first null hypothesis. Although, sodium fluoride solution with low pH (pH= 3.6) removes the smear layer and exposes dentin tubules (6), the thickness of true hybrid layer formed on dentin without smear layer is similar to dentin with thin smear layer, approximately $0.5 \mu\text{m}$ thick (3). According to TAY *et al.* (20), the immediate bond strength between mild-self etch adhesive system and dentin with and without smear layer were not significantly different.

On the other hand, the second null hypothesis was rejected because significant reductions in bond strength were observed for distilled water and CHX when the 24-h were compared to thermocycling (Table 2). The stability of the bonded interface relies on the creation of a compact and homogenous hybrid layer (10). Nevertheless, mild self-etch adhesives, as the one used in this study, applied on thin smear layer increased the formation of an interfacial gap (18), leading to the possible separation of the hybridized smear layer from the true hybrid layer (20). In a recent study, a mild self-etch adhesive failed predominantly under the hybrid layer

after aging, which may have been the result of insufficient penetration of self-etch primers in the dentin (32). SABOIA *et al.* (33) speculated that during thermocycling the repetitive contraction/expansion stresses generated at the hybrid layer synergistically enhanced the hydrolytic degradation of the adhesive components and collagen fibrils, thereby weakening the physical properties of the resin–dentin bond such as bond strength.

Specimens pretreated with NaF solution maintained its bond strength after thermocycling. Pretreatment of dentin surfaces with acidic solutions has been also recommended to eliminate the smear layer and enable a direct contact between the adhesive resin and dentin (34). However, main challenge for demineralizing agents is to dissolve the smear layer without demineralizing the tooth surface too profoundly (5). ITOTA *et al.* (35) showed that the bond strength of mild-self adhesive system on demineralized dentin is lower than mineralized dentin. The application of 1.23% sodium fluoride solution removes the smear layer and does not promote significant changes in the dentinal surface (6). The preservation of hydroxyapatite will also provide calcium for chemical bonding to the functional self-etch monomers (5). Within the limitations of this study, we can speculate that smear layer removal using a low pH (3.6) NaF provided better attachment between the resin monomers and dentinal components, thereby establishing a more stable hybrid layer and providing resistance to hydrolytic degradation.

Low pH of mild self-etch adhesives ($\text{pH} \approx 2$) have shown to activate dentinal enzymes (36-38). Matrix metalloproteinases (MMPs) and cysteine cathepsins may be responsible for the degradation of collagen fibrils exposed at the adhesive interface (9, 10, 31, 38, 39). The use of chlorhexidine as an MMP-inhibitor was first described by HEBLING *et al.* (40) Subsequently, several authors have reported that the use of chlorhexidine associated with total-etch adhesive systems promoted the maintenance of bond strength over time (8-11). However, when chlorhexidine was used prior to the application of two-step self-etching adhesives did not maintain the resin-dentin bond strength after thermocycling (31).

For a long time, fluoride ions were used to protect hard tissues due to a favorable balance in the demineralizing and remineralizing of the enamel and dentin (41). However, recent studies have suggested that sodium fluoride has the potential to prevent collagen degradation as an MMP-inhibitor (12, 13). Purified forms of both MMP-2 and MMP-9 were completely inhibited by 200 ppm F^- (12). It is speculated that the fluoride concentration of NaF (12,300 ppm F^-) would also be able to inhibit dentinal MMPs (12). Nevertheless, little is known about the molecular mechanisms by which fluoride ions block gelatinolytic activities (12,13). Considering that MMPs are zinc and calcium-dependent, and ion F is highly electronegative, it seems logical that excess F could make these cations unavailable to participate in the catalytic

process, according to KATO *et al.* (12). The inhibition of MMPs-2 and -9 by sodium fluoride is reversible at lower F concentrations, but irreversible at higher (5,000 ppm) of F concentrations (12). This inhibitory activity of NaF associated with the the possible formation of a more homogeneous hybrid layer provided the maintenance of resin-dentin bond strengths. Thus, further studies should be conducted to confirm the potential of sodium fluoride in inhibiting dentinal MMPs, as well as its role in the preservation of collagen and maintenance of bond strength.

Within the limitations of this in vitro experimental model was observed that dentin pretreatment with acidic sodium fluoride could be useful to improve the bond stability of the bonding procedures when associated to two-step self-etch adhesive.

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CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

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TABLES

Table 1 – Adhesive system used, batches, chemical compositions and application protocols.

Product	Composition	Manufacturer (#Batch n°)	Application mode
Clearfil SE Bond	Primer: MDP, HEMA, water, photoinitiator Bond: MDP, BisGMA, HEMA, TEGDMA, Hydrophobics dimethacrylates, photoinitiator	Kuraray Medical, Tokyo, Japan (#51476)	Apply primer for 20 s, gently air-dry; apply bond and light cure for 10 s.
Abbreviations: BisGMA: bisphenol-A-diglycidylmethacrylate; HEMA: hydroxyethylmethacrylate; MDP: 10-methacryloyloxi-decyl-phosphate; TEGDMA: triethylene-glycol-dimethacrylate.			

Table 2 – Bond strengths according to the pretreatment solution. The results are in MPa \pm SD (*).

Groups	Clearfil SE Bond	
	24 hours	60.000 thermocycles
Distilled water	30.98 \pm 5.96 (33) ^{A,a}	22.96 \pm 6.88 (30) ^{B,b}
CHX	30.18 \pm 4.06 (22) ^{A,a}	21.91 \pm 4.37 (26) ^{B,b}
NaF	35.60 \pm 6.52 (28) ^{A,a}	30.17 \pm 4.23 (26) ^{A,a}

Identical superscript letters indicate no statistical significance between values. Different superscript upper case letters (analysis in columns) and different superscript lower case letters (in rows) indicate statistically significant differences

(*) corresponds to the number of sticks tested per group in each period.

Table 3 – Distributions of the mode of fracture according to the study groups. Relative percentages are in ().

Clearfil SE Bond										
Groups	24 hours					60.000 thermocycles				
	A	M	CR	CD	PF	A	M	CR	CD	PF
Distilled water	6 (16)	27 (73)	0 (0)	4 (11)	0 (0)	8 (20)	22 (55)	4 (10)	2 (5)	4 (10)
CHX	4 (14)	18 (64)	3 (11)	2 (7)	1 (4)	6 (18)	20 (61)	2 (6)	0 (0)	5 (15)
NaF	3 (8)	25 (69)	2 (6)	5 (14)	1 (3)	3 (8)	23 (66)	5 (14)	1 (4)	3 (8)

A: adhesive failure; M: mixed failure; CR: cohesive failure in resin; CD: cohesive failure in dentin; PF: premature failure

Capítulo 2

3.2 CAPÍTULO 2

Influence of dentin biomodification with epigallocatechin-3-gallate on the bond strength of self-etch adhesive: twelve-month results

Running title: Influence of catechin on bond strength to dentin

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ABSTRACT

Aims: To evaluate the effectiveness of dentin biomodification with epigallocatechin-3-gallate (EGCG) on the resin-dentin bonds overtime. **Materials and Methods:** Twenty seven extracted human third molars were prepared to expose a flat dentin surface and divided into 3 groups. Dentin surfaces were dried and treated with 20 μ L aliquots of either distilled water (control); 2% chlorhexidine digluconate solution (CHX) or 0.1% EGCG aqueous solution. Solutions were rubbed for 60 s followed by bonding with Adper Easy One, and 5-mm-thick resin crown build-up. Bonded teeth were stored in distilled water for 24 h and then longitudinally sectioned to obtain bonded sticks. One-third of the specimens were immediately tested in tension at 0.5 mm/min, while the remaining specimens were tested after six and twelve months of storage in distilled water at 37°C. Data were analyzed with Two-way ANOVA and Holm-Sidak method. **Results:** After 24 h of storage, mean bond strength values were not significantly different among all groups ($p>0.05$). Bond strengths of EGCG and CHX remained stable after 6 and 12 months. ($p>0.05$). **Conclusions:** Pretreatment with EGCG or CHX preserved the bonding of Adper Easy One to dentin after six and twelve months of storage.

Keywords: Catechin; Dentin; Matrix metalloproteinases; Dental adhesive; Resin-dentin bond

INTRODUCTION

The most common cause of replacement of resin composite restorations is secondary caries.^{1,2} This fact occurs due to gradual degradation of the components of the hybrid layer, such as collagen fibrils, resulting in the loss of resin-dentin bond strength and bacterial microleakage.^{3,4} Promising strategies are being developed to reinforce the type I collagen and reduce their degradation, further to prevent dental caries.⁵ Dentin pretreatment with bioactive agents, synthetic or natural, may produce a strong and long lasting tooth-biomaterial interface.⁵⁻⁷

Chlorhexidine has been widely used as an antimicrobial agent due to its broad spectrum of action in Gram-negative bacteria and Gram-positive, particularly *Streptococcus mutans*.⁸ However, there is a concern about the use of chlorhexidine as bioactive agent when adhesive procedures are employed.⁹⁻¹¹ Pretreatment of dentin surfaces with this agent may reduce the bond strength of self-etching adhesives to dentin.¹¹ Therefore, chlorhexidine should be used with caution when self-etch adhesives are selected.

Tea polyphenols have been demonstrated to be effective as antimicrobial compound against a variety of pathogenic microorganisms.¹²⁻¹⁴ Interestingly, among the catechin family, epigallocatechin-3-gallate exhibits outstanding bactericidal efficacy due to the gallate group (gallic acid ester),¹⁴ while other catechins do not have this effect.^{15,16} Epigallocatechin-3-gallate is effective in reducing acid production in dental plaque and on the growth of *Streptococcus mutans* particularly due to suppression of specific virulence factors associated with its acidogenicity and acidity.¹⁷⁻¹⁹ Epigallocatechin-3-gallate also prevents episodes of dental demineralization, inhibiting the progress of caries in dentin.²⁰

Additionally, some catechins, such as epigallocatechin-3-gallate, were able to stabilize the collagen chain.²¹⁻²³ Epigallocatechin-3-gallate increases the number of collagen fibrils crosslinks and reduces the biodegradation of collagen.²³ Recent studies showed that the use of epigallocatechin-3-gallate associated with etch-and-rinse adhesive systems was effective in preserving resin-dentin bond strength up to 6 months.^{7,24} Although epigallocatechin-3-gallate is being extensively studied, their use associated with self-etch adhesives was not previously investigated.

The objective of this study was to evaluate the effectiveness of dentin biomodification with epigallocatechin-3-gallate on the resin-dentin bonds overtime. The tested null hypotheses were that 1) there will be no differences in the immediate bond strength caused by the different pretreatments; and 2) there will be no differences in bond strength between storage periods for all groups.

MATERIALS AND METHODS

Twenty seven unerupted third molars were collected after the patients' informed consent had been obtained under a protocol reviewed and approved by the local Ethics Committee (#103/11). Selected teeth were stored in 0.1% thymol solution and used within one month after extraction. Occlusal enamel was removed using a #120-grit silicon carbide (SiC) paper mounted to an electric polishing machine (Aropol 2V; Arotec®, São Paulo, SP, Brazil) to expose a flat coronal dentin surface. The dentin surface was prepared with #600-grit SiC paper under copious water for 60 seconds to standardize the smear layer.

The teeth were randomly allocated by the Excel software (Excel 2003, Microsoft Corporation, One Microsoft Way, Redmond, WA, USA) into 3 groups ($n=9$) according to the pretreatment solution prior to adhesive application. Dentin surfaces of all teeth were dried with oil/water-free air for 10 s and treated with 20 μ L of either distilled water (control), 2% chlorhexidine digluconate solution (CHX) (FGM, Joinville, SC, Brazil - batch #210211) or 0.1% epigallocatechin-3-gallate aqueous solution (EGCG) (Sigma-Aldrich, Saint Louis, USA - batch #029K 1228). The solutions were rubbed on the surface with a microbrush for 60 s, and excess of each solution removed with absorbent paper.

The one-step self-etch adhesive system - Adper Easy One (3M ESPE, St. Paul, MN, USA) – was applied according to manufacturer's instructions (Table 1) and light-cured with Variable Intensity Polymerizer – VIP Junior (Bisco Inc., Schaumburg, IL, USA) with power density of 500 mW/cm^2 . Five increments of 1 mm-thick of composite resin were build-up on the bonded surface (Filtek Z250; 3M ESPE, St. Paul, MN, USA - batch #1030600122) and each increment was light-cured for 20 s. The bonded teeth were stored in distilled water at 37°C for 24 h.

After storage, the bonded teeth were longitudinally sectioned in both “x” and “y” directions across the bonded interface using a diamond saw in a Labcut 1010 (Extec, Enfield, CT, USA) under water cooling to obtain bonded sticks with cross-sectional area of approximately 1.0 mm^2 . The cross-sectional area of each stick was measured with a digital caliper (Absolute Digimatic, Mitutoyo, Tokyo, Japan) to the nearest 0.01 mm and recorded for subsequent calculation of bond strength values.

Sticks from each tooth were randomly allocated by the Excel software (Excel 2003, Microsoft Corporation, One Microsoft Way, Redmond, WA, USA) and assigned to three storage times: 24 hours, 6 months and 12 months. All sticks were stored in distilled water at 37°C and the storage solution was changed every two weeks. For microtensile test, each bonded stick was

attached with cyanoacrylate glue (Super Bonder Gel, Loctite, São Paulo, SP, Brazil) to a modified Geraldeli testing apparatus (Odeme Biotechnology, Joaçaba, SC, Brazil) and subjected to a tensile force at 0.5 mm/min in a universal testing machine (Instron 3345; Instron Inc., Canton, MA, USA). The load at fracture was used to calculate bond strength (MPa).

The failure mode was evaluated using a stereoscope at 80X magnification (StereoZoom® Leica S8 APO, Leica Microsystems, Wetzlar, Germany), and classified as cohesive when fracture occurred exclusively within dentin (CD) or resin composite (CR); adhesive (A) when was at the dentin/resin interface, or mixed (M) when two modes of failure (adhesive and cohesive) occurred simultaneously.

Statistical procedures were performed with the Sigmastat 3.5 (Systat Software Inc., San Jose, CA, USA) for Windows statistical program software. A Shapiro-Wilk test was applied to all groups to analyze the normal distribution of errors and the Barlett test for the homoscedasticity. Bond strength values were statistically analyzed with Two-way ANOVA (pretreatment and storage) and Holm-Sidak method was used for *post hoc* comparisons. Statistical significance was set at $p < 0.05$. Teeth were used as a statistical unit and the number of prematurely debonded specimens was recorded, although this was not included in the analysis.

RESULTS

The results were affected by treatment ($p < 0.001$; $F = 17.562$), but not by storage time ($p = 0.295$; $F = 1.242$). Interactions were statistically significant ($p = 0.01$; $F = 3.573$) (Table 2).

After 24 h storage, there was no significant difference among the bond strength values of all groups ($p > 0.05$). At 6 months and 12 months, bond strength mean values of EGCG, and CHX groups were not statistically different ($p > 0.05$) and they were significantly higher than those of distilled water (control, $p < 0.05$). In addition, the resin-dentin bond strength was preserved up to 12 months after application of EGCG and CHX ($p > 0.05$).

Most of failures were mixed in all tested groups at all storage times. At 24 h, adhesive failures were more observed than cohesive failures (resin and dentin) for specimens of CHX group. The number of adhesive failures increased with ageing, except in CHX group (Table 3).

DISCUSSION

Dentin pretreatment with EGCG or CHX did not affect immediate bond strength as the control group; therefore failed to reject the first null hypothesis. On the other hand, the second null hypothesis was rejected since that only experimental solutions used as pretreatment were effective in preserving the bond strength after 6 and 12 months of water storage.

The use of chlorhexidine as bioactive agent prior to application of self-etch adhesives is controversial. Some studies^{9-11,25} suggested that such use should be avoided because there could be interactions between chlorhexidine and the adhesive components. These interactions could decrease their wettability and the level of dentin conditioning.⁹ However, the results of the present study demonstrated that chlorhexidine application prior to bonding procedures did not present adverse effects on immediate resin-dentin bond strength for self-etch adhesive (Table 2) and this data is confirmed by other studies.²⁶⁻³⁰

Dentin pretreatment with epigallocatechin-3-gallate has also been shown not to be detrimental to bond strengths to dentin.^{7,24} In the study published by Du et al.²⁴ there were no difference between degrees of conversion among concentrations of epigallocatechin-3-gallate used, although these authors speculated that higher concentration of epigallocatechin-3-gallate, e.g., 0.03% could disturb the polymerization of the adhesive, thus affecting bond strength. However, EGCG at 0.1% was used in the present study and did not significantly affect bond strength (Table 2). Neri et al.³¹ demonstrated that incorporation of epigallocatechin-3-gallate at 0.1% in Adper Easy One did not present adverse effects on degree of conversion. Probably, the EGCG at 0.1% have not been entrapped within the linear chains after curing without interfering with monomers conversion and bond strength.

The mode of failure of the tested groups confirmed the bond strength results at 24 h, as the control, CHX and EGCG groups showed almost similar results, with an increased percentage of mixed failures (Table 3). The results of the control group corroborated with others studies,^{30,32} which showed high rates of mixed failure with Adper Easy One. This was explained by the micromechanical as well as chemical interaction of the functional monomer included in this adhesive with the hydroxyapatite crystals that remain at the surface.³²

Enzymatic degradation of the collagen matrix by host derived enzymes plays a significant role in the destruction of the bonded interface.³³ Matrix metalloproteinases (MMPs) and cysteine cathepsins have been identified in dentin in inactive form.^{34,35} However, some researches have reported that the low pH of self-etch adhesive systems (pH 2.3-5) could activate

dentin enzymes.^{34,36,37} The self-etch adhesive used in this study has a pH value of 2.4 and so, is capable of enhancing dentin proteolytic activity without denaturation of the enzymes. It has been suggested that the action of these enzymes may be responsible for the degradation of collagen fibrils exposed at the adhesive interface.^{6,34,35} After 6 and 12-month storage in distilled water at 37 °C, it was observed the reduction of the bond strength values for the control group (Table 2). These data corroborate the findings of other *in vitro* studies.^{28,30,38} Probably, the action of MMPs and cysteine cathepsins has determined to reduce the bond strength of the control group as well as the increase of number of adhesive and premature (Table 3).

On the other hand, the use of bioactive substances that inhibit the enzymes may help to preserve the hybrid layer and increase the durability of resin-dentin bonded interfaces.^{4,6,7} Besides being an excellent antimicrobial agent, chlorhexidine is also an inhibitor of the activity of MMP-2, MMP-8 and MMP-9.³⁹ The use of chlorhexidine improved the integrity of the collagen network within hybrid layer after aging.⁴⁰⁻⁴² When chlorhexidine was applied as a pretreatment resulted in the maintenance of bond strength over 6 months³⁰ and 12 months, corroborating with our results.

As chlorhexidine, epigallocatechin-3-gallate has been used as dentin pretreatment associated with etch-and-rinse adhesive systems and it was effective in preserving bond strength up to 6 months.^{7,24} According to the results of the present study, epigallocatechin-3-gallate seems to be able to maintain the stability of resin-dentin bond for up to 12 months (Table 2). One reason for maintaining the bond strength over time by epigallocatechin-3-gallate may be due of its ability to enhance of collagen biomechanical properties.^{23,43} Collagen stabilization depends directly of the specific chemical structures of the bioactive agent involved in the process.^{5,43} Thus, epigallocatechin-3-gallate molecule containing galloyl groups can stabilize dentin collagen through hydrogen bonding and hydrophobic interactions.^{21,22,43} The greater ability by epigallocatechin-3-gallate to form collagen cross-links increases the ultimate tensile strength and elastic modulus of demineralized dentin.⁴³

Another plausible reason to improve the durability of the resin-dentin bond strength, is the fact of epigallocatechin-3-gallate inhibits of collagen-degrading enzymes action, such as MMP^{43,44} and cysteine cathepsins^{43,44}. The mechanism of action of epigallocatechin-3-gallate is likely more complex, involving multi-functional activities, such as down regulation of endogenous proteases expression,⁴⁴ protease inactivation/silencing⁴³⁻⁴⁴ and preventing the free access of collagenase to active sites on the collagen chains^{23,43}. The limitation of this study is the lack of evidence of inhibition enzyme activity by EGCG within the clinical application parameters used. Although epigallocatechin-3-gallate present promising results as dentin

pretreatment, it is necessary simplify the protocol of application. Thus, further studies should incorporate this catechin directly into the self-etch adhesive systems and analyze its effect on physicochemical properties.

CONCLUSIONS

Epigallocatechin-3-gallate and clorexidine when used as bioactive agent before Adper Easy One application did not affect the immediate resin-dentin bond strength and preserved the resin-dentin bond strength up to twelve months.

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TABLES

Table 1 – Adhesive system and bonding procedure.

Product	Composition	Manufacturer (#Batch n°)	Application mode
Adper Easy One*	Adhesive–Bis-GMA, HEMA, Methacrylated phosphoric esters dimethacrylates, 1,6 hexanediol dimethacrylate, polyalquenoic acid copolymer, silica nanofiller (7 nm), initiators, water and ethanol	3M ESPE, St.Paul, MN, USA (#402261)	1-apply one coat of adhesive for 20 s; 2-air-drying for 5 s; 3-light-curing for 10 s.

Abreviations: Bis-GMA: bisphenol A diglycidyl methacrylate; HEMA: 2-hydroxyethyl methacrylate. *This brand name is the same product as Adper Easy Bond.

Table 2 – Bond strength values (MPa \pm SD (*)) according to pretreatment solution.

Groups	Adper Easy One		
	24 hours	6 months	12 months
Distilled water	23.5 \pm 4.3 (32) ^{A,a}	16.7 \pm 6.1 (30) ^{B,b}	13.8 \pm 4.5 (37) ^{B,b}
CHX	24.5 \pm 6.6 (37) ^{A,a}	26.5 \pm 5.9 (37) ^{A,a}	25.2 \pm 5.2 (43) ^{A,a}
EGCG	24.7 \pm 4.4 (32) ^{A,a}	25.4 \pm 5.5 (33) ^{A,a}	26.8 \pm 5.0 (40) ^{A,a}

Identical superscript letters indicate no statistical significance between values. Capital letters compare treatments and lower cases compare storage time.

(*) corresponds to the number of sticks tested per group in each period

Table 3 – Distribution of mode of fracture of each group expressed as *n* (relative percentage)

Groups	Adper Easy One														
	24 hours					6 months					12 months				
	A	M	CR	CD	PF	A	M	CR	CD	PF	A	M	CR	CD	PF
Distilled water	3 (8)	29 (76)	3 (8)	0 (0)	3 (8)	2 (5)	28 (74)	2 (5)	1 (3)	5 (13)	11 (27)	25 (63)	1 (3)	0 (0)	3 (7)
CHX 2%	12 (27)	25 (55)	5 (11)	0 (0)	3 (7)	10 (23)	27 (61)	4 (9)	0 (0)	3 (7)	10 (21)	32 (68)	4 (9)	0 (0)	1 (2)
EGCG 0.1%	2 (5)	30 (77)	6 (15)	0 (0)	1 (3)	6 (18)	27 (79)	0 (0)	0 (0)	1 (3)	8 (19)	31 (74)	2 (5)	0 (0)	1 (2)

A: adhesive failure; M: mixed failure; CR: cohesive failure in resin; CD: cohesive failure in dentin; PF: premature failure

Capítulo 3

3.3 CAPÍTULO 3

Physicochemical properties of a methacrylate-based dental adhesive incorporated with epigallocatechin-3-gallate

Running title: Properties of adhesive doped with catechin

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ABSTRACT

This study aimed to evaluate the influence of epigallocatechin-3-gallate (EGCG) incorporation on the physicochemical properties of a methacrylate-based dental adhesive. EGCG was added to Adper Easy One (3M-ESPE), except in control group, to obtain concentrations of 0.01% w/w and 0.1% w/w of EGCG-doped adhesives. For water sorption (WS) and solubility (SL) surveys, resin discs were assayed following ISO recommendations ($n=10$). The degree of conversion (DC) was analyzed by FTIR whereas flexural strength (FS) was tested in three-point bending with bar specimens ($n=10$). Data were subjected to one-way ANOVA and Tukey's test ($p<0.05$). No significant difference in the DC, WS and FS were found between the different concentrations of EGCG ($p>0.05$). Adhesives containing 0.1% or 0.01% of EGCC demonstrated similar values of SL ($p>0.05$); and lower than those found for adhesive without EGCC ($p<0.05$). In conclusion, the addition of EGCC to adhesive reduced the solubility without affect the other properties evaluated.

Keywords: Mechanical phenomena; Dental adhesives; Flavonoids.

INTRODUCTION

Dental adhesives have created several alternatives for direct build-ups and other restorative treatments since their introduction more than forty years ago (1). However, their bonding performance on dentin may not be as durable as previously assumed (2). The resin-dentin bonds attained by using most of the contemporary adhesives can deteriorate over time (3,4).

A recent review has indicated that the hybrid layer created by the *in situ* polymerization of resin monomers (generally dimethacrylates) infiltrated in a partially demineralized dentin organic matrix is the weak link in the adhesive restorations (5). Aging may affect most of the components in the resin-dentin interface individually or several simultaneously (6). The dentin bond degradation may occur via two mechanisms: (1) the hydrolytic degradation of the polymer within the hybrid and adhesive layers and (2) the breakdown of the resin-sparse collagen fibrils (1,2). By self-etching the dental substrates without a separate application of phosphoric acid, the zone of resin-sparse collagen is reduced (6).

Overall, the self-etch adhesives contain relatively more hydrophilic components since acidic functional monomers and water are needed to a feasible etching of dentin and enamel (7,8). Therefore, these more hydrophilic adhesives tend to rapidly absorb water (7). Water transudation accelerates the swelling, plasticization and the elution of unreacted monomers (9). Furthermore, the accelerated degradation of the collagen by host-derived enzymes (i.e. matrix-metalloproteinases and cathepsins) plays a significant role on the degradation of dentin bonds (10). Thus, both mechanisms could lead to degradation of the dentin bond (7,10) as well as the drop on the physical properties of dental adhesives (11) by the polymer hydrolysis.

Some natural chemicals, such as flavonoids (present in fruits, vegetables, nuts, seeds and flowers) have shown enzymatic inhibition (4) and potential collagen cross-linking (12). Epigallocatechin-3-gallate (EGCG) is the principal flavonoid of green tea (*Camelia sinesis*), and it has been investigated due to its potential against cancer (13). Notably, this polyphenol has been demonstrated to be an effective inhibitor of matrix-metalloproteinases (MMPs) and cysteine cathepsins (13). It has been shown that EGCG can increase the collagen crosslinking and prevent the free access of collagenase to the active sites on the collagen chains (14). Recent studies demonstrated the beneficial effects of epigallocatechin-3-gallate mixed in dental adhesive which maintained resin-dentin bond strength over time (3,4). Nevertheless, the effects of this mixture on the physicochemical properties of self-etch dental adhesives have never been previously evaluated.

Thus, the aim of this study was to evaluate the influence of epigallocatechin-3-gallate incorporation on the physicochemical properties of a commercial methacrylate-based dental bonding agent. The hypothesis to be tested was that different concentrations of EGCG do not cause a detrimental effect on the water sorption/solubility, the degree of conversion and the flexural strength of the adhesive.

MATERIAL and METHODS

1. Doping the adhesive with EGCG

A commercial one-step self-etch adhesive, Adper Easy One (EO, also known as Adper Easy Bond, 3M ESPE, St. Paul, MN, USA – batch #402261), was used in this study. Epigallocatechin-3-gallate (EGCG, Sigma Aldrich, St. Louis, MO, USA) was added to the adhesive system in 0.01% w/w (EGCG 0.01) or 0.1% w/w (EGCG 0.1) concentrations. In order to obtain a homogenous mixture, the EGCG-doped adhesives were shaken using a tube agitator (QL-901, Biomixer, São Paulo, SP, Brazil) in darkness for 1 min. The homogeneity of the dilution was carefully checked and it was used only if no crystals were noted. The three tested adhesive systems (EO - Control, EGCG 0.01 and EGCG 0.1) were evaluated *in vitro* using water sorption, water solubility, degree of conversion and flexural strength experiments.

2. Water sorption/solubility

Water sorption and solubility were determined following ISO 4049:2000 except for specimen dimensions as previously undertaken by Ito et al.(15).

The disc specimens ($n=10$) were prepared by using a Teflon matrix (10 mm diameter and 1 mm thickness). A Mylar strip and a glass slide were placed on the discs after dispensing the adhesive systems using micro-pipettes. The light-activation was undertaken for 40 s using a halogen lamp light source (VIP junior, Bisco Inc., Schaumburg, IL, USA) with 500 mW/cm² irradiance at both sides of each specimen and they were ground and slowly polished up to a thickness of 0.5 mm using 600-grit SiC polishing papers.

The discs were stored in a silica-containing desiccator at 37°C and were repeatedly weighed after a 24 h intervals on an analytical balance (AUX-220, Shimadzu, Tokyo, Japan) with an accuracy of 0.0001 g up to a constant mass (m_1) was obtained (i.e., variation less than 0.1 mg in three weight measures). The volume of each specimen were measured with a 0.001 mm precision digital caliper (Absolute Digimatic, Mitutoyo, Tokyo, Japan) by analyzing the diameter and thickness, and the volume (V) was expressed in mm³. Thereafter, the specimens were stored in sealed glass vials with 10 ml of distilled water at 37°C for 7 days. Afterwards, the specimens were weighed after gently wiped on absorbent papers to obtain a constant mass (m_2) and then they were returned to the desiccator. The specimens were finally weighed as aforementioned up to stabilization of a constant mass (m_3). Water sorption (WS) and solubility (SL) ($\mu\text{g}/\text{mm}^3$) were calculated using the following formulae:

$$WS = \frac{m2 - m3}{V} \qquad SL = \frac{m1 - m3}{V}$$

The data were statistically analyzed using one-way ANOVA and Tukey's test ($p < 0.05$).

3. Degree of conversion

The degree of conversion (DC) of the adhesive resins was assessed by Fourier Transform Infrared Spectroscopy (FTIR) (Perkin-Elmer Spectrum 100, Perkin Elmer, Shelton, USA). Each adhesive system was dispensed into a small agate mortar and thoroughly mixed with potassium bromide (KBr) using a pestle, at a ratio of 4:100 w/w. The pellets of KBr/adhesive solution were prepared with a hand press (Hand Press Kit 161-1100, PIKE Technologies, Madison, WI, USA). FTIR spectrum of the uncured adhesive was obtained from each sample using 32 scans in a range of 4000-400 cm^{-1} , at 4 cm^{-1} resolution in transmission mode.

The adhesive resins were light-activated for 20 s using the light source (VIP junior, Bisco Inc., Schaumburg, IL, USA). Additional FTIR spectra were obtained immediately after light curing. The analyses were performed at room temperature with 50% relative humidity. Ten specimens per group ($n=10$) were tested. The rate of unreacted carbon-carbon double bonds (C=C) was determined from the ratio of absorbance intensities of aliphatic C=C (peak at 1636 cm^{-1}) against an internal standard (aromatic carbon-carbon bond peak at 1608 cm^{-1}) before and after curing. Degree of conversion was determined by subtracting the C=C from 100%. Data were statistically analyzed using one-way ANOVA and Tukey's test ($p < 0.05$).

4. Flexural Strength

A three-point bending test was used to assess the flexural strength (FS) of bar-shaped specimens. Twenty microliters of each adhesive resin was placed into a Teflon matrix to prepare the bar-shaped specimens (25 mm length x 2 mm width x 2 mm height) according to ISO 4049:2000. The adhesive resins were following air-dried for 20 s for solvent evaporation. Before the light-activation, a Mylar strip was placed on the top of the matrix and covered with a glass slide.

The adhesives were light-activated for 40 s with the light source (VIP junior, Bisco Inc., Schaumburg, IL, USA) at 3 different positions (right, middle and left) of sample. After light activation, the specimens were removed from the matrix, and the bottom surface underwent additional light-activation for 40 s. Specimens were stored for 24 h in distilled water at 37°C and subjected to a three-point bending test using a universal testing machine (Instron 3345, Instron Corp., Canton, MA, USA) at a crosshead speed of 1.0 mm/min. The dimensions of each specimen were captured using a digital caliper (0.01 mm precision, Absolute Digimatic, Mitutoyo, Tokyo, Japan). The FS calculated and transformed to MPa according to the formula:

$$FS = \frac{3FI}{2wb^2}$$

F=maximum force (N) at the fracture; I=the distance between the supports (fixed at 20 mm), w=specimen width; and b=specimen thickness.

Data were statistically analyzed using one-way ANOVA and Tukey's test ($p < 0.05$).

RESULTS

The results of the study are presented in Table 1. There was no significant difference in the WS between all groups ($p=0.158$). On the other hand, the SL outcomes were significantly different ($p<0.001$) presenting reduced solubility by increasing the EGCG concentration.

The degree of conversion was significantly similar between all tested adhesive resins ($p=0.214$). There were no significant differences in the FS values between all groups ($p=0.313$).

DISCUSSION

The main active component of green tea is epigallocatechin-3-gallate (EGCG). It has been shown several outstanding therapeutic effects on oral health (16). Two principal properties of EGCG related to restorative/conservative dentistry are its bacteriostatic activity (17) on *Streptococcus mutans*, one of the main bacteria responsible for the progress of dental caries, and its MMP inhibition potential which strikingly prevents dentin collagen degradation (18). Furthermore, dentin treatment using EGCG significantly improved the mechanical properties of demineralized dentin which suggests potential collagen crosslinking (19). Previous investigations also demonstrated the action of EGCG in very low concentrations (3,17). Therefore, the use of very low concentrations of EGCG as used in the present study might be suitable to attain its potential therapeutic effects.

The water sorption of dental polymers may be positively correlated with the hydrophilicity of the overall components (7-9,15). Most dental methacrylate-based monomers as well as photoinitiators contain polar functionalities such as esters and hydroxyls (20). The polarity of adhesive systems influences their affinity with water molecules (8). Though epigallocatechin-3-gallate is a relatively polar molecule due to several hydroxyls (21,22), no increase in the water sorption was attained in EGCG-doped adhesives (Table 1). Pallan et al. showed that the water sorption of different resins was associated with their formulations (hydrophilic components/features) instead of the presence of epigallocatechin-3-gallate (23). Indeed, the low concentrations of EGCG (0.01% and 0.1%) in the present investigation were not able to cause a significant increase in the overall hydrophilicity of the one-step self-etch adhesive, which could result insignificantly increased water sorption.

Simplified adhesives (i.e. one-step self-etch adhesives) often exhibit relatively high water sorption and solubility (9,15). High solubility was also found when epigallocatechin-3-gallate (1 wt% and 2 wt%) was incorporated to adhesive resins (23). It was speculated that the increase in solubility of EGCG-doped resins occurs due to the release of EGCG along with residual monomers and oligomers. Contrariwise, in the present study, the incorporation of EGCG (0.01% and 0.1%) reduced the SL of the tested resin. Such outcome may be explained by the difficult achievement of a homogeneous mixture between the branched catechin (EGCG) molecule and the polymer network. Potential hydrogen bonds between the EGCG hydroxyls, HEMA and Bis-GMA (23) are also feasible linkages which could impair the releasing of components resulting in lower solubility.

The degree of conversion is a useful strategy to assess the basic physicochemical properties of a dental resin, such as maximum polymerization reaction (24). The present findings demonstrated that the incorporation of EGCG in low concentrations into a specific one-step self-etch adhesive did not cause any negative effect on the DC (Table 1). These results are corroborated by recent studies (3,23). Du et al. speculated that high EGCG concentrations (more than 0.03%) could induce the formation of linear polymer chains, and the free radical scavenging effect of EGCG may jeopardize the polymerization (3). Indeed, the low concentrations of EGCG may have avoided these phenomena without interfering with polymerization conversion.

The final conversion after light-curing affects several mechanical properties such as the flexural strength (24). Nevertheless, polymers with similar degree of conversion may exhibit different crosslink densities which may afford contrasting flexural strengths (25). Once EGCG incorporation did not affect the FS (Table 1), we may assume that the crosslinking of the tested resins is similar. In fact, the same crosslinking of the polymer matrix play an important role on the resemblance in the water sorption among all concentrations of EGCG.

The observed results showed that the incorporation of EGCG into Adper Easy One, a one-step self-etch adhesive, did not affect its physicochemical properties, except the solubility. This leads to a partial acceptance of the hypothesis. A limitation of this study was not quantifying the release of EGCG from the adhesive system. Therefore, further studies are encouraged to assess the releasing of EGCG in order to inhibit proteinases and improve the resin-dentin bonds over time. In conclusion, EGCG incorporation (0.01% and 0.1%) into a self-etch dental adhesive may be useful for therapeutic adhesion to dental hard substrates, particularly to dentin, due to the potential of improving the longevity of adhesive procedures.

RESUMO

O presente estudo teve como objetivo avaliar a influência da incorporação de epigallocatequina-3-galato (EGCG) nas propriedades físico-mecânicas de um sistema adesivo. O EGCG foi adicionado ao Adper Easy One (3M-ESPE), exceto para o grupo controle, para a obtenção das concentrações de 0,01% e 0,1% p/p. No ensaio de sorção (S) e solubilidade (SL), foram confeccionados discos de resina de acordo com as recomendações da ISO ($n=10$). O grau de conversão (GC) foi analisado através de FTIR, enquanto a resistência flexural (RF) foi avaliada em um teste de flexão em três pontos com espécimes em forma de barra ($n=10$). Os dados foram submetidos à Análise de Variância a um critério e teste de Tukey ($p<0.05$). Não houve diferença significativa entre as concentrações de EGCG testadas no GC, S e RF ($p>0,05$). Adesivos contendo EGCG a 0,1% ou 0,01% apresentaram valores similares de SL ($p>0,05$); e inferiores aos valores obtidos pelo adesivo não incorporado por EGCG. Conclui-se que a adição de EGCG ao adesivo reduziu a solubilidade sem afetar as outras propriedades avaliadas.

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TABLES

Table 1. Mean (95% confidence intervals) of physicochemical properties tested.

Groups (<i>n</i> =10)	Water Sorption ($\mu\text{g}/\text{mm}^3$)	Solubility (mg/mm^3)	Degree of conversion %	Flexural Strength (MPa)
EO (Control)	180.2 (175.1 -185.3)	82.4 (79.3 - 85.5) ^a	68.5 (66.5 - 70,1)	18.7 (15.6 - 21.8)
EGCG 0.01% w/w	173.9 (165.3 - 182.5)	59.7 (53.3 -66.1) ^b	69.4 (67.5 - 71.3)	16.0 (13.9 - 18.1)
EGCG 0.1% w/w	180.0 (173.8 -186.2)	62.9 (57.5 - 68.3) ^b	70.6 (68.6 - 72.6)	18.5 (14.8 - 22.2)

*Distinct superscript letters indicate statistical difference in the same columns ($p < 0.05$).

Capítulo 4

3.4 CAPÍTULO 4

Development and characterization of PLGA microparticles for controlled release of epigallocatechin-3-galate

Characterization of catechin-loaded polymeric microparticles

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ABSTRACT

The aim of the study was to develop and characterize of poly (lactide-co-glycolide) acid (PLGA) microparticles for controlled release of epigallocatechin-3-galate (EGCG), using two types of PLGA. A parallel stability study revealed that EGCG remained stable in solution over 50 days, independent of the concentration (5, 20 or 40 $\mu\text{g/mL}$) and storage condition (room temperature, incubator or freezer). PLGA 50:50/EGCG microparticles had spherical shape and smooth surfaces. Conversely, PLGA 75:25/EGCG microparticles did not presented spherical shape and had an irregular surface, some microparticles presented external voids. Microparticle size of PLGA 50:50/EGCG averaged 1.010 μm and PLGA 75:25/EGCG was 7.159 μm . The yield of production was 55.33 and 37.11% to PLGA50:50 /EGCG and PLGA75:25 /EGCG, respectively. PLGA 50:50/EGCG showed the encapsulation efficacy lower than PLGA 75:25/EGCG. At 24 h, PLGA 75:25/EGCG showed higher EGCG release (64.04%) than PLGA 50:50/EGCG (7.77%) ($p < 0.001$). On the other hand, there was not statistical difference between the EGCG cumulative release values for PLGA 50:50/EGCG (95.13%) and PLGA 75:25 (100%) at 2952 h ($p = 0.409$). Both microparticles showed a pulsatile drug release pattern, which would benefit the need of different peaks of action. It can be concluded that epigallocatechin-3-gallate loaded-PLGA microparticles could be a useful alternative for controlled release of this drug.

Keywords: catechin; polymers; drug delivery systems

1. INTRODUCTION

Green tea (*Camellia sinensis*) is one of the most popular beverages in the world, and its consumption is directly associated with beneficial effects on human health (Okello et al., 2015). Epigallocatechin-3-gallate is the principal component of green tea extract, accounting for about 48%-55% of the total catechins (Jun et al., 2010). This polyphenol has received considerable attention from the scientific community because it can interfere and control some pathological processes (Khurana et al., 2013).

Epigallocatechin-3-gallate is effective against cancer (Irimie et al., 2015), reduces the oxidative stress of cells and inflammation process (Liu et al., 2014), prevent cardiovascular diseases (Kim et al., 2010) and reduce the risk of neurodegenerative diseases (Mandel et al., 2004; Walker et al., 2015). In addition, It has been demonstrated that epigallocatechin-3-gallate prevents infectious diseases, such hepatitis B (Huang et al., 2014), caries (Xu et al., 2011) and periodontal disease (Asahi et al., 2014), by inhibition of some microorganisms activity (Reygaert, 2014). However, to produce therapeutic effects is necessary to guarantee the stability and bioavailability of epigallocatechin-3-gallate in the medium (Song et al., 2014).

The oral bioavailability of green tea catechins is low in humans, resulting in plasma concentrations 5 to 50 times less than concentrations shown to exert biological activities in *in loco* (Hong et al., 2001). When green tea is consumed, it is estimated that only 5% of the original dose of epigallocatechin-3-gallate is absorbed and go to systemic circulation (Zhu et al., 2000). Two factors considered to be contributing to the limited oral bioavailability are the sensitivity of catechin to the digestive system and absorption barriers found in the human gastrointestinal tract (Song et al., 2014). In an attempt to improve the bioavailability of catechins in human organism has been researched protection of epigallocatechin-3-gallate molecules with polymers (Srivastava et al., 2013) and the application of this catechin *in situ* (Kato et al., 2010; Shin et al., 2014), both yield promising results.

The Poly (lactide-co-glycolide acid) blends are classic polymer used to develop drug delivery systems due to their superior biodegradability, biocompatibility and be approved by US FDA and European Medicine Agency, including for the invasive routes. (Li et al., 2008) The release of the drug occurs through contact of the microparticles with body fluids (Cappelano et al., 2014). The liquids penetrates into the microparticles, dissolves the drug, and causes polymer chain cleavage. Once dissolved, the drug starts to diffuse out into the surrounding bulk fluid (Gaignaux et al., 2012). The time degradation of poly (lactide-co-glycolide acid) and pattern of drug release depend on the molecular weight of polymer

(lactide:glycolide rates) and hydrophobicity (Makadia and Siegel, 2011). To the best of the current authors' knowledge, no studies exist that have examined the pattern release of catechin performed with different types of poly (lactide-co-glycolide) acid.

Thus, the aim of the present study was to promote the development and characterization of PLGA microparticles for controlled release of epigallocatechin-3-gallate, using two types of PLGA. The null hypothesis tested was that there will be no statistical difference between the percentage of cumulative released of epigallocatechin-3-gallate after 24 and 2952 hours, regardless type of poly (lactide-co-glycolide acid).

2. MATERIALS AND METHODS

1. Materials

Poly (D-L lactide-co-glycolide) acid (PLGA): Resomer® RG502H (PLGA 50:50 - batch #STBD2887V) and Resomer® RG756S (PLGA 75:25 - batch #STBC6378V) were purchased from Sigma Aldrich, Germany. Epigallocatechin-3-gallate (EGCG - batch #SLBL1959V) and ethyl acetate (batch #DCBB6676) were purchased from Sigma Aldrich, United States of America. Dichloromethane (DCM - batch #65456) was obtained from Dinâmica, Brazil. All other chemicals were of analytical reagent grade and were used as received.

2. Analytical validation

2.1 Epigallocatechin-3-gallate quantification

Validation was performed to guarantee linearity, precision and accuracy in the measurements. Six concentrations, ranging from 2.5 to 30 µg/mL, were assayed using an UV-visible spectrophotometer (Evolution 60S, Thermo Scientific, Ohio, USA). The standard curve presented feasible within the assayed concentrations ($R^2 = 0.9997$).

2.2 EGCG stability in solution

A stability experiment was performed to evaluate the degradation kinetics in aqueous medium (Milli-Q water; pH 7.40) of this drug, after storage under different conditions. Concentrations of 5, 20 and 40 µg/mL of EGCG were measured after storage in room temperature (25 °C), incubator (37 °C) and freezer (4 °C). Solutions were measured at pre-set time intervals along 50 days to guarantee its practical usage.

3. Preparation of EGCG-load PLGA microparticules

Formulations were prepared in the ratio PLGA: EGCG of 16:1. Two different forms of PLGA was used: 50:50 and 75:25 as mentioned in the materials section. Due to differences in solubility among the drugs and polymer, an emulsification process was proposed. Briefly, PLGA 5.12% w/v was dissolved in DCM and EGCG 0.64% dissolved in ethyl acetate were mixed, under magnetic stirring for 10 min at 25°C using a high shear mixer (Ultraturrax IKA T10B; IKA/Works, Inc. NC, USA) at 19,000 rpm for 5 min.

Resulting PLGA/EGCG emulsion was immediately spray dried using a Büchi B-290 mini spray drier (Büchi Labortechnik AG, Flawil, Switzerland), according to Souza et al. (2010) All obtained EGCG-load PLGA microparticules were collected from the glass containers and stored at 4°C. A blank formulation (PLGA only) was obtained in the same manner and used as a reference.

4. Physical and chemical characterization of EGCG-loaded microparticles

4.1 Morphological and size determination

Microparticles were morphologically examined by scanning electronic microscopy (FE-SEM Ultra Plus; Carl Zeiss, Baden-Württemberg, Germany). Samples of dried microparticles were placed on double-sided carbon adhesive stickers and analyzed freshly in the apparatus.

Microparticles size and polydispersity index were determined by light scattering (Zetasizer Nano ZS, Malvern, Worcestershire, United Kingdom) in aqueous solution of Tween™ 20 (0.1%) (Sigma Aldrich, St. Louis, MO, USA). Measurements were made in triplicate.

4.2 Yield of production, Encapsulation efficacy and Drug loading

The yield of production of microparticles was determined according to the ratio between the obtained mass and theoretical mass of the polymer and drug used.

The amount of EGCG entrapped within the microparticles was determined using the solvent-separation method. For that, 10 mg of microparticles accurately weighted were added and dissolved in 0.4 mL of DCM, followed by the addition of 0.8 mL of Milli-Q water and stirred vigorously using a tube agitator (QL-901, Biomixer, São Paulo, SP, Brazil). The samples were centrifuged for 10 min at 10,000 rpm (NT810, Novatecnica, São Paulo, Brazil) to separate the two phases. The procedure was repeated until the complete extraction of the drug.

Encapsulation efficacy (EE) and Drug loading (DL) ($n=3$) were calculated using the following formulae:

$$EE (w/w \%) = \frac{M_{enc}}{M_0} \quad DL(w/w\%) = \frac{M_{enc}}{M_{ms}}$$

Where M_{enc} is the real content of drug in the microparticles, M_0 is the theoretical amount of drug in the formulation and M_{ms} is the mass (mg) of microparticles used in the assay.

5. *In vitro* EGCG release studies

Five milligrams of EGCG-loaded microparticles ($n=4$) were placed in the glass vials in direct contact to the release medium (1.5 mL of Milli-Q water) and kept in an incubator (TE-392/1-MP; Tecnal Equipamentos para Laboratório, Piracicaba, SP, Brazil) at 37 °C. The release medium was collected at pre-setted times and measured immediately by a UV-Vis Spectrophotometer (DU-730; Beckman Coulter, Fullerton, CA, USA). Release medium was changed when release the concentration overcame the limit of standard curve values. Solutions were measured along over 4 months.

Statistical procedures were performed with the statistical software Sigmastat 3.5 (Systat Software Inc., San Jose, CA, USA) for Windows statistical program software. A Shapiro-Wilk test was applied to all groups to analyze the normal distribution of errors and the Barlett test for the homoscedasticity. To analyze EGCG cumulative release in 24 h and 2952 h was used Two-way ANOVA on Ranks (independent factors: PLGA blends and storage time). Post-hoc comparisons was analyzed by Holm-Sidak method. The level of significance was set at $p<0.05$.

3. RESULTS AND DISCUSSION

3.1 Analytical validation

3.1.1 EGCG stability in solution

There is great controversy in the literature regarding the stability of catechins in an aqueous medium (Wang et al. 2008). It seems that storage temperature can affect the stability of tea catechins (Chen et al., 2001; Wang et al., 2008). Wang et al. (2000) showed that approximately 30% of total catechins was lost when green tea was kept at 40 °C for 6 months. Other studies (Wang et al., 2008; Cheng et al., 2001) observed that tea catechins degraded slowly at temperatures between 25 and 100°C. Nevertheless, in the current study, EGCG levels remained stable over 50 days, independent of concentration (5, 20 or 40 µg/mL) and storage condition (room temperature, incubator or freezer) (Fig. 1). The degradation of epigallocatechin-gallate is commonly observed at high temperatures, above 70 °C (Wang et al., 2008).

3.2 Physical and chemical characterization of EGCG-loaded microparticles

3.3 Morphological and size determination

Particle size distribution and particle shape, together with some selected chemical properties, usually constitute the critical variables of a pharmaceutical manufacturing process (Shekunov et al, 2007). Spray dried particles are, normally, spherical, and their size can be described by their geometric diameter (Shekunov et al., 2007). In the current study, the micrographs showed that PLGA 50:50/EGCG microparticles had spherical shape and smooth surfaces (Fig. 2). However, PLGA 75:25/EGCG microparticles did not present spherical shape and had an irregular surface and, some microparticles presented external voids (Fig. 3). According to Vehring et al. (2008), external voids space can be created by process conditions and formulations that cause early separation of a soft surface layer, which folds to form a wrinkled morphology. The increased hydrophobicity of the polymer may have been crucial on the microparticles formation and caused a certain resistance on the polymer coating of the drug, which is majorly hydrophilic.

Particle size can be measured because influence the dissolution rate, controlled release and bioavailability of active pharmaceutical ingredients (Burgess et al., 2004). Microparticle size of PLGA 50:50/EGCG averaged 0.780 µm and PLGA 75:25/EGCG was 7.159 µm (Table 1). As necessary as the average particle size is the analysis of size distribution of particles in a

product (Vehring, 2008). PLGA 75:25/EGCG presented high polydispersity index (0.980) (Table 1), indicating that microparticles were not uniform and homogeneous, as represented in Figure 3. On the other hand, PLGA 50:50/EGCG showed homogeneous size of microparticles (Fig. 2), thus obtained a low PDI (0.219) (Table 1).

3.2.2. Yield of production, Encapsulation efficacy and Drug loading

The yield of production was 55.33 and 37.11% to PLGA50:50 /EGCG and PLGA75:25 /EGCG, respectively (Table 1). One limitation of spray dryer technique is the difficulty of recovering the atomized content (Zgoulli et al, 1999; Ambike et al., 2005). There may be a significant loss of the product during spray-drying process, due to adhesion of the microparticles to the inside wall of the spray-drier apparatus, and can also produce agglomeration of the microparticles (Takada et al., 1995). Low yield production observed in the PLGA75:25 /EGCG may be due to size and aggregation of particles (Fig. 3) that facilitated its sedimentation within the spray dryer chamber, making it difficult to collect further in the cyclone.

The encapsulation efficacy of the microparticles depends on different factors like concentration of the polymer, interaction between drug and polymer and solubility of polymer in solvent (Jyothi et al., 2010). PLGA 50:50/EGCG showed the encapsulation efficacy lower than PLGA 75:25/EGCG (Table 1). The increase in glycolic acid content in the poly (D-L lactide-co-glycolide) acid resulted in reduced solubility of the polymer in dichloromethane (Morishita and Park, 2010), and probably reduced the encapsulation efficacy of the PLGA 50:50/EGCG compared with PLGA 75:25/EGCG. The size may also have impacted the entrapment level observed within the different blends.

On the other hand, particles with maximal drug loading reduces the quantity of carrier required for the administration of sufficient amount of active compound (drug) to the target site as well as drug wastage during manufacturing (Govender et al., 1999). Both particles, PLGA 50:50/EGCG and PLGA 75:25/EGCG showed low levels of drug loading (Table 1).

3.4 *In vitro* EGCG release studies

There is a statistically significant interaction between microparticles types and release times ($p < 0.001$; $F = 17.611$). At 24 h, PLGA 75:25/EGCG showed higher EGCG release (64.04%) than PLGA 50:50/EGCG (7.77%) ($p < 0.001$). On the other hand, there was not

statistical difference between the EGCG cumulative release values for PLGA 50:50/EGCG (95.13%) and PLGA 75:25 (100%) at 2952 h ($p=0.409$).

PLGA 50:50/EGCG presented three-step burst release effect, at 96 h (21.15%), 360 h (61.56%) and 2952 h (95.13%), while the PLGA 75:25/EGCG showed a different pattern release with two-step burst release at 48 h (70.91%) and 504 h (100%) (Fig. 4A). Although, both microparticles showed a pulsatile drug release, PLGA 50:50/EGCG presented more controlled release pattern over the experimental period as evidenced by drug amount release at each interval. Additionally, PLGA 50:50/EGCG showed a delayed release after 2256 h (Fig. 4B).

Recently, pulsatile drug delivery systems are gaining a lot of interest and attention (Mangubat et al., 2015; Sha et al., 2015). A major objective of controlled delivery systems in the treatment of several diseases is to deliver the drug in higher concentrations during the time of greatest need according to the onset of the disease or syndrome (Pragna et al., 2013). Some researches have used the epigallocatechin-3-gallate associated with polymers, such drug delivery system, to treatment of atherosclerosis in rabbits (Hong et al., 2014), to prevent postsurgical adhesion (Shin et al., 2014), caries (Mankovskaia et al., 2013) and pancreatic cancer (Sun et al., 2014). In this context, PLGA 50:50/EGCG and PLGA 75:25/EGCG microparticles could be an alternative for use in Medicine and Dentistry.

4. CONCLUSION

Based on all experimental results, it can be concluded that epigallocatechin-3-gallate-loaded poly (D-L lactide-co-glycolide) acid microparticles, especially the ones based in PLGA 50:50, could be a useful tool for controlled drug release.

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TABLE

Table 1. Main characteristics of EGCG loaded-PLGA microparticles.

Microparticles	Yield of production (%)	Particle size (μm) (Pdl)	Encapsulation efficacy (%)	Drug loading (%)
PLGA 50:50	44.91	1.010 (0.219)	-	-
PLGA50:50 /EGCG	55.38	0.780 (0.687)	42.35	2.49
PLGA 75:25	48.31	5.832 (0.538)	-	-
PLGA 75:25 / EGCG	37.11	7.159 (0.980)	100	6.03

FIGURES

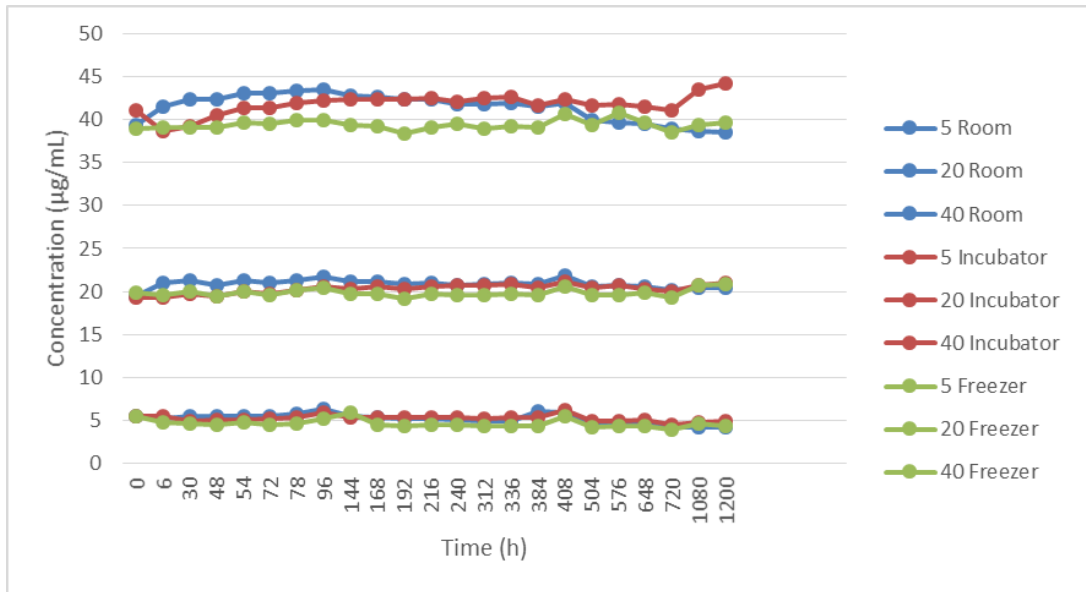


Fig. 1. Stability in solution of EGCG; where 5, 20 and 40 refer to the concentration in $\mu\text{g/mL}$, after storage under different conditions: room (25°C), incubator (37°C) and freezer (4°C) up to 50 days.

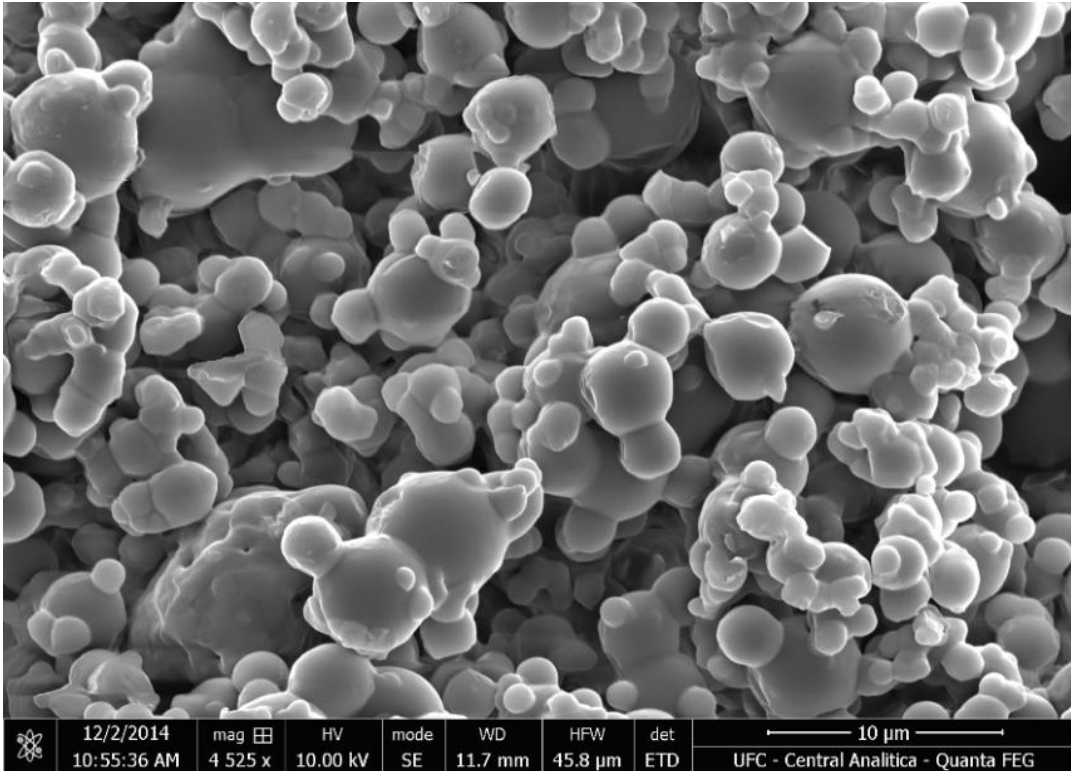


Fig. 2. SEM image of PLGA 50:50/EGCG microparticles

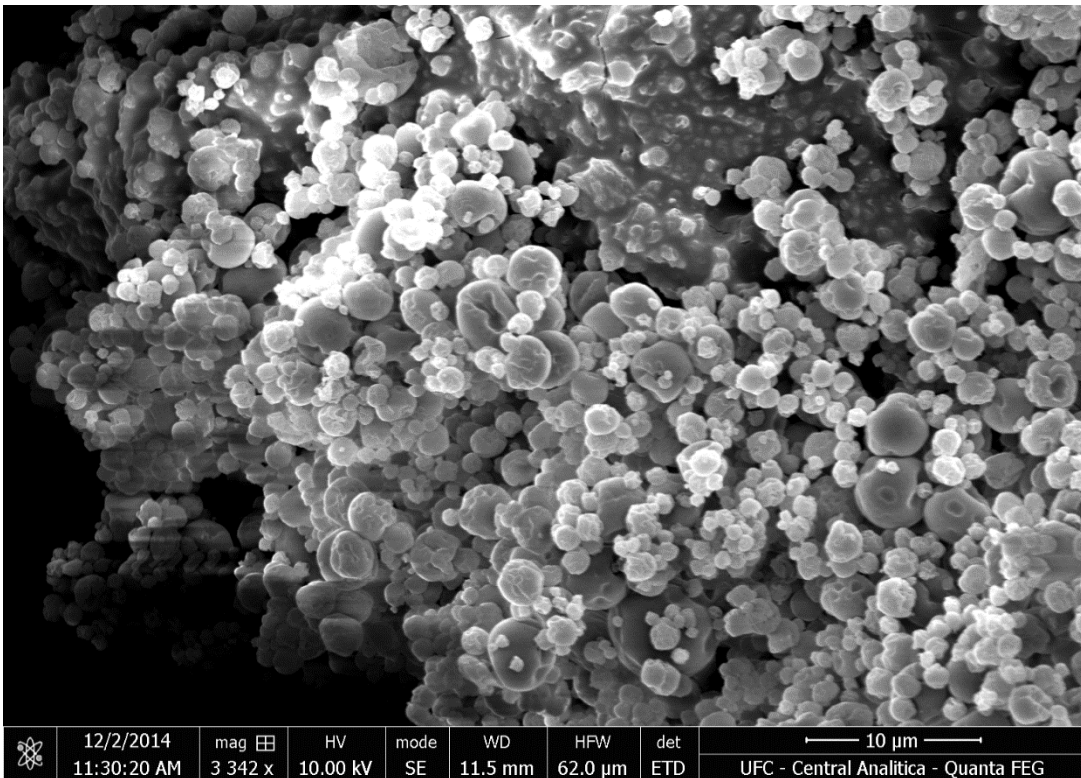
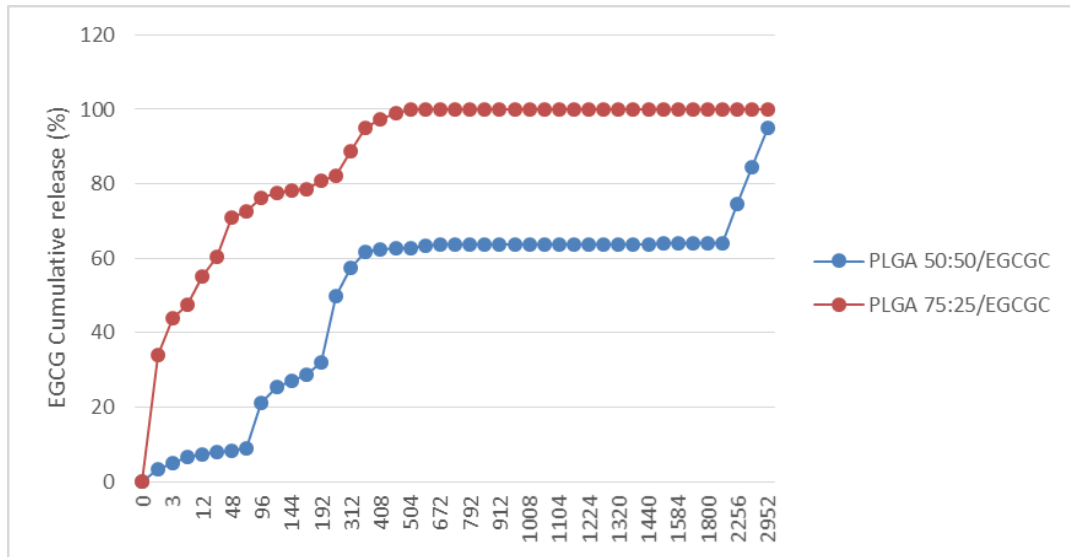


Fig. 3. SEM image of PLGA 75:25/EGCG microparticles

(A)



(B)

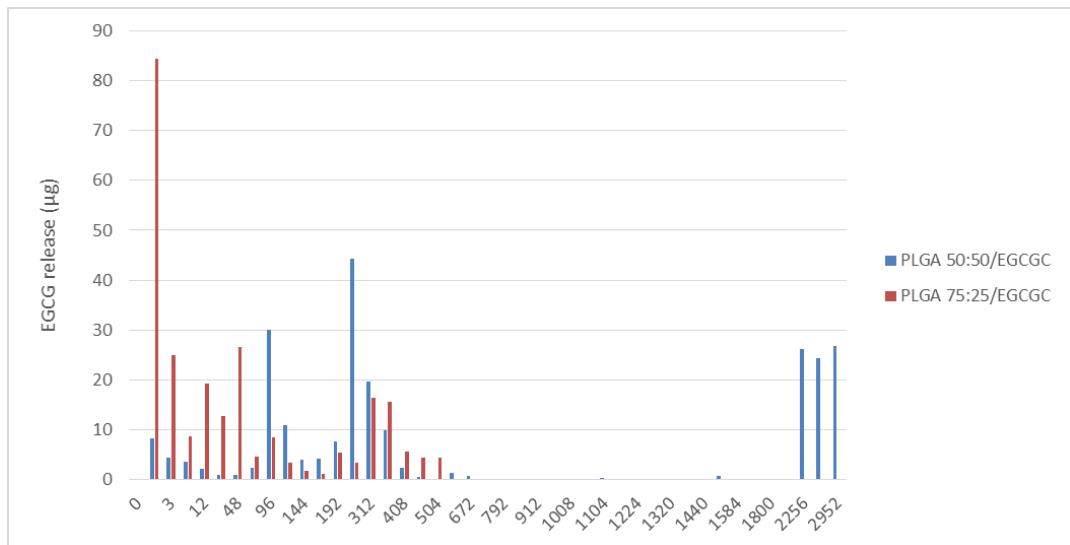


Fig. 4. *In vitro* EGCG release studies: EGCG profiles of PLGA microparticles (A); EGCG release over time from PLGA microparticles (B)

Capitulo 5

3.5 CAPÍTULO 5

Physicochemical properties and pattern of drug release of etch-and-rinse adhesive system incorporated with catechin-loaded polymeric microparticles

Running title: Dental adhesive doped with EGCG-loaded microparticles

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ABSTRACT

Objective: This study aimed to evaluate the epigallocatechin-3-gallate-load poly (lactide-co-glycolide) acid (PLGA) microparticules incorporation on the physicochemical properties of a two-step etch-and-rinse adhesive system, and the release rate of epigallocatechin-3-gallate (EGCG).

Methods: EGCG was added to the Single Bond 2, directly in 0.01% w/w (EGCG 0.01%) and 0.1% w/w (EGCG 0.1%) concentrations or microencapsulated with PLGA 50:50 or PLGA 75:25 microparticules in 0.5, 1, and 2% w/w. In phase 1, was evaluated EGCG cumulative release (CR) using an UV-Vis Spectrophotometer. In phase 2, degree of conversion (DC) was analyzed by FTIR whereas flexural strength (FS) and elastic modulus (E) were tested in three-point bending with bar specimens ($n=10$). For water sorption (WS) and solubility (SL) surveys, resin discs were assayed following ISO recommendations ($n=10$). Specimens sticks-shaped were used to evaluate the resin-dentin bond strength (μ TBS). One-way ANOVA was used to DC, FS, E, WS, SL, μ TBS and for analyze EGCG release was used Two-way ANOVA on Ranks. Comparisons post hoc was analyzed by Holm-Sidak method. The level of significance was set at $p<0.05$.

Results: 1% PLGA50:50/EGCG achieved d the highest release in quantitative terms reaching the highest released mass (77.30 μ g). There was no significant difference in the DC, E, WS, SL and μ TBS values among all groups ($p>0.05$). FS values were significantly increased in the 1% PLGA50:50/EGCG ($p<0.05$).

Conclusions: Single Bond 2 doped with EGCG-load PLGA 50:50 microparticules at 1% could be an alternative to promote controlled release without causing detrimental effects of physicochemical properties.

Clinical Relevance: Adhesives systems doped with epigallocatechin-3-gallate, directly or in PLGA microparticules, do not cause detrimental effect of restorative procedures.

Keywords: Catechin; Dentin; Physical properties; Dental adhesive; Drug delivery systems

INTRODUCTION

Biomodification of demineralized collagen matrices with cross-linking agents has attracted the attention of researchers in recent years.¹⁻⁴ The use of bioactive agents during the bonding procedures improves the biomechanical properties of dentin by increasing the tensile strength and modulus of elasticity of the collagen fibrils.^{1,5} Moreover, most strategies of dentin biomodification reduces the proteolytic activity of enzymes, such as cysteine cathepsins (CTs)^{4,6} and matrix metalloproteinases (MMPs),^{4,7,8} that degrade the collagen fibrils of hybrid layer. Therefore, dentin biomodification agents may have an important role on the preservation of resin-dentin interfaces.²

A large variety of bioactive agents can be found in the plants.^{1,4,8} Of particular interest to the adhesive dentistry is the epigallocatechin-3-gallate, the principal flavonoid of green tea (*Camelia sinensis*).^{3,9-11} This catechin prevents episodes of dental demineralization and inhibits the progress of caries in dentin.¹² In addition, epigallocatechin-3-gallate is an effective inhibitor of activity of MMP-2 and MMP-9^{4,8} and CT-B.^{4,6} It has been shown that epigallocatechin-3-gallate can increase the collagen cross-linking and prevent the free access of collagenase to the active sites on the collagen chains.¹³

Recent studies demonstrated that the use of epigallocatechin-3-gallate at 0.01%⁹ and 0.1%³ were effective in preserving resin-dentin bond strength up to 6 months. However, there is concern about the ability of adhesive systems to release the catechin for more long periods.¹⁰ The release of epigallocatechin-3-gallate, in water, when incorporated directly into polymeric materials was significantly reduced after 24 hours.¹⁰ Thus, the performance of epigallocatechin-3-gallate as an inhibitor of the enzyme activity could be insufficient to inhibit the enzymatic activity and maintain the bond strength after a few years.

The controlled release may be obtained by entrapping the epigallocatechin-3-gallate in biodegradable microparticles of poly (lactide-co-glycolide) acid.¹⁴ Poly (lactide-co-glycolide) acid is the most used polymer to develop drug delivery systems due to be biodegradable, biocompatible and approved by US FDA.¹⁵ The pattern of drug release may be affected by the composition of poly (lactide-co-glycolide) acid, since the poly lactic acid is more hydrophobic than poly glycolic acid.¹⁶ The release of the drug occurs through contact of the microparticles with aqueous body.¹⁷ Water molecules penetrates into the microparticles, dissolves the drug, and causes polymer chain cleavage. Once dissolved, the drug starts to diffuse out into the surrounding bulk fluid.¹⁸ Although the use of drug delivery systems seem promising, little is known about the consequences of their incorporation in the properties of adhesive systems.

Therefore, this study was designed to evaluate the effect of the epigallocatechin-3-gallate-load poly (lactide-co-glycolide) acid (PLGA) microparticules incorporation on the physicochemical properties of a two-step etch-and-rinse adhesive system. To achieve this objective, the study was divided into two phases. The aim of the phase 1 was to select the more effective EGCG pattern release in different types of poly (lactide-co-glycolide) acid microparticules incorporated in the two-step etch-and-rinse adhesive systems. On the other hand, the aim of phase 2 was to evaluate the physicochemical properties of a two-step etch-and-rinse adhesive system incorporated with epigallocatechin-3-gallate, directly or in poly (lactide-co-glycolide) acid microparticules, selected in the first phase. The first null hypothesis was that there is no significant difference between the groups regarding the EGCG release of adhesives systems, after 24 and 4320 hours. The second null hypothesis to be tested was that different incorporation modes of EGCG have no effect on the physicochemical properties.

MATERIAL and METHODS

1. Materials

Poly (D-L lactide-co-glycolide) acid (PLGA): Resomer® RG502H (PLGA 50:50 - batch #STBD2887V) and Resomer® RG756S (PLGA 75:25 - batch #STBC6378V) were purchased from Sigma Aldrich, Germany. Epigallocatechin-3-gallate (EGCG - batch #SLBL1959V) and ethyl acetate (batch #DCBB6676) were purchased from Sigma Aldrich, United States of America. Dichloromethane (DCM - batch #65456) was obtained from Dinâmica, Brazil. All other chemicals were of analytical reagent grade and were used as received.

2. Preparation of EGCG-load PLGA microparticules

Formulations were prepared in the ratio PLGA: EGCG of 16:1. Two different forms of PLGA were used: 50:50 and 75:25 mentioned in materials. Due to differences in solubility among the drugs and polymer, an emulsification process was proposed. Briefly, PLGA 5.12% w/v was dissolved in DCM and EGCG 0.64% dissolved in ethyl acetate were mixed, under magnetic stirring for 10 min at 25°C using a high shear mixer (Ultraturrax IKA T10B; IKA/Works, Inc. NC, USA) at 19,000 rpm for 5 min.

Resulting PLGA/EGCG solution was immediately spray dried using a Büchi B-290 mini spray drier (Büchi Labortechnik AG, Flawil, Switzerland), according to Souza *et al.*¹⁹. All obtained EGCG-load PLGA microparticules was collected in glass containers and stored in desiccators at 4 °C.

A blank formulation (PLGA only) was obtained and used as a reference.

3. Doping the adhesive with EGCG

A commercial two steps etch-and-rinse adhesive system, Adper Single Bond 2 (SB, 3M ESPE, St. Paul, MN, USA), was used in this study (Table 1). EGCG was added to the adhesive system in different incorporation modes, directly or microencapsulated, according to Figure 1. In

order to obtain a homogenous mixture, the EGCG-doped adhesives were shaken using a tube agitator (QL-901, Biomixer, São Paulo, SP, Brazil) in darkness for 1 min. The homogeneity of the dilution was carefully checked and it was used only if no crystals were noted.

4. Release assay of adhesives containing EGCG (Phase 1)

In view of the difference in hydrophobicity and biodegradation rates of the PLGAs (PLGA 50:50 and PLGA 75:25) used in this study, there was a release assay in order to observe their performance into the adhesive system (Adper Single Bond 2).

A series of reference solutions ranging from 2.5 to 40 µg/mL of EGCG in distilled water were prepared to obtain a linear relationship between absorbance peak height and drug concentration. A UV-Vis Spectrophotometer (DU-730; Beckman Coulter, Fullerton, CA, USA) was used to evaluate and confirm the absorbance peak of EGCG at 275 nm. The standard curve performance was within acceptable range for bioanalytical method acceptance ($R^2 = 0.99982$).

Nine disc-shaped specimens (6.0 mm diameter and 1.0 mm thickness) of each adhesive system were prepared by using a silicon matrix. A Mylar strip and a glass slide were placed on the discs after dispensing the adhesive systems using micropipettes. The light-activation was undertaken for 40 s using a light source (Ellipar Freelight 2, 3M ESPE, St. Paul, MN, USA) with 600 mW/cm² irradiance at both sides of each specimen and they were ground and slowly polished up to a thickness of 0.5 mm using 600-grit SiC polishing papers.

Discs were divided in three glass containing 1 ml of distilled water at 37 °C and stored until 180 days (4320 hours). The absorbance peaks heights of storage solutions were analyzed with a UV-Vis Spectrophotometer and converted to drug release rates based on the established linear calibration. The absorbance of blank microparticles was used to eliminate the polymer influence.

The adhesive system incorporated with PLGA microparticles loaded with EGCG that showed a pulsatile profile and highest mass released was selected for phase 2 of this study.

5. Analysis of the physicochemical properties of adhesive systems (Phase 2)

Four adhesive systems (SB - Control, EGCG 0.01%, EGCG 0.1% and 1% PLGA/EGCG) were evaluated *in vitro* using water sorption, water solubility, degree of conversion, flexural strength, elastic modulus, and microtensile bond strength experiments.

5.1. Degree of conversion

The degree of conversion (DC) of the adhesive resins was assessed by Fourier Transform Infrared Spectroscopy (FTIR) (Perkin-Elmer Spectrum 100, Perkin Elmer, Shelton, CT, USA). Each adhesive system was dispensed into a small agate mortar and thoroughly mixed with potassium bromide (KBr) using a pestle, at a ratio of 4:100 w/w. The pellets of KBr/adhesive solution were prepared with a hand press (Hand Press Kit 161-1100, PIKE Technologies, Madison, WI, USA). FTIR spectrum of the uncured adhesive was obtained from each sample using 32 scans in a range of 4000-400 cm^{-1} , at 4 cm^{-1} resolution in transmission mode.

The adhesive resins were light-activated for 20 s using the light source (Ellipar Freelight 2, 3M ESPE, St. Paul, MN, USA). Additional FTIR spectra were obtained immediately after light curing. The analyses were performed at 25 °C with 70% relative humidity. Ten specimens per group ($n=10$) were tested. The rate of unreacted carbon-carbon double bonds (C=C) was determined from the ratio of absorbance intensities of aliphatic C=C (peak at 1636 cm^{-1}) against an internal standard (aromatic carbon-carbon bond peak at 1608 cm^{-1}) before and after curing. Degree of conversion was determined by subtracting the C=C from 100%.

5.2. Flexural Strength and Elastic Modulus

A three-point bending test was used to assess the flexural strength (FS) and elastic modulus (E) of bar-shaped specimens following ISO 4049:2000 except for specimen dimensions that were adapted for the microflexural test as previously undertaken by Gaglione *et al.*²⁰

Each adhesive system was placed into a silicon matrix to prepare the bar-shaped specimens (7.0 mm length x 2.0 mm width x 1.0 mm height). The adhesive systems were following air-dried for 40 s for solvent evaporation and light-activated for 40 s with the light source (Ellipar Freelight 2, 3M ESPE, St. Paul, MN, USA).

After light activation, the specimens were removed from the matrix, and the bottom surface underwent additional light-activation for 40 s. Specimens were stored for 24 h in distilled water at 37 °C and subjected to a three-point bending test using a universal testing machine (Instron 3345, Instron Corp., Canton, MA, USA) at a crosshead speed of 1.0 mm/min. Prior the test, the dimensions of each specimen were captured using a digital caliper (Absolute Digimatic, Mitutoyo, Tokyo, Japan) to the nearest 0.01 mm and recorded with Bluehill 2 software (Instron Corp., Canton, MA, USA), which calculated FS (MPa) and E (GPa) values, according dimensions and tension.

5.3. Water sorption/solubility

Water sorption and solubility were determined following ISO 4049:2000 except for specimen dimensions as previously undertaken by Collares *et al.*²¹

Ten disc specimens (6.0 mm diameter and 1.0 mm thickness) were prepared as previously described in EGCG release test. The discs were stored in a silica-containing desiccator at 37°C and were repeatedly weighed after a 24 h intervals on an analytical balance (AUX-220, Shimadzu, Tokyo, Japan) with an accuracy of 0.0001 g up to a constant mass (m_1) was obtained (i.e., variation less than 0.1 mg in three weight measures). The volume of each specimen were measured with a 0.001 mm precision digital caliper (Absolute Digimatic, Mitutoyo, Tokyo, Japan) by analyzing the diameter and thickness, and the volume (V) was expressed in mm^3 . Thereafter, the specimens were stored in sealed glass vials with 1.5 ml of distilled water at 37°C for 7 days. Afterwards, the specimens were weighed after gently wiped on absorbent papers to obtain a constant mass (m_2) and then they were returned to the desiccator. The specimens were finally weighed as aforementioned up to stabilization of a constant mass (m_3). Water sorption (WS) and solubility (SL) ($\mu\text{g}/\text{mm}^3$) were calculated using the following formulae:

$$WS = \frac{m_2 - m_3}{V} \quad SL = \frac{m_1 - m_3}{V}$$

6. Microtensile Bond Strength Test (μTBS)

Thirty six unerupted, caries-free third molars were collected after the patients' informed consent had been obtained under a protocol reviewed and approved by the local Research and Ethics Committee (# 459.659). The selected teeth were stored in 0.01% thymol solution and used within one month after extraction. Occlusal enamel was removed using a diamond saw in a Labcut 1010 (Extec, Enfield, CT, USA) under water-cooling to expose a flat coronal dentin surface. The dentin surface was then prepared with a wet #600-grit SiC paper for 60 s to standardize the bonding surface.

The teeth were randomly allocated by the Excel software (Excel 2013, Microsoft Corporatin, One Microsoft Way, Redmond, WA, USA) into 4 groups (n=9) according to the adhesive system used. The exposed dentin surfaces of all teeth were etched with 35% phosphoric acid gel (Scotchbond Phosphoric Acid Etchant; 3M ESPE, St. Paul, MN, USA - batch #1219600378) for 15 s, rinsed for 30 s with distilled water, and dried with oil-/water-free air for 30 s. The teeth were re-hydrated with 20 μ L of distilled water and excess solution was removed with absorbent paper, leaving the dentin surface visibly moist. The etch-and-rinse adhesive system Adper Single Bond 2 (3M ESPE, St. Paul, MN, USA) was applied according to manufacturer's instructions (Table 1). After light curing the adhesive (Ellipar Freelight 2, 3M ESPE, St. Paul, MN, USA), five 1 mm thick increments of composite resin were built up (Filtek Z250XT; 3M ESPE, St. Paul, MN, USA - batch #37277). Each increment was light-cured (Ellipar Freelight 2) for 20 s, with a power density of 600 mW/cm². The bonded teeth were stored in distilled water at 37°C for 24 h.

After storage, the bonded teeth were longitudinally sectioned in both “x” and “y” directions across the bonded interface using a diamond saw in a Labcut 1010 (Extec, Enfield, CT, USA) under water cooling to obtain bonded sticks with cross-sectional area of approximately 1.0 mm². The cross-sectional area of each stick was measured with a digital caliper (Absolute Digimatic, Mitutoyo, Tokyo, Japan) to the nearest 0.01 mm and recorded for subsequent calculation of bond strength values.

Sticks from each tooth were randomly allocated by the Excel software (Excel 2013, Microsoft Corporatin, One Microsoft Way, Redmond, WA, USA) and assigned to three storage times: 24 hours, 6 months and 12 months. For microtensile test, 24 hours bonded sticks were attached with cyanoacrylate glue (Super Bonder Flex Gel, Loctite, São Paulo, SP, Brazil) to a modified Geraldeli testing apparatus (Odeme Biotechnology, Joaçaba, SC, Brazil) and subjected to a tensile force at 0.5 mm/min in a universal testing machine (Emic, São José dos Pinhais, PR, Brazil). The load at fracture was used to calculate bond strength (MPa). Specimens of 6 and 12 months remain stored for later testing.

The failure modes were evaluated at 80 X magnification (StereoZoom® Leica S8 APO, Leica Microsystems, Wetzlar, Germany) and classified as cohesive when they were located exclusively within dentin (CD) or resin composite (CR); adhesive (A) when failure occurred at the dentin/adhesive interface; or mixed (M) when two modes of failure occurred simultaneously.

7. Statistical analysis

Statistical procedures were performed with the Sigmapstat 3.5 (Systat Software Inc., San Jose, CA, USA) for Windows statistical program software. A Shapiro-Wilk test was applied to all groups to analyze the normal distribution of errors and the Barlett test for the homoscedasticity. For analyze cumulative EGCG release in 24 h and 4320 h was used Two-way ANOVA on Ranks (independent factors: storage time and adhesive system). One-way ANOVA was used to compare, GC, FS, E, WS, SL and μ TBS within each adhesive system. Comparisons *post hoc* was analyzed by Holm-Sidak method. The level of significance was set at $p < 0.05$. Teeth were used as a statistical unit and the number of prematurely debonded specimens was recorded, although this was not included in the analysis.

RESULTS

EGCG release profiles are shown in Figures 2 and 3. In phase 1, mean of EGCG cumulative release (%) were significantly influenced by adhesive system ($p < 0.001$; $F = 9.399$) and storage time ($p < 0.001$; $F = 140.924$). The interactions between variables were significant ($p < 0.001$; $F = 5.909$). At 24 h, there was no significant difference between EGCG release values among groups ($p > 0.05$). After 4320 h, there was no significant difference among 0.5% PLGA 50:50/EGCG, 1% PLGA50:50/EGCG and EGCG 0.01% ($p > 0.05$), and these adhesive systems reached total release (100%) during the assayed period. On the other hand, among the PLGA75:25 groups, the highest release obtained did not reach more than 68% of the overall drug content (0.5% PLGA75:25/EGCG).

All groups incorporated with PLGA microparticles showed a pulsatile release profile, characterized by moments of cessation and subsequent rapid increase of release. The 1% PLGA50:50/EGCG group presented four-step burst release effect, at 288 h (42.62%), 528 h (63.00%) 768 h (79.67%) and 1200 h (100%) (Figure 2). Additionally, 1% PLGA50:50/EGCG achieved the highest release in quantitative terms reaching the highest released mass (77.30 μg), among all studied groups and completing the full release at the end of 50 days (Figure 3). In contrast, the 0.5% PLGA 50:50/EGCG group showed the lowest release (39.84 μg) (Figure 3).

Table 2 showed DC, FS, E, WS and SL means values and standard deviation for adhesive systems. In the phase 2, there was no significant difference in the DC, E, WS and SL values among all groups ($p > 0.05$). However, FS results were influenced by treatment ($p = 0.009$; $F = 4.432$). The incorporation of 1% PLGA 50:50/EGCG significantly increased the flexural strength ($p < 0.05$). While the FS mean values for other groups were not statistically different ($p > 0.05$).

Mean μTBS values are displayed in Table 3. After 24 h of storage, there was not statistically significant difference between the mean bond strength values of all groups ($p > 0.05$). At 24 h, most of failures were mixed in all of the tested groups and, and cohesive failures (resin and dentin) were more prevalent than adhesive failures for specimens of all groups, except to SB group (control) (Table 4).

DISCUSSION

The idea of using adhesives systems as vehicles for the delivery of therapeutic agents is attractive²². Many bioactive agents have incorporated into the adhesive systems in an attempt to improve the durability of resin–dentin adhesive interfaces *in vitro*^{3,9,23,24} and *in vivo*.^{25,26} However, the interactions between resins monomers and drug particles can affect physicochemical properties of adhesive systems, such as monomer flexibility, degree of conversion, crosslinking density, hydrophilicity, and drug release.¹⁰

The adhesive systems should release enzymatic inhibitors in minimum concentrations to inactive MMPs and CTs. Epigallocatechin-3-gallate inhibited MMP-2 and MMP-9 at concentrations of 6 μM ($\approx 20 \mu\text{g/ml}$) and 0.8 μM ($\approx 3 \mu\text{g/ml}$), respectively^{7,8}, while CT-B were inhibited at 6,500 $\mu\text{g/ml}$.⁴ In the present study, after 4320 h of epigallocatechin-3-gallate release in 1 mL of distilled water, all groups demonstrated rates cumulative release that could inhibit MMPs, but not CTs. However, the amount of water within the hybrid layer created by simplified adhesives is low, once that fluid flow rate is approximately 4 $\mu\text{L cm}^{-2} \text{min}^{-1}$.²⁷ Thus, it is possible that the concentration of EGCG released within the interface is higher than that presented in the current study, indicating a limitation of release assay.

When adhesive system was loaded directly with EGCG at 0.01% and 0.1% presented a low and constant EGCG release. Pallan *et al.*¹⁰ observed similar release pattern using experimental resins incorporated with epigallocatechin-3-gallate at 0.2%, 1% and 2%. The low release of epigallocatechin-3-gallate may be of concern because MMPs with residual activity could continue degrading the collagen fibrils of the hybrid layer. However, Du *et al.*⁹ demonstrated that direct incorporation of catechin at 0.01% to adhesive systems were able to maintain the bond strength up to 6 months, showing that maybe the low epigallocatechin-3-gallate release does not affect the bond strength.

In contrast, the adhesive systems incorporated with PLGA microparticles presented a pulsatile-release profiles. Poly (lactide-co-glycolide acid) is known in medical science as the best biomaterial available for controlled drug delivery in relation to design and performance.^{16,28,29} Pulsatile release allows that higher doses are reached in consecutive moments,³⁰ which probably can be important to inactivate dentin endopeptidases that degrade the hybrid layer. After 4320 h of EGCG release, the majority of PLGA 75:25 groups presented cumulative release (%) significant lower than PLGA 50:50 groups ($p < 0.05$). Therefore, the first null hypothesis was rejected.

Mechanical properties of the resin-based bonding systems are strongly depend upon the DC.³¹ Epasinghe *et al.*²⁴ speculated that epigallocatechin-3-gallate may disturb the free radical polymerization of adhesive systems due to their free radical scavenging effect. Nevertheless, some studies demonstrated that the incorporation of epigallocatechin-3-gallate in low concentrations into adhesive systems did not cause any negative effect on the DC.⁹⁻¹¹ These facts corroborate with present study results. Probably, the catechin seems to have been entrapped within the linear chains after curing without interfering with monomers conversion.¹¹ In addition, PLGA microparticules did not interfere on the DC of two-step etch-and-rinse adhesive system. It is possible that DC was not affected by the addition of EGCG-loaded microparticles due to the existence of chemical compatibility between the resin monomers of adhesive system and the poly (lactide-co-glycolide) acid, once that both have polymeric nature.¹⁶

Epigallocatechin-3-gallate at 0.01% and 0.1% incorporated directly into adhesive systems did not affect the FS and E, corroborate with Neri *et al.*¹¹ On the other hand, when etch-and-rinse adhesive system was incorporated with EGCG loaded PLGA microparticules there was an increase in FS. Probably the undissolved particles may have acted as reinforcement leading to the higher values in FS.³² Therefore, the second null hypothesis was rejected.

Polymers are considered insoluble structures that exhibit relatively high chemical stability.³³ Nevertheless, when exposed to oral environment the polymeric materials tend to begin a continuous process of water absorption and loss of soluble components, these known phenomena, respectively, sorption and solubility.^{10,34} Polymer hydrophilicity is directly linked to its chemical composition.³³ Simplified adhesives systems, as Adper Single Bond 2, are composed of methacrylate monomers, which chemical structures contain polar groups (esters and hydroxyls groups) that having affinity for water.^{33,34} Water molecules bind to polar groups through hydrogen bonding and breaks covalent bonds between the polymers, and consequently promotes the leaching of components of the adhesive system.^{33,35} Some studies³⁶⁻³⁸ related high sorption and solubility values to Adper Single Bond 2, corroborating with our results. Thus, it is plausible to believe that the incorporation of the hydrophilic drugs in the adhesive systems could result in an increase of sorption and solubility.¹¹ Epigallocatechin-3-gallate is a polyphenol that have a greater hydrogen bonding capability due to the presence of three vicinal hydroxyl groups from the galloyl moiety.⁴ However, incorporation of EGCG directly at 0.01% and 0.1% or in polymeric microparticules at 1% did not increase the water sorption of simplified adhesive system. One reason for WS results can be attributed chemical properties of poly (lactide-co-glycolide), since it is relatively hydrophobic and water insoluble,³⁹ so it would have

a limited ability to attract water into the polymer. In addition, Pallan *et al.*¹⁰ showed that addition of epigallocatechin-3-gallate up to 2% did not affect WS of polymeric materials. It appears that low concentrations of this polyphenol was not capable of causing significant changes in the polarity of adhesive systems.^{10,11}

One of the consequences of water sorption is the solubility phenomenon.^{34,36} Solubility is the process characterized by leaching the material components of polymeric materials such solvents, unreact monomers and oligomers.³³ In the scientific literature polymeric materials doped directly with Epigallocatechin-3-gallate has shown conflicting solubility results. Pallan *et al.*¹⁰ showed that the increase of solubility values of experimental resins by Epigallocatechin-3-gallate incorporation had concentration-dependent effect. However, when one-step self-etch adhesive system was doped with Epigallocatechin-3-gallate at 0.01% and 0.1% resulted in a decrease of solubility.¹¹ In the present study EGCG did not interfere in SL values, independent of concentration used and incorporation modes (directly or loaded PLGA microparticles) in two-step self-adhesive.

Immediate bond strength depends on the adequate resin monomers penetration into the interfibrillar spaces of the collagen web in the demineralized dentin, leading to the formation of the hybrid layer.⁴⁰ Nevertheless, differences in the composition of adhesive systems can influence in the bond strength of resin-dentin interface.⁴¹ In the current study, the incorporation of epigallocatechin-3-gallate into the adhesive systems did not affect the resin-dentin bond strength after 24h corroborating with Du *et al.*⁹ Epigallocatechin-3-gallate molecules interact with dentin collagen fibrils producing crosslinking and reduces the enzymatic activity of MMPs and CTs, which may be important to the longevity of the adhesive procedures.⁴

Although epigallocatechin-3-gallate incorporation in adhesive systems present promising results in sound dentin, it is necessary more long evaluation periods to confirm the maintenance of resin-dentin bond strength and further studies evaluating the catechin efficiency in the preservation of adhesive interfaces performed in affected caries dentin.

CONCLUSION

Single Bond 2 doped with EGCG-load PLGA 50:50 microparticules at 1% promote controlled release without causing detrimental effects of physicochemical properties.

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TABLES

Table 1 – Adhesive system and bonding procedures

Product	Composition	Manufacturer (#Batch n°)	Application mode
Adper Single Bond 2	Adhesive–Bis-GMA, HEMA, dimethacrylates, silica nanofiller (5 nm), polyalquenoic acid copolymer, initiators, water and ethanol	3M St.Paul, MN, USA (batch #1312201025)	ESPE, 1-two coats of adhesive; 2-air-drying for 10 s at 20 cm; 3-light-curing for 10 s.

Abbreviations: Bis-GMA: bisphenol A diglycidyl methacrylate, HEMA: 2-hydroxyethyl methacrylate. *This brand name is the same product as Adper Scotchbond 1 XT, Adper Single Bond Plus and Adper Single Bond 1 XT.

Table 2. Mean (standard deviation) of physicochemical properties tested.

Groups (<i>n</i> =10)	Degree of conversion %	Flexural Strength (MPa)	Elastic Modulus (GPa)	Water Sorption (mg/mm ³)	Solubility (µg/mm ³)
SB (Control)	59.8 (2.0)	12.2 (3.1) ^a	0.2 (0.06)	172.7 (7.2)	73.4 (5.8)
EGCG 0.01%	58.3 (3.3)	12.4 (2.7) ^a	0.2 (0.1)	175.5 (6.3)	74.7 (2.8)
EGCG 0.1%	59.0 (3.6)	11.9 (3.7) ^a	0.2 (0.09)	173.6 (9.6)	76.3 (5.2)
1% PLGA50:50/EGCG	58.7 (2.8)	16.5 (3.4) ^b	0.2 (0.04)	166.9 (7.4)	77.0 (4.3)

*Distinct superscript letters indicate statistical difference in the same columns ($p < 0.05$).

Table 3 – Bond strength values (MPa \pm SD (*)) according to adhesive systems used.

Groups (<i>n</i> =9)	Adper Single Bond 2	
	24 hours (ns)	
SB	34.6 \pm 6.9 (47)	
EGCG 0.01%	36.6 \pm 5.7 (53)	
EGCG 0.1%	38.6 \pm 4.7 (52)	
1% PLGA50:50/EGCG	35.1 \pm 6.7 (40)	

ns: no statistical significance between values ($p > 0.05$).

(*) corresponds to the number of sticks tested per group.

Table 4 – Distribution of mode of fracture of each group expressed as *n* (relative percentage)

Groups	Adper Single Bond 2				
	24 hours				
	A	M	CR	CD	PF
SB	7 (14)	40 (78)	1 (2)	2 (4)	1(2)
EGCG 0.01%	1 (2)	52 (88)	3 (5)	1 (2)	2(3)
EGCG 0.1%	3 (5)	49 (82)	3 (5)	3 (5)	2(3)
1% PLGA50:50/EGCG	2 (4)	38 (81)	3 (7)	2 (4)	2(4)

A: adhesive failure; M: mixed failure; CR: cohesive failure in resin; CD: cohesive failure in dentin; PF: premature failure

FIGURES

Incorporation mode of EGCG	Adhesive system	Description
-	SB (Control)	-
Directly	SB + EGCG 0.01% (EGCG 0.01%)	Containing 0.01% of EGCG
	SB+EGCG 0.1%	Containing 0.1% of EGCG
Microencapsulated	SB+0.5% PLGA50:50/EGCG	Containing 0.5% (w/w) of microparticles (PLGA50:50) loaded with EGCG (\approx 0.02% EGCG)
	SB+1% PLGA50:50/EGCG	Containing 1.0% (w/w) of microparticles (PLGA50:50) loaded with EGCG (\approx 0.04% EGCG)
	SB+2% PLGA50:50/EGCG	Containing 2.0% (w/w) of microparticles (PLGA50:50) loaded with EGCG (\approx 0.08% EGCG)
	0.5% PLGA75:25/EGCG	Containing 0.5% (w/w) of microparticles (PLGA75:25) loaded with EGCG (\approx 0.02% EGCG)
	SB+1% PLGA75:25/EGCG	Containing 1.0% (w/w) of microparticles (PLGA75:25) loaded with EGCG (\approx 0.04% EGCG)
	SB+2% PLGA75:25/EGCG	Containing 2.0% (w/w) of microparticles (PLGA75:25) loaded with EGCG (\approx 0.08% EGCG)

Figure 1 – Experimental adhesives systems

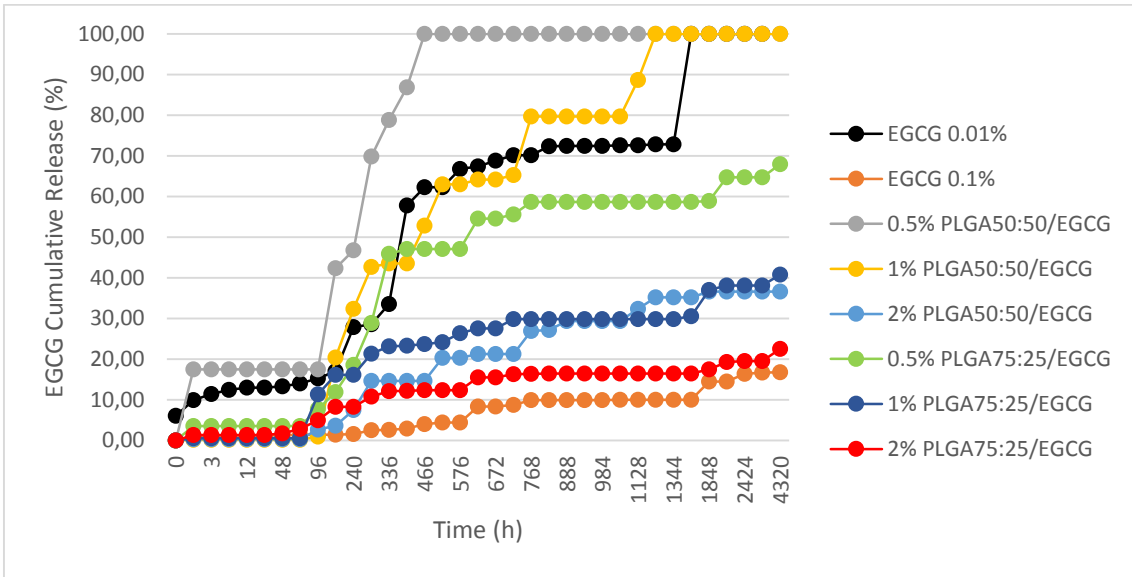


Figure 2. EGCG cumulative release (%) from adhesive systems during the entire evaluation period (4320 hours).

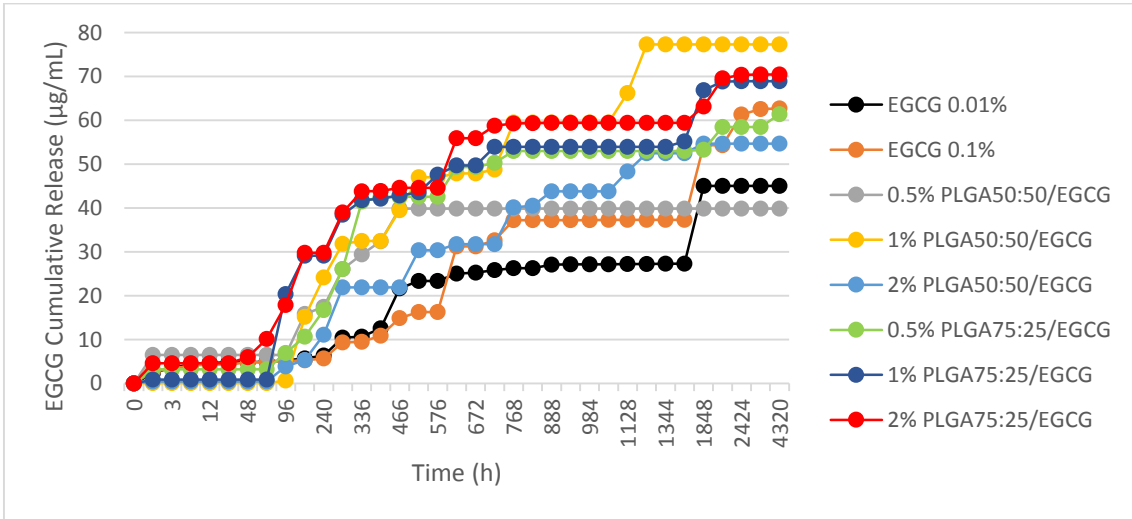


Figure 3. EGCG cumulative release (µg/mL) from adhesive systems during the entire evaluation period (4320 hours).

Conclusão Geral

4 CONCLUSÃO GERAL

Diante dos resultados obtidos no presente estudo e considerando as condições experimentais empregadas, pode-se concluir que:

- 1) O pré-tratamento dentinário com solução ácida de fluoreto de sódio (pH 3,6) pode ser uma alternativa viável para melhorar a longevidade dos procedimentos adesivos executados com sistemas adesivos autocondicionantes de 2 passos;
- 2) A biomodificação dentinária, com o flavonóide epigalocatequina-3-galato e o digluconato de clorexidina, é eficiente em manter a resistência de união de sistema adesivo autocondicionante de passo único ao longo de 6 e 12 meses;
- 3) A incorporação de epigalocatequina-3-galato, em baixas concentrações (0,01% e 0,1%), em um sistema adesivo autocondicionante de passo único reduz a solubilização dos seus componentes. Contudo, o epigalocatequina-3-galato não interfere significativamente nos valores de sorção, grau de conversão e resistência flexural;
- 4) Micropartículas de ácido polilático glicólico carregadas com epigalocatequina-3-galato, especialmente formuladas com PLGA 50:50, poderiam ser uma ferramenta útil para a liberação controlada de fármacos.
- 5) O Single Bond 2 incorporado com micropartículas de PLGA 50:50 carregadas com EGCG a 1% promove liberação controlada da droga sem prejudicar as propriedades físico químicas do sistema adesivo.

Portanto, conclui-se que as diferentes abordagens de biomodificação dentinária, de uma forma geral, além de não prejudicarem as propriedades físicas dos sistemas adesivos estudados, podem também proporcionar benefícios às interfaces de união, podendo mantê-las mais estáveis ao longo do tempo.

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Anexos

ANEXO A – Instruções para os autores do periódico “*European Journal of Oral Sciences*”, referente ao Capítulo 1.

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 - Select the designation of each file in the drop down next to the Browse button.
 - When you have selected all files you wish to upload, click the “Upload Files” button.
- Be sure to upload a complete manuscript with all pages and sections as specified under 5.2 (below). It is of importance that a manuscript is adapted to journal format.
- Before uploading a manuscript, you must turn off Word’s automatic function for tracking of changes in the text. The uploaded manuscript should not display any track-changes.
- Review your submission (in HTML and PDF format) before completing your submission by sending to the Journal. Click the 'Submit' button when you are finished reviewing.

3.3. Manuscript Files Accepted

Manuscripts should be uploaded as Word (.doc) or Rich Text Format (.rft) files (not write-protected). Illustrations/Figures should be uploaded separately as TIFF, EPS, GIF, JPEG, PICT or Bitmap files. Do not embed illustrations in a .doc file and do not use PowerPoint. However, only high-resolution TIFF or EPS files are suitable for printing if the manuscript is accepted for publication. The files will be automatically converted to HTML and PDF on upload and will be used for the review process. The text file must contain the entire manuscript including title page, abstract page, text, references, tables, and figure legends, but no embedded figures. In the text, please reference any figures as “Figure 1”, “Figure 2” etc to match the Tag name you choose for all individual figure files uploaded. Tables may also be uploaded separately. Manuscripts should be formatted as described below. Please note that any manuscripts uploaded as Word 2007 (.docx) will be automatically rejected, implying that any .docx file should be saved as .doc before uploading.

3.4. Suspension of Submission Mid-way in the Submission Process

You may suspend a submission at any phase before clicking the 'Submit' button and save it to make the final submission later. The manuscript can then be located under 'Unsubmitted Manuscripts' and you can click on 'Continue Submission' to continue your submission when you choose to.

3.5. E-mail Confirmation of Submission

After submission you will receive an e-mail to confirm receipt of your manuscript. If you do not receive the confirmation e-mail after 24 hours, please check your e-mail address carefully in the system. If the e-mail address is correct please contact your IT department. The error may be caused by some sort of spam filtering on your e-mail server. Also, the e-mail should be received if the IT department adds our e-mail server (uranus.scholarone.com) to their whitelist.

3.6. Editorial Processing

After a first editorial screening, manuscripts will be forwarded to one of the Journal's Editors for further scientific evaluation and processing. Thus, queries and comments concerning a specific manuscript should primarily be directed to the managing Editor. Manuscripts submitted to the European Journal of Oral Sciences will be reviewed by two or more experts in the field. The European Journal of Oral Sciences uses single blinded review. The names of the reviewers will thus not be disclosed to the author submitting a paper.

3.7. Manuscript Status

You can access ScholarOne Manuscripts (formerly known as Manuscript Central) any time to check your 'Author Centre' for the status of your manuscript. The Journal will inform you by e-mail once a decision has been made.

3.8. Submission of Revised Manuscripts

To upload a revised manuscript, please locate your manuscript under 'Manuscripts with Decisions' and click on 'Submit a Revision'. You should be careful not to upload the revised version under a new manuscript number as if it were another article. Be sure to use the earlier

manuscript number (which will then get an R addendum). Please remember to delete any old files uploaded when you upload your revised manuscript. Do not forget to submit an accompanying letter with itemized answers to all questions and remarks made by the reviewers and the Editor.

4. MANUSCRIPT TYPES ACCEPTED

Original Articles: An original article should comprise a conclusive, full-length scientific investigation. It should describe the rationale behind the study, the materials and methods used, and the results obtained. There should also be a discussion of the implications of the results as well as a list of literature references cited.

Scientific studies investigate phenomena and acquire new knowledge – or correct or integrate previous knowledge. They are based on the collection of data through observation and experimentation, and subject to specific principles of reasoning. The European Journal of Oral Sciences gives priority to analytical articles, investigating why and how something occurred rather than reporting empirical observations.

Review Articles: May be invited by the Editors. Proposals for such articles should be discussed with the appropriate Editor prior to preparation and submission. Review articles comprise attempts to synthesize the existing literature pertaining to a specific scientific question using methods and principles of reasoning that are as transparent as possible. It follows that systematic reviews are preferred over more narrative reviews. Review articles will be subjected to peer review.

Focus Articles: May be invited by the Editors. Proposals for such articles should be discussed with the appropriate Editor prior to preparation and submission. Focus articles may build on the same principles as the Review article, but are usually shorter and aim at stimulating a broader scientific discussion by ‘contesting conventional wisdom’ and allowing the author(s) to argue a specific point pertaining to a matter of current scientific importance. Focus articles will be subjected to peer review.

Short Communications: Short communications should aim at being no longer than two printed pages. They should contain important, new, definitive information of sufficient significance to warrant publication. Short communications need not follow the usual division into Material and methods etc. but should have a short Abstract.

Extra issues: Congress proceedings, larger papers or monographs may be published as Supplements or Part II issues, the full cost being paid by the congress organizer or similar. A condition is that the proposed extra issue is deemed to have a significant scientific value. In some cases, the Journal will partly fund extra issues; this is at the discretion of the Editor-in-Chief. Further information may be obtained from the Editor-in-Chief.

5. MANUSCRIPT FORMAT AND STRUCTURE

It is expected that all manuscripts submitted to the European Journal of Oral Sciences should follow journal format as described in the Author Guidelines and as displayed in recent issues of the Journal. Failure to do so reflects negatively on the work itself and may be a cause for immediate revision or even rejection of a manuscript.

5.1. Format

Language: The language of publication is English. Authors whose native language is not English are strongly advised to obtain assistance from someone proficient in scientific English. Manuscripts not submitted in the proper format or in poor English may be returned without review. A list of independent suppliers of editing services can be found at http://authorservices.wiley.com/bauthor/english_language.asp. All services are paid for and arranged by the author, and use of one of these services does not guarantee acceptance or preference for publication.

Abbreviations, Symbols and Nomenclature: Correct unit abbreviations should be used. Examples include “yr”, “wk”, “d”, “h”, “min”, “s” and “ μm ” rather than “years”, “weeks”, “days”, “hrs”, “minutes”, “sec” and “ μ ”, respectively. For abbreviations of physical and chemical units and symbols, designation of isotopically labelled compounds, abbreviations which may be used without definition etc., the Biochemical Journal web site is a valuable resource. Scientific names of bacteria, binomials in italics, must be given in full when first mentioned. Subsequent mention may abbreviate genus, taking care that this abbreviation is unambiguous (Staph. or Strep. instead of S.).

5.2. Structure

All manuscripts submitted to the European Journal of Oral Sciences should include: Title page, Abstract Page, Introduction, Material and Methods, Results, Discussion, Acknowledgments, References, Figure Legends, Tables, and Figures, arranged in that order.

Authors are urged to consult a recent issue of the Journal to be familiar with style and format. The whole manuscript should be double-spaced, paginated, and submitted in correct English. The beginning of paragraphs should be properly marked with an indent. Avoid end-of-line hyphens.

Title Page: The title page should contain the following information in the order given: 1) the article title; 2) authors' full names without degrees or titles; 3) authors' institutional affiliations including city and country; 4) a running title, not exceeding 40 letters and spaces; 5) name, address, telephone, telefax and e-mail address of the author responsible for correspondence. The title should be concise but informative, include animal species used (if appropriate) and should not include any non-standard acronyms or abbreviations. The Journal does not favour titles of an affirmative character.

Abstract: A separate abstract page should contain the following: 1) authors' surnames and initials; 2) title of manuscript; 3) the abbreviation Eur J Oral Sci; 4) the word Abstract followed by a summary of the complete manuscript; 5) up to five key words according to Index Medicus;

6) name, address, telefax and e-mail address of the author to whom requests for reprints should be sent. This contact information should refer to a professional rather than to a residential/private address.

The Abstract should give a condensed overview of the study, summarizing its background, aim, methodology and results with only few but relevant details, and the authors' principal conclusions. It should be short and concise, without headings and not divided into paragraphs, and with a maximum of 200 words. It should not contain any non-standard acronyms or abbreviations.

5.3 Main Text of Original Articles

Material and Methods: Procedures should be described in such a detail as to make it possible to repeat the work. Subheadings may be used to improve clarity.

It is assumed that authors have considered the ethical aspects of their research and ensured that the work was approved by an appropriate Ethical Committee. This should be stated. In human experimentation, informed consent from individuals must have been given. (See above under 2.2)

Sources of supply of commercial products should be given with the address (town, state and country) in parenthesis.

For an improved quality and transparency, reports of randomized trials must conform to the CONSORT guidelines and will be evaluated in light of the recommendations in this statement. (See above under 2.3)

Since many investigations rely on statistical treatment, authors are advised to consult a person with in-depth statistical knowledge.

If a manuscript describes original nucleotide/amino acid sequence data, these should be submitted to GenBank by the authors and the accession numbers included in the manuscript. (See above under 2.4)

Authors of papers published in the Journal are obliged to honor any reasonable request by qualified investigators for unique propagative materials, such as cell lines, hybridomas, DNA clones and antibodies that are described in the paper.

Results and Discussion: The Results section should clearly and concisely report findings, as a rule in the past tense, without subjective comments and reference to previous literature. Double documentation of data in text, tables or figures is not acceptable. Tables/figures should not include data that can be given in the text in one or two sentences. The Discussion section presents the interpretation of the findings; this is the only proper section for subjective comments. Authors are strongly urged to avoid undue repetition of what has already been reported in Results. For the sake of clarity, the Results section may have subheadings; this is usually not the case with the Discussion.

Acknowledgements: Under acknowledgements please specify contributors to the article other than the authors accredited. This may include recognition of e.g. financial support, gifts of research material, assistance with statistics and language. Please also include specifications of any potential conflict of interests if appropriate.

Short Communications need not follow the usual division into Material and methods etc. but should have a short abstract.

Review and Focus Articles should include a Title page, an Abstract page and a Reference list as regular Original Research Articles. Although a Review article (particularly following a systematic review) may adhere to the format of the Original Research Article, Review and Focus articles need not contain Materials and Methods, Results or Discussion sections, and may instead employ other headings as relevant for the topic addressed.

5.4. References

Number references consecutively in the order in which they are first mentioned in the text. Identify references in texts, tables, and legends by Arabic numerals (within parenthesis). Check to ensure that all listed references are cited in the text. If an author's name is mentioned in the text, small capital letters should be used.

Non-refereed material and, if possible, non-English publications should be avoided. Congress abstracts, unaccepted papers, unpublished observations, and personal communications may not be placed in the Reference list. References to 'unpublished findings' and to 'personal communication' (provided explicit consent has been given by the sources) may be inserted in parentheses in the text. Unpublished articles should be referred to only if proof can be given that they are accepted for publication. Copies of such articles may be requested for evaluation of the manuscript submitted.

Authors are urged to study the examples of correct reference formats given below. For abbreviations of journals, consult the List of the Journals Indexed in Index Medicus. List all authors; do not use et al. in the Reference list. Avoid issue numbers in journal articles. Give first and last page of references in full.

Journals

Standard journal article:

JERNVALL J, THESLEFF I. Reiterative signaling and patterning during mammalian tooth morphogenesis. *Mech Dev* 2000; 92: 19–29.

Article in supplement or special issue:

MUNDY GR. Cellular and molecular regulation of bone turnover. *Bone* 1999; 24 (Suppl): 35S–38S.

Corporate (collective) author:

WHO COLLABORATING CENTRE FOR ORAL PRECANCEROUS LESIONS. Definition of leukoplakia and related lesions: an aid to studies on oral precancer. *Oral Surg Oral Med Oral Pathol* 1978; 46: 518–539.

Unpublished article:

FLEISCHMANNOVA J, MATALOVA E, TUCKER AS, SHARPE PT. Mouse models of tooth abnormalities. *Eur J Oral Sci* 2008; 116: in press.

Books and other monographs:

Personal author(s):

PINDBORG JJ. Atlas of diseases of the oral mucosa, 5th ed. Copenhagen: Munksgaard, 1992; 50–66.

Chapter in book:

RUCH JV. Tooth morphogenesis and differentiation. In: LINDE A, ed. Dentin and dentinogenesis. Vol. I. Boca Raton, FL: CRC Press, 1984; 47–79.

No author given:

International statistical classification of diseases and related health problems. 10th revision, 2nd Ed, Vol 1. Geneva: World Health Organization, 2005; 550–564.

5.5. Tables, Figures and Figure Legends

Tables: Tables should be numbered consecutively with Arabic numerals. Each table should include a compulsory, concise explanatory title and an explanatory legend. A table should be organized with due regard for the proportion of the printed column/page. Specifically, tables which are too wide must be avoided, as these have to be printed vertically.

Figure Legends: Include Figure Legends after the reference section of the Main Text.

Figures: Articles will not be published unless the Figures fulfill journal quality criteria in terms of scientific information, general style, legibility of text and numbers, as well as electronic format and resolution. Double documentation of data in text, tables or figures is not acceptable. Always consider whether data might be better given in the text or in a table. All graphs,

drawings, and photographs are considered Figures and should be numbered in sequence with Arabic numerals. Each figure should have a legend (number and list legends after the reference section of the main text). Figures should be planned to fit the proportions of the printed page or one column's width. Authors are encouraged to arrange micrographs into multipane

6. Early View

The European Journal of Oral Sciences is covered by Wiley-Blackwell's Early View service. Early View articles are complete full-text articles published online in advance of their publication in a printed issue. Articles are therefore available as soon as they are ready, rather than having to wait for the next scheduled print issue. Early View articles are complete and final. They have been fully reviewed, revised and edited for publication, and the authors' final corrections have been incorporated. Because they are in final form, no changes can be made after online publication. The nature of Early View articles means that they do not yet have volume, issue or page numbers, so Early View articles cannot be cited in the traditional way. They are therefore given a Digital Object Identifier (DOI), which allows the article to be cited and tracked before it is allocated to an issue. After print publication, the DOI remains valid and can continue to be used to cite and access the article.

ANEXO B – Registro de Aprovação do Comitê de Ética em Pesquisa da Universidade Federal do Ceará, referente ao Capítulo 1.



Universidade Federal do Ceará
Comitê de Ética em Pesquisa

Of. Nº 74/12

Fortaleza, 16 de Março de 2012.

Protocolo COMEPE nº: 35/12

Pesquisador responsável: Sérgio Lima Santiago.

Título do Projeto: "Efeito do fluoreto de sódio na resistência de união de sistema adesivo autocondicionante à dentina"

Levamos ao conhecimento de V.S^a, que o Comitê de Ética em Pesquisa da Universidade Federal do Ceará – COMEPE, dentro das normas que regulamentam a pesquisa em seres humanos, do Conselho Nacional de Saúde – Ministério da Saúde, Resolução nº 196 de 10 de outubro de 1996 e complementares, aprovou o protocolo e o TCLE do projeto supracitado na reunião do dia 15 de Março de 2012.

Outrossim, informamos, que o pesquisador deverá se comprometer a enviar o relatório final do referido projeto.

Atenciosamente,

Dr. Leonardo A. Freire Bezerra
Coordenador do Comitê
de Ética em Pesquisa
UFCE

ANEXO C – Instruções para os autores do periódico “*Journal of Biomedical Materials Research Part B: Applied Biomaterials*”, referente ao Capítulo 2.

Author Guidelines

Journal of Biomedical Materials Research Part B: Applied Biomaterials
Information for Contributors

Aims and Scope

Journal of Biomedical Materials Research Part B: Applied Biomaterials is an official journal of the Society for Biomaterials, the Japanese Society for Biomaterials, the Australasian Society for Biomaterials, and the Korean Society for Biomaterials. It is a peer-reviewed journal serving the needs of biomaterials professionals who devise, promote, apply, regulate, produce, and market new biomaterials and medical devices. Papers are published on device development, implant retrieval and analysis, manufacturing, regulation of devices, liability and legal issues, standards, reviews of different device areas, and clinical applications. Published manuscripts fit into one of six categories: original research reports, clinical device-related articles, short research and development reports, review, special report, or columns and editorials. Manuscripts from all countries are invited but must be in English. Authors are not required to be members of a Society for Biomaterials.

Types of Articles Considered for Publication

Original Research Reports: Full-length papers consisting of complete and detailed descriptions of a research problem, the experimental approach, the findings, and appropriate discussion. Findings should represent significant new additions to knowledge.

Clinical Device-Related Articles: Full-length papers addressing such issues as material processing, device construction, regulatory matters, clinical trials, and device retrieval.

Reviews: Scholarly and critical topic-oriented reviews that present a state-of-the-art view. While most reviews are solicited, persons interested in contributing may contact the Editor.

Special Reports: Reports of special topic-oriented symposia, device retrieval protocols, or other special reports not described in the above categories, yet of interest to the applied biomaterials research and development community. Potential contributors should contact the Editor before submitting special reports.

Columns and Editorials: While columns and guest editorials are preponderantly solicited, persons interested in becoming columnists or contributing editorials are encouraged to contact the Editor.

Submission of Manuscripts

Online Submission:

Journal of Biomedical Materials Research Part B: Applied Biomaterials is now receiving submitted manuscripts online at <http://mc.manuscriptcentral.com/jbmr-b>.

Submit all new manuscripts online. Launch your web browser and go to <http://mc.manuscriptcentral.com/jbmr-b>. Check for an existing user account. If you are submitting for the first time, and you do not find an existing account, create a new account. Follow all instructions.

At the end of a successful submission, a confirmation screen with manuscript number will appear and you will receive an e-mail confirming that the manuscript has been received by the journal. If this does not happen, please check your submission and/or contact tech support using the GetHelp Now link in the right corner of any screen.

Upon Acceptance: Manuscript files will now automatically be sent to the publisher for production. It is imperative that files be in the correct format to avoid a delay in the production schedule.

JBMR Part B has adopted a policy that requires authors to make a statement concerning potential conflict of interest relating to their submitted articles. The Editorial Board asks authors of original reports and reviews to disclose, at the time of submission: (1) any financial or employment arrangements they may have with a company whose product figures prominently in the submitted manuscript or with a company making a competitive product; and (2) any grants or contracts from a government agency, a nonprofit foundation, or a company supporting the preparation of the manuscript or the described research. This information will be available to the reviewers of the manuscript. If the article is accepted for publication, the editor will discuss with the authors the manner in which such information may be communicated to the reader.

At the time of submission, JBMR Part B asks authors to certify that all animals utilized in their research were cared for according to the policies and principles established by the Animal Welfare Act and the NIH Guide for Care and Use of Laboratory Animals.

Review Process: All original reports and reviews receive critical review by at least two reviewers with expertise in the major subject area of the paper. Reviewers may recommend "Acceptance as is," "Acceptance with modification," or "Rejection." If modification is required, the manuscript is returned to the author(s). The revised manuscript is then re-reviewed by the original reviewers, and even re-revised if necessary. Differences in opinion are resolved by submission either to a third reviewer or the Editor.

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For authors signing the copyright transfer agreement: If the OnlineOpen option is not selected the corresponding author will be presented with the copyright transfer agreement (CTA) to sign. The terms and conditions of the CTA can be previewed in the samples associated with the Copyright FAQs below:

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Organization and File Formats

Manuscript: For optimal production, prepare manuscript text in size 12 font on 8-1/2 x 11 inch page, double-spaced, with at least 1-inch margins on all sides. Text files should be formatted as .doc or .rtf files. The results and discussion sections must be written separately and cannot be combined. Refrain from complex formatting; the Publisher will style your manuscript according to the Journal design specifications. Do not use desktop publishing software such as PageMaker or Quark Xpress or other software such as Latex. If you prepared your manuscript with one of these programs, export the text to a word processing format. Please make sure your word processing programs "fast save" feature is turned off. Please do not deliver files that contain hidden text: for example, do not use your word processor's automated features to create footnotes or reference lists. Manuscripts including references (but not figures or tables) should be no longer than 18 pages.

Please be sure to submit your illustrations and tables as separate files; the system will automatically create a pdf file of your paper for the reviewers.

Original research and short reports should appear in the following order: title page (including authors and affiliations), abstract, keywords, introduction, materials and methods, results, discussion, acknowledgments, references, figure legends. Number pages consecutively starting with the title page as page 1. Abbreviations must conform to those listed in Council of Biology Editors' CBE Style Manual, 5th Edition.

When mentioning a material, chemical reagent, instrument, or other product, use the generic name only. If further identification (proprietary name, manufacturer's name and address) is absolutely required, list it in parentheses.

Title Page: List the full title of the paper and each author's full name (first name, middle initial(s), surname), department, institution, city, and state (and country if other than the United States). Indicate the name and address of the author to whom reprint requests should be sent.

Abstract and Keywords: Include an abstract of about 200 words maximum summarizing the aims, findings, and conclusions of the paper. Below the abstract, list five keywords or phrases that best characterize the subject matter of the manuscript.

Running Heads: Supply a short title of no more than 65 characters, including spaces and punctuation, to be used for running head copy.

References: Number references consecutively as they appear in the text. Material accepted for publication but not yet published may be listed in the References, but unpublished observations, personal communications, and material submitted for publication but not yet accepted should be cited parenthetically within the text (and not included among the numbered references). Style references entries using the Council of Biology Editors Style Manual, 5th Edition formats:

For journal articles:

Alexander A, Green WS. Total hip replacements: A second look. *J Soc Biomater* 1989;45:345–366.

For books/chapters:

Ricci JL, Guichet J-M. Total hip replacement: A third look. Cindra AB, Franklin DE, editors. *State of the art orthopaedics*, vol 3, Hips. New York: Wiley; 1988:56–59.

For abstracts:

Davidson GRH. Total hip replacement: A fifth look. *Trans ABCS* 1987;22:341–345.

For presentations:

Goodenough T. Total hip replacement: A sixth look. Presented at the 3rd Annu Mtg Orthop Res Soc, Boston, December 5–7, 1989.

Figure Legends: Please supply complete captions for all figures. Captions are to appear on a separate page at the end of the manuscript.

Tables: Please save Tables separately and supply numbers and titles for all. All table columns should have an explanatory heading. Tables should be submitted as doc or rtf files (it is preferred that tables are prepared using Word's table edit tool.)

Illustrations: When preparing digital art, please consider:

Resolution:

The minimum requirements for resolution are:

1200 DPI/PPI for black and white images, such as line drawings or graphs.

300 DPI/PPI for picture-only photographs

600 DPI/PPI for photographs containing pictures and line elements, i.e., text labels, thin lines, arrows.

These resolutions refer to the output size of the file; if you anticipate that your images will be enlarged or reduced, resolutions should be adjusted accordingly.

Formats:

For the editorial review process, GIF and JPEG files are acceptable; upon submission of a revision, TIFF or EPS files will be required. For the editorial review process, color images may be submitted in RGB color; upon revision, CMYK color will be required. Delivery of production-quality files early in the review process may facilitate smooth and rapid publication once a manuscript has been accepted.

Note that these file formats are not acceptable for printing: JPG, GIF, PNG, PCX, PNG, XBM, Word, and Excel. We recommend creating your graphics in Photoshop, Illustrator, or Freehand and importing them into your page applications as TIFFs with all fonts included. Do not scan figures as JPEGs and convert to TIFFs. For further guidance on preparing digital figure files, authors are encouraged to visit <http://cjs.cadmus.com/da/applications.asp>.

To ensure that your digital graphics are suitable for print purposes, please go to RapidInspector™ at <http://rapidinspector.cadmus.com/zwi/index.jsp>. This free, stand-alone software application will help you to inspect and verify illustrations right on your computer.

A legend must be provided for each illustration and must define all abbreviations used therein. Legends should be placed at the end of the manuscript text file.

Color Illustrations: Color figures are generally printed in the Journal at the author's expense. The publisher will provide cost estimates prior to printing. A limited number of color figures that are of critical importance and that significantly enhance the presentation will be considered for publication at the publisher's expense subject to editorial recommendation. Final decision on publication of color figures will be at the discretion of the Editor. All color figures will be reproduced in full color in the online edition of the journal at no cost to authors. For best reproduction, bright, clear colors should be used. Dark colors against a dark background do not reproduce well; please place your color images against a white background wherever possible.

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Note to NIH Grantees: Pursuant to NIH mandate, Wiley-Blackwell will post the accepted version of contributions authored by NIH grant-holders to PubMed Central upon acceptance. This accepted version will be made publicly available 12 months after publication. For further information, see www.wiley.com/go/nihmandate.

ANEXO D – Registro de Aprovação do Comitê de Ética em Pesquisa da Universidade Federal do Ceará, referente ao Capítulo 2.



Universidade Federal do Ceará
Comitê de Ética em Pesquisa

Of. Nº 105/11

Fortaleza, 23 de maio de 2011

Protocolo COMEPE nº 103/11

Pesquisador responsável: Sérgio Lima Santiago

Título do Projeto: "Efeito de soluções de pré-tratamento dentinário na resistência de união de sistemas adesivos simplificados à dentina"

Levamos ao conhecimento de V.S^a. que o Comitê de Ética em Pesquisa da Universidade Federal do Ceará – COMEPE, dentro das normas que regulamentam a pesquisa em seres humanos, do Conselho Nacional de Saúde – Ministério da Saúde, Resolução nº 196 de 10 de outubro de 1996 e complementares, aprovou o protocolo e o TCLE do projeto supracitado na reunião de nº 140, realizada em 12/05/2011.

ANEXO E – Instruções para os autores do periódico “*Brazilian Dental Journal*”, referente ao Capítulo 3.

Author Guidelines

Brazilian Dental Journal
Information for Contributors

Scope and policy

The Brazilian Dental Journal publishes Full-Length Papers, Short Communications and Case Reports, dealing with dentistry or related disciplines. Only original papers will be considered for publication. In submitting a manuscript, the authors should state in the cover letter that the

material has not been published previously and is not under consideration by another journal in either electronic or printed versions.

ELECTRONIC ADDRESS FOR SUBMISSION

<http://mc04.manuscriptcentral.com/bdj-scielo>

MANUSCRIPTS MUST BE SUBMITTED IN ENGLISH. Authors whose primary language is not English must have their manuscript reviewed by someone proficient in English. Manuscripts accepted for publication will be submitted to the Technical Review for revision of English grammar and scientific writing and to fit the text into the Journal's standards. The cost of the Technical Review will be charged to the authors. Submission of a manuscript to BDJ implies the acceptance of these terms. The decision of acceptance for publication relies on the Editors and is based on the recommendation of the Editorial Board and/or ad hoc reviewers. Authors of manuscripts not recommended for publication will receive an email explaining the decision. The concepts emitted in the papers published in the BDJ are the sole responsibility of the authors, not necessarily reflecting the Editorial Board's opinion.

Form and preparation of manuscripts

THE FOLLOWING GUIDELINES MUST BE FOLLOWED CAREFULLY.

General

- The authors must submit the manuscript in Word and in PDF, comprising the title page, text, tables, figure captions and figures (photographs, micrographs, radiographs, schematic drawings, graphs, computer-generated images, etc).
- The manuscript must be typed in Times New Roman 12 font, with 1.5 spacing, 2.5-cm margins at each side. DO NOT USE bold letters, watermarks or other resources to make the text visually attractive.

Pages should be numbered consecutively, starting with the summary.

Full-length manuscripts are assembled in the following sections:

- 1) Title Page
- 2) Summary and Key Words
- 3) Introduction; Material and Methods; Results; Discussion
- 4) Summary in Portuguese (an item necessary for Latin American Indexing Services that will be provided for non-Brazilian authors by the Journal)

- 5) Acknowledgements (if any)
- 6) References
- 7) Tables
- 8) Figure captions
- 9) Figures

All titles of sections (Introduction, Material and Methods, etc) must be capitalized in regular font type (not bold).

Results and Discussion MUST NOT be joined in a single section.

Short Communications and Case Reports should be divided into appropriate sections.

Products, equipments and materials: the trade name must be followed by the manufacturer's name, city, state and country, within parentheses upon first mention. For further mentions, only the manufacturer's name is required.

All abbreviations must be explained at first mention.

Title page

The first page must contain the title of the manuscript, a short title (maximum of 40 characters, to be used as a running head), author(s) name(s) (no more than 6) and their Department(s), School(s) and/or University (s). DO NOT INCLUDE the author's titles (DDS, MSc, PhD, etc.) or position (Professor, Graduate student, etc.).

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The first page of the manuscript must contain: title of the manuscript, short tile with no more than 40 characters, and NO authors' names or identification.

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The second page should contain a summary of no more than 250 words, stating the aims, methods, results, and any conclusions drawn from the study. Do not use topics and paragraphs and do not cite references in the Summary.

A list of key words (no more than 5) should be included below the summary in lowercase letters, separated by commas.

Introduction

Summarize the purpose of the study, giving only pertinent references. Do not review existing literature extensively. State clearly the working hypothesis.

Material and Methods

Material and methods should be presented in sufficient detail to allow confirmation of the observations. Indicate the statistical methods used, if applicable.

Results

Present the results in a logical sequence in the text, tables and figures, emphasizing the important information.

Do not repeat in the text data contained in the tables and illustrations. The important observations should be emphasized.

Do not repeat the same data in tables and figures.

Describe the statistical data in this section.

Discussion

Summarize the findings without repeating in detail the data given in the Results section.

Relate your observations to other relevant studies and point out the implications of the findings and their limitations. Cite pertinent studies.

Present your conclusions at the end of the Discussion, indicating how your study is pertinent and/or its clinical implications. Presentation of the conclusions in topics should be avoided.

Summary in Portuguese (for Brazilian authors only)

The Summary in Portuguese should be IDENTICAL to the English version (Summary). DO NOT INCLUDE title and key words in Portuguese.

Acknowledgements

Financial support by government agencies should be acknowledged. If appropriate, technical assistance or assistance from colleagues may be acknowledged.

References

References must follow the Journal's style. Authors should refer to a current issue of the BDJ for guidance on reference citation and presentation of the reference list.

References must be numbered consecutively in the text in order of citation, within parentheses, without space between numbers: (1), (3,5,8), (10-15). DO NOT USE superscript numbers.

For papers with two authors, cite both authors in the text, as follows: Ex: "According to Santos and Silva (1)...". If there are more than 3 authors, cite only the first author and add "et al.". Ex: "Pécora et al. (2) reported that..."

All authors of each paper should be included in the Reference List unless there are 7 or more. In this case, the first 6 authors should be given, followed by "et al.".

The reference list must be typed at the end of the manuscript in numerical sequence. No more than 25 references may be cited.

Citation of abstracts and books, as well as articles published in non-indexed journals should be avoided, unless absolutely necessary. Do not cite references in Portuguese.

Abbreviations of journal titles should conform to those used in Dental Index. The style and punctuation of references must follow the format illustrated below:

Journal articles

1. Lea SC, Landini G, Walmsley AD. A novel method for the evaluation of powered toothbrush oscillation characteristics. *Am J Dent* 2004;17:307-309.

Book

2. Shafer WG, Hine MK, Levy BM. *A Textbook of Oral Pathology*. 4th ed. Philadelphia: WB Saunders; 1983.

Chapter in a Book

3. Walton RE, Rotstein I. Bleaching discolored teeth: internal and external. In: *Principles and Practice of Endodontics*. Walton RE (Editor). 2nd ed. Philadelphia: WB Saunders; 1996. p 385-400.

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Each table with its title must be typed after the text. Tables should be numbered with Arabic numerals. DO NOT USE vertical lines, bold letters and capital letters (except the initials).

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Tables must contain all necessary information and be understandable without allusions to the text.

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ANEXO F – Instruções para os autores do periódico “*International Journal of Pharmaceutics*”, referente ao Capítulo 4.

International Journal of Pharmaceutics

Information for Contributors

The International Journal of Pharmaceutics publishes innovative papers, reviews, mini-reviews, rapid communications and notes dealing with physical, chemical, biological, microbiological and engineering studies related to the conception, design, production, characterisation and evaluation of drug delivery systems in vitro and in vivo. "Drug" is defined as any therapeutic or diagnostic entity, including oligonucleotides, gene constructs and radiopharmaceuticals.

Areas of particular interest include: pharmaceutical nanotechnology; physical pharmacy; polymer chemistry and physical chemistry as applied to pharmaceutics; excipient function and characterisation; biopharmaceutics; absorption mechanisms; membrane function and transport; novel routes and modes of delivery; responsive delivery systems, feedback and control mechanisms including biosensors; applications of cell and molecular biology to drug delivery;

prodrug design; bioadhesion (carrier-ligand interactions); and biotechnology (protein and peptide formulation and delivery).

Note: For details on pharmaceutical nanotechnology, see Editorials in 279/1-2 281/1, and 288/1.

Types of paper

(1) Full Length Manuscripts

(2) Rapid Communications

(a) These articles should not exceed 1500 words or equivalent space.

(b) Figures should not be included otherwise delay in publication will be incurred.

(c) Do not subdivide the text into sections. An Abstract should be included as well as a full reference list.

(3) Notes

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(b) Do not subdivide the text into sections. An Abstract and reference list should be included.

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Authorship

All authors should have made substantial contributions to all of the following: (1) the conception and design of the study, or acquisition of data, or analysis and interpretation of data, (2) drafting the article or revising it critically for important intellectual content, (3) final approval of the version to be submitted.

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Introduction

State the objectives of the work and provide an adequate background, avoiding a detailed literature survey or a summary of the results.

Material and methods

Provide sufficient detail to allow the work to be reproduced. Methods already published should be indicated by a reference: only relevant modifications should be described.

Results

Results should be clear and concise.

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This should explore the significance of the results of the work, not repeat them. A combined Results and Discussion section is often appropriate. Avoid extensive citations and discussion of published literature.

Conclusions

The main conclusions of the study may be presented in a short Conclusions section, which may stand alone or form a subsection of a Discussion or Results and Discussion section.

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3. Three or more authors: first author's name followed by 'et al.' and the year of publication.

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List: References should be arranged first alphabetically and then further sorted chronologically if necessary. More than one reference from the same author(s) in the same year must be identified by the letters 'a', 'b', 'c', etc., placed after the year of publication.

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Strunk Jr., W., White, E.B., 2000. *The Elements of Style*, fourth ed. Longman, New York.

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JOURNAL OF DENTISTRY

AUTHOR INFORMATION PACK

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The Abstract should not exceed 250 words and should be presented under the following subheadings: Objectives, Methods; Results; Conclusions (For Reviews: Objectives; Data; Sources; Study selection; Conclusions). A 50 word 'Clinical Significance' statement should appear at the end of the abstract advising readers of the clinical importance and relevance of their work. These subheadings should appear in the text of the abstract. Please repeat the title of the article at the top of the abstract page.

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Although a graphical abstract is optional, its use is encouraged as it draws more attention to the online article. The graphical abstract should summarize the contents of the article in a concise, pictorial form designed to capture the attention of a wide readership. Graphical abstracts should be submitted as a separate file in the online submission system. Image size: Please provide an image with a minimum of 531 × 1328 pixels (h × w) or proportionally more. The image should be readable at a size of 5 × 13 cm using a regular screen resolution of 96 dpi. Preferred file types: TIFF, EPS, PDF or MS Office files. See <http://www.elsevier.com/graphicalabstracts> for examples.

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Provide a maximum of 6 keywords, using British spelling and avoiding general and plural terms and multiple concepts (avoid, for example, 'and', 'of'). Be sparing with abbreviations: only abbreviations firmly established in the field may be eligible. These keywords will be used for indexing purposes.

Artwork

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Please submit tables as editable text and not as images. Tables can be placed either next to the relevant text in the article, or on separate page(s) at the end. Number tables consecutively in accordance with their appearance in the text and place any table notes below the table body. Be sparing in the use of tables and ensure that the data presented in them do not duplicate results described elsewhere in the article. Please avoid using vertical rules.

References

Citation in text

Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. Unpublished results and personal communications are not recommended in the reference list, but may be mentioned in the text. If these references are included in the reference list they should follow the standard reference style of the journal and should include a substitution of the publication date with either 'Unpublished results' or 'Personal communication'. Citation of a reference as 'in press' implies that the item has been accepted for publication.

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Journal articles

Lynch CD, Frazier KB, McConnell RJ, Blum IR, Wilson NHF. State-of-the-art techniques in Operative Dentistry: contemporary teaching of posterior composites in UK and Irish dental schools. *British Dental Journal* 2010; **209**: 129 - 36.

Wilson NHF, Mjör I. The teaching of class I and class II direct composite restorations in European dental schools. *Journal of Dentistry* 2000; **28**: 15-21.

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Books

Lynch CD. *Successful posterior composites*. London: Quintessence Publishing Co., 2008.

Book chapters

Phillips SJ, Whisnant JP. The role of dentine under restorations. In: Laragh JH, Brenner BM, editors. *The science of restorative dentistry*. 2nd ed. Oxford: Elsevier; 2003. p.266-78.

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One author has been designated as the corresponding author with contact details:

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All necessary files have been uploaded, and contain:

- Keywords
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Further considerations

- Manuscript has been 'spell-checked' and 'grammar-checked'
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ANEXO H – Registro de Aprovação do Comitê de Ética em Pesquisa da Universidade Federal do Ceará, referente ao Capítulo 5.

UNIVERSIDADE FEDERAL DO
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PARECER CONSUBSTANCIADO DO CEP

DADOS DO PROJETO DE PESQUISA

Título da Pesquisa: EFEITO DA INCORPORAÇÃO DE CATEQUINA EM SISTEMA ADESIVO CONVENCIONAL NA RESISTÊNCIA DE UNIÃO À DENTINA

Pesquisador: Nadine Luisa Soares de Lima Guimarães

Área Temática:

Versão: 1

CAAE: 22468813.0.0000.5054

Instituição Proponente: Departamento de Odontologia Restauradora

Patrocinador Principal: Departamento de Odontologia Restauradora

DADOS DO PARECER

Número do Parecer: 459.659

Data da Relatoria: 14/11/2013

Apresentação do Projeto:

Os materiais adesivos são bastante utilizados na odontologia e têm demonstrado excelentes resultados na prática clínica diária. Atualmente, os sistemas adesivos são classificados em convencionais e autocondicionantes. O presente estudo como objetivo comparar o efeito do flavonóide Epigallocatequina-3-galato (EGCG) incorporado ao sistema adesivo convencional ou como pré-tratamento da dentina na resistência de união, através de ensaios de microtração imediatos e após armazenagem por 6 e 12 meses.

Serão utilizados 40 terceiros molares humanos hígidos. Após remoção do esmalte e dentina oclusal, utilização da lixa de carbeto de silício (SiC) N°

100 acoplada em uma polítrix elétrica para completa remoção de esmalte e a de N° 600 durante 1 minuto para gerar um padrão de smear layer em

todos os espécimes, os espécimes serão distribuídos aleatoriamente em 5 Grupos (n=8): Grupo I (Água destilada + Adper_z Single Bond 2®); Grupo

II (Água destilada + EGCG puro incorporado ao Adper_z Single Bond 2®); Grupo III (EGCG em cápsula incorporado ao Adper_z Single Bond 2®);

Grupo IV (Solução aquosa de EGCG + Adper_z Single Bond 2®) e; Grupo V (Solução de EGCG cápsula + Adper_z Single Bond 2®). Primeiramente,

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Bairro: Rodolfo Teófilo CEP: 60.430-270
UF: CE Município: FORTALEZA
Telefone: (85)3368-8344 Fax: (85)3223-2903 E-mail: comepe@ufc.br

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será feito condicionamento das superfícies dentárias com ácido fosfórico por 15 segundos, em seguida essas superfícies serão lavadas com spray ar/água por 15 segundos, secas por 30 segundos, e reumidecidas com as soluções de limpeza de acordo com cada Grupo (15 μ L, ativamente por 60s). O excesso será removido com papéis absorventes, deixando a superfície dentinária visivelmente úmida. Na sequência, será aplicado o sistema adesivo Adper ξ Single Bond 2 ® de formulação específica para cada Grupo, de acordo com as recomendações do fabricante. A porção coronária será reconstruída com 4 incrementos (1 mm cada) de resina composta (Filtek Z250XT ®). Após estocagem em água destilada a 37 $^{\circ}$ C por 24h, espécimes em forma de palito (constituídos de resina e dentina unidas pela interface adesiva) serão obtidos por meio de cortes seriados dos dentes. Após a obtenção dos palitos, estes serão submetidos a uma força de tração em uma máquina de ensaios universais à velocidade de 0,5 mm/ minuto até que ocorra a fratura. Os palitos que irão ser testados após 6 meses e 1 ano ficarão armazenados em em água destilada. Serão avaliados os modos de fratura com o auxílio de uma Lupa Estereoscópica e classificadas em: 1) Falha coesiva em resina composta (FCR); 2) Falha coesiva em dentina (FCD); 3) Falha adesiva (FA); e 4) Falha mista (FM). A partir dos dados obtidos em Mpa será realizado teste de normalidade e, se constatada a presença de distribuição normal das amostras, os dados serão submetidos ao teste ANOVA (Análise de Variância) para comparação entre os grupos e em seguida, eventuais diferenças entre os grupos serão analisadas por teste de comparação múltipla. No caso dos resultados falharem no teste de normalidade, será aplicado teste Kruskal-Wallis. O nível de significância será de p0,05 em todas as situações.

Objetivo da Pesquisa:

Comparar o efeito do flavonóide Epigallocatequina-3-galato (EGCG) incorporado ao sistema adesivo convencional ou como pré-tratamento da dentina na resistência de união, através de ensaios de microtração imediatos e após armazenagem por 6 e 12 meses.

Avaliação dos Riscos e Benefícios:

Riscos:

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Continuação do Parecer: 459.659

Não há riscos envolvidos pois trata-se de uma pesquisa em laboratório com dentes humanos extraídos por motivos que não envolvem este estudo.

Benefícios:

De acordo com os resultados obtidos, será possível propor novas técnicas e materiais a serem usados na odontologia restauradora.

Comentários e Considerações sobre a Pesquisa:

Pesquisa relevante na área de dentística com metodologia clara e objetiva e compatível com o objetivo do trabalho

Considerações sobre os Termos de apresentação obrigatória:

Apresentou: folha de rosto, dispensa do TCLE, termo de doação de dentes, orçamento detalhado, currículo Lattes, carta de apreciação ao COMEP, cronograma, autorização do local onde será realizada a pesquisa, declaração de concordância, declaração de vínculo da aluna no mestrado, declaração de custeio da pesquisa, projeto com anexos.

Recomendações:

Conclusões ou Pendências e Lista de Inadequações:

Não se aplica.

Situação do Parecer:

Aprovado

Necessita Apreciação da CONEP:

Não

Considerações Finais a critério do CEP:

FORTALEZA, 18 de Novembro de 2013

Assinador por:

FERNANDO ANTONIO FROTA BEZERRA
(Coordenador)

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