



Redescription of *Hatschekia exigua* Pearse, 1951 (Copepoda: Hatschekiidae), parasitic on the Squirrelfish *Holocentrus adscensionis* (Osbeck, 1765) (Actinopterygii: Holocentridae) off Ceará, Northeastern Brazil, with first description of the male

João Victor Couto · Anna Júlia Pontes ·
Caroline Vieira Feitosa ·
Felipe Bisaggio Pereira · Fabiano Paschoal

Received: 17 November 2023 / Accepted: 22 January 2024 / Published online: 3 April 2024
© The Author(s), under exclusive licence to Springer Nature B.V. 2024

Abstract The original description of *Hatschekia exigua* Pearse, 1951 neglected innumerable features of taxonomic value as well as morphometric data and illustrations. Posteriorly, other author tried to access the type material, but their poor state of preservation compromised a detailed redescription. Since then, this species is in need for new morphological data, mainly from fresh material. In the present work, three specimens of *Holocentrus adscensionis* from Mucuripe

Bay, Fortaleza, State of Ceará, Northeastern Brazil, were parasitized by copepods on their gills. Parasites were fixed and preserved in 80% ethanol and cleared in 85% lactic acid for morphological observations using light microscopy. The females were identified as *Hatschekia exigua* by the cephalothorax representing about one-fourth of total body length, with lateral margins expanded into lateral lobes, first exopod with basal segment armed with one outer setae and terminal segment with three setae, first endopod with basal segment unarmed and terminal segment with five setae, leg 3 reduced to two setae and leg 4 reduced to single seta. Comparison with the type series revealed morphological differences in trunk and small appendages, which may be related to alterations in the specimens, caused by the mounting methodology and poor preservation. A detailed morphological analysis of the male revealed for the first time that they differ from their closest congeners by having five setae on the last endopodal segment of leg 1, by smooth intercoxal sclerites on legs 1 and 2 and by a proximolateral process on the third segment of antenna. Moreover, this work represents the first report of *H. exigua* in Brazil and the first hatschekiid copepod found off the coast of Ceará, highlighting that the diversity of Hatschekiidae in this oceanographic region still needs further investigation.

J. V. Couto · F. B. Pereira
Programa de Pós-Graduação em Parasitologia, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Av. Antônio Carlos, 6627, Pampulha, Belo Horizonte, MG 31270-901, Brazil

A. J. Pontes
Faculdade de Ciências Biológicas e Saúde, Universidade do Estado do Rio de Janeiro, Campus Zona Oeste, Av. Manuel Caldeira de Alvarenga, 1203, Campo Grande, Rio de Janeiro, RJ 23070-200, Brazil

C. V. Feitosa
Instituto de Ciências do Mar - Labomar, Universidade Federal do Ceará, Avenida da Abolição, 3207, Fortaleza, CE 60165-081, Brazil

F. Paschoal (✉)
Programa de Pós-Graduação em Biodiversidade e Conservação, Departamento de Oceanografia e Limnologia, Universidade Federal do Maranhão, Av. dos Portugueses, 1966, Bacanga, São Luís, MA 65080-805, Brazil
e-mail: paschoalfabiano@gmail.com

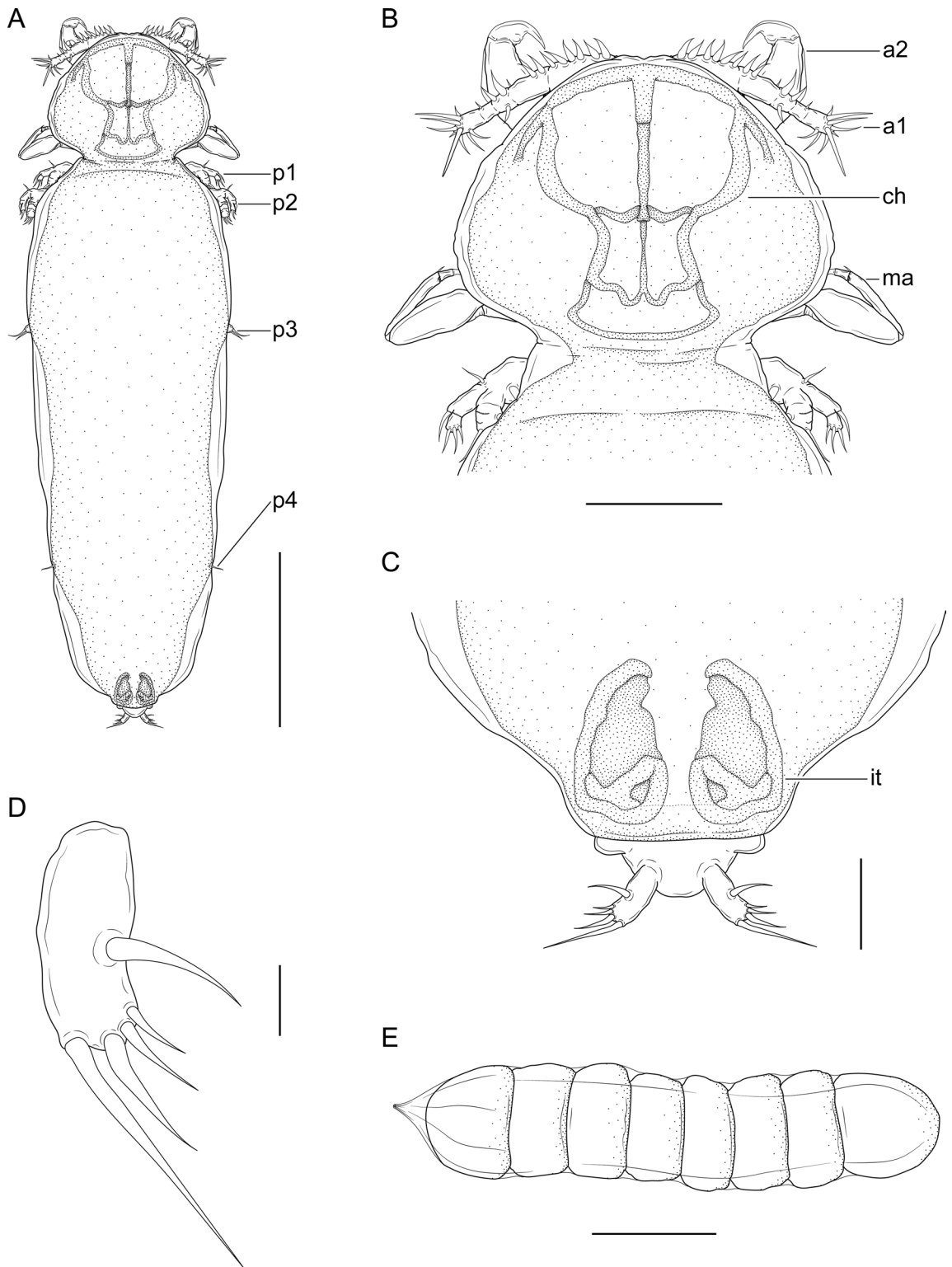


Fig. 1 *Hatschekia exigua* Pearse, 1951 (adult female). **A** habitus, dorsal, p1= leg 1, p2= leg 2, p3= leg 3, p4= leg 4; **B** detail of cephalothorax, dorsal, a1= antennule, a2= antenna, ch= dorsal chitinous

supporting frame, ma= maxilla; **C** posterior part of trunk, ventral, it= internal tissues; **D** caudal ramus, ventral; **E** egg sac, lateral. Scale bars: **A**= 300 μm, **B**= 100 μm, **C**= 40 μm, **D**= 20 μm, **E**= 400 μm

Introduction

Siphonostomatoid copepods of the family Hatschekiidae Kabata, 1979 represent one of the most diverse groups of parasitic copepods commonly found on the gills of marine actinopterygians (Boxshall & Halsey, 2004; Uyeno & Ali, 2013; Paschoal et al., 2022). Currently, this family comprises about 168 species, distributed within nine valid genera: *Bassettithia* Wilson, 1922, *Brachihatschekia* Castro-Romero & BaezaKuroki, 1989, *Congericola* Beneden, 1854, *Hatschekia* Poche, 1902, *Laminohatschekia* Boxshall, 1989; *Mihbaicola* Uyeno, 2013, *Prohatschekia* Nunes-Ruivo, 1954, *Pseudocongericola* Yü, 1933 and *Wynnoweria* Boxshall, 1987 (Walter & Boxshall, 2023).

Although *Hatschekia* represents the most speciose genus within Hatschekiidae, with about 149 species described worldwide (Paschoal et al., 2022; Walter & Boxshall, 2023), research on these copepods in the Southwestern Atlantic is scarce, with only three species reported so far, i.e., *Hatschekia conifera* Yamaguti, 1939 from the Atlantic Pomfret *Brama brama* (Bonnaterre) (Bramidae) off Mar del Plata and San Matías gulf, Argentina, *Hatschekia nagasawai* Paschoal, Couto, Pereira & Luque, 2022 from the Porkfish *Anisotremus virginicus* (Linnaeus) (Haemulidae) off Rio de Janeiro, Brazil and *Hatschekia priacanthus* Izawa, 2016, from the Atlantic Bigeye *Priacanthus arenatus* (Cuvier) (Priacanthidae) off Rio de Janeiro, Brazil (Luque & Tavares, 2007; Cantatore et al., 2012; Paschoal et al., 2022). Additionally, there are two undetermined species of *Hatschekia*, from the Coney *Cephalopholis fulva* (Linnaeus) (Serranidae) and the Bluewing *Prionotus punctatus* (Bloch) (Triglidae), off Rio de Janeiro, Brazil (Paschoal et al., 2022).

Hatschekia exigua Pearse, 1951 was originally described by Pearse (1951) based on three females collected from the gills of *Holocentrus adscensionis* Osbeck (Holocentridae), off Bimini, Bahamas. Despite the remarkable sampling effort and the relevance of his work, the original description of *H. exigua* is poorly detailed, lacking some structures with taxonomic importance and morphometric data, which can mislead the diagnosis and identification by other researchers (Jones, 1985). Subsequently, this species was revisited by Jones (1985) in a revision of the genus, pointing several morphological incongruences

with the original description, as the segmentation and armature of the antennule and the morphology of leg I rami. Jones (1985) also affirmed that the type specimens were distorted and poorly preserved, compromising the observation of important morphological features and hindering a detailed redescription. Moreover, the author stated that a full redescription of *H. exigua* is needed, based on fresh material, collected from the type host (Jones, 1985).

During a survey of parasitic copepods on *Ho. adscensionis* from the Brazilian coastal zone, some specimens of *H. exigua* were recovered from the fish gills. In this paper, we provide a complete and detailed redescription of the female and the first description of the male of this species.

Materials and Methods

Three specimens of *Ho. adscensionis* (total length 25.3–29; mean \pm standard deviation 27.6 ± 2) were bought from local fishermen in the Mucuripe Bay (3°43'18"S, 38°28'51"W), Fortaleza, State of Ceará, Northeastern Brazil. Copepods were collected from the gills, fixed, and preserved in 80% ethanol until morphological analysis. For microscopical observation, specimens were cleared in 85% lactic acid and the appendages were dissected and examined using the wooden slide procedure described by Humes and Gooding (1964). Drawings were made using a Nikon Eclipse Ei (Nikon Instruments Inc, New York, USA) attached to an image capture system. Measurements were based on 14 adult females and six males, using the parameters proposed by Uyeno and Nagasawa (2009a; see Fig. 1), given as the range followed by mean and standard deviation in parentheses, all in micrometers. The type material of *H. exigua* deposited in the invertebrate collection of the National Museum of Natural History (accession code NMNH-88531), Smithsonian Institution of USA, studied by Pearse (1951) was also examined. The morphological terminology follows Huys and Boxshall (1991). Ecological terminology adopted for parasites is according to Bush et al. (1997). Host identification was based on Figueiredo and Menezes (1980), and nomenclature and classification were updated according to Eschmeyer's Catalog of Fishes (Van der Laan et al., 2023). To avoid ambiguity of some generic names, the following abbreviations were used: "*H.*" for

Hatschekia and “*Ho.*” for *Holocentrus*. Voucher specimens were deposited in the Coleção Carcinológica of the Museu de Zoologia da Universidade de São Paulo (acronym MZUSP), Brazil. Access to genetic heritage was registered in the Sistema Nacional de Gestão do Patrimônio Genético e do Conhecimento Tradicional Associado (acronym SisGen), under the number A03E910, according to the Brazilian Federal requirements.

Systematics

Order Siphonostomatoidea Burmeister, 1835

Family Hatschekiidae Kabata, 1979

Genus *Hatschekia* Poche, 1902

Hatschekia exigua Pearse, 1951

Type-host and type-locality: The Squirrelfish *Holocentrus adscensionis* (Osbeck) (Actinopterygii: Holocentridae); Bimini, Bahamas, Caribbean Sea (geographical coordinates not available) (Pearse, 1951).

Host and locality of present material: *Ho. adscensionis*; Mucuripe Bay (3°43'18"S, 38°28'51"W), Fortaleza, State of Ceará, Northeastern Brazil.

Site on host: Gills.

Prevalence and mean intensity: 100% (three fish infested out of three analyzed); mean of 6 copepods per infested fish (range 4–9).

Voucher specimens: 14 females (MZUSP-45252) and six males (MZUSP-45253). Four specimens were dissected and kept in the personal collection of the first author.

Redescription of adult female [based on 18 specimens; Figs. 1–3]. Body elongate (Fig. 1A), comprising distinct cephalothoracic head, short neck and long cylindrical trunk; external cuticular layer smooth. Body length 690–1138 (1004 ± 128.1), excluding caudal rami. Cephalothorax (Figs. 1A, B) forming octagonal to ovoid segment, expanded laterally forming blunt lobes and expanded posteriorly, shorter than wide $192\text{--}273$ (218 ± 23.5) \times $201\text{--}304$ (260 ± 30.2), representing about one-fourth of total body length; dorsal surface bearing chitinous supporting frame resembling key-hole shape, symmetrically with median longitudinal thick bar; each anterolateral corner with process anteriorly and two median transverse slender bars connecting to median thick one; posterior part deep (Fig. 1B). Trunk fusiform, longer than wide, $473\text{--}940$ (770 ± 129.7) \times $189\text{--}408$ (276 ± 60.9), lacking posterolateral lobes or processes, with

anterior narrow and short “neck”; expanding to maximum width at region slightly posterior to level of first leg level and gradually narrowing towards posterior end; internal tissues separated from cuticle and apparently sheathed with cuticular membrane. Urosome (Fig. 1C) comprising fused genital complex and abdomen, wider than long $32\text{--}60$ (48 ± 9.5) \times $45\text{--}75$ (60 ± 11.2). Caudal ramus (Fig. 1D) incompletely fused to urosome, longer than wide $24\text{--}35$ (30 ± 3.3) \times $12\text{--}18$ (15 ± 1.7), with five naked setae: four distal and one lateral. Egg sacs (Fig. 1E) shorter than trunk, uniseriate, with mean of eight eggs per sac, range from six to ten eggs per sac ($n = 5$).

Rostrum absent. Antennule (Fig. 2A) indistinctly five segmented, $118\text{--}171$ (152 ± 15.1) long; armature formula: 6, 3, 3, 1, 7 + 2 aesthetascs. Antenna (Fig. 2B) with three segments: proximal (coxa) unarmed, mid (basis) ornamented with circular pit and terminal claw short and robust. Antenna $169\text{--}228$ (204 ± 19.4) long; length of proximal, mid, and terminal segments $30\text{--}66$ (53 ± 10.6), $82\text{--}120$ (103 ± 12.2) and $40\text{--}57$ (48 ± 5.4), respectively. Parabasal papilla (Fig. 2B) blunt, thumb-like with annulated surface. Oral cone robust. Mandible (Fig. 2D) styli-form tapering posteriorly. Maxillule (Fig. 2E) bilobate, both lobes armed with two tapering setae of unequal sizes. Maxilla (Fig. 2C) with four segments: proximal unarmed; second rod-like with one basal seta and row of blunt, fine spinules on middle outer surface; third elongate with one distal seta. Terminal segment small, with one short seta and bifid claw. Maxilliped absent.

Legs 1 and 2 biramous, each joined by bar-like smooth intercoxal sclerite (Fig. 3C). Exopods represented by two incompletely fused segments and two-segmented endopods. All setae on rami naked and thick. Armature formula as follow:

	Protopod	Exopod	Endopod
Leg 1	1–1	1–0; 4	0–0; 5
Leg 2	1–0	1–0; 4	0–1; 4

Leg 1 (Fig. 3A) $92\text{--}114$ (103 ± 6.7) long, with coxa and basis fused forming large protopod and retaining trace of suture. Protopod $56\text{--}65$ (60 ± 2.9) long, with large inner seta and slender outer seta. Exopod longer than endopod, $35\text{--}51$ (43 ± 5.7) and $29\text{--}38$ (34 ± 3.2) long, respectively. Leg 2 (Fig. 3B)

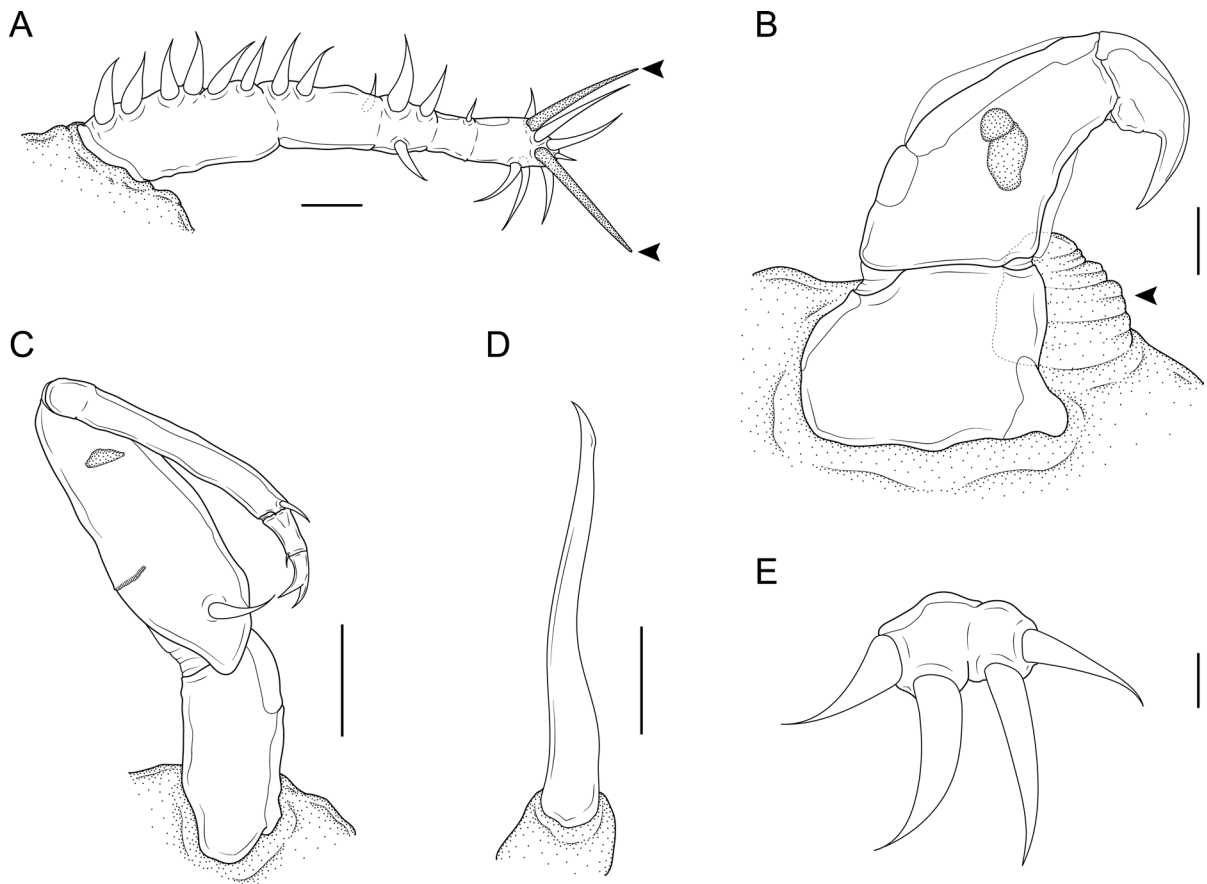


Fig. 2 *Hatschekia exigua* Pearse, 1951 (adult female). **A** antennule, dorsal, arrows pointing to aesthetascs; **B** antenna, ventral, arrow pointing to parabasal papilla; **C** maxilla, ventral;

D mandible, ventral; **E** maxillule, ventral. Scale bars: **A–B**= 20 μ m, **C**= 30 μ m, **D**= 5 μ m, **E**= 10 μ m

111–146 (131 ± 11.4) long, with coxa and basis fused to form protopod, retaining trace of suture. Protopod 71–84 (76 ± 4.6) long, with slender basal outer seta. Exopod longer than endopod, 40–65 (55 ± 8.1) and 37–60 (52 ± 5.5) long, respectively. Protopods and rami of legs 1 and 2 ornamented with rows of blunt and fine spinules on anterior surface.

Leg 3 (Fig. 3D) represented by two simple naked setae inserted in rounded papilla at middle of trunk. Leg 4 (Fig. 3E) represented by small naked seta located at posterior quarter of trunk with slightly swollen base.

Description of adult male [based on six specimens; Figs. 4, 5]. Body (Fig. 4A) comprising cephalothorax, short neck cylindrical trunk containing spermatophores, and with lateral seta representing leg

6 (Figs. 4A, C) on genital operculum. Body length 396–450 (432 ± 24.5), excluding caudal rami. Cephalothorax (Figs. 4A, B) longer than wide 140–153 (149 ± 6) \times 119–127 (123 ± 3.3), representing about one-third of total body length; dorsal surface bearing chitinous supporting frame (Fig. 4B), symmetrical with median longitudinal thick bar; form somewhat trapezoidal, divided anteriorly, with two rounded processes in anterolateral corners; first third formed by square process inserted in transverse bars, each with one blunt process at anterior margin, and spatula-shaped process deep in posterior margins; remaining part continuous, forming base of trapezium. Urosome (Fig. 4C) longer than wide 35–43 (40 ± 3.6) \times 35–39 (36 ± 1.7). Caudal ramus (Fig. 4C) incompletely fused to urosome, longer than wide 41–50 (46 ± 3.9)

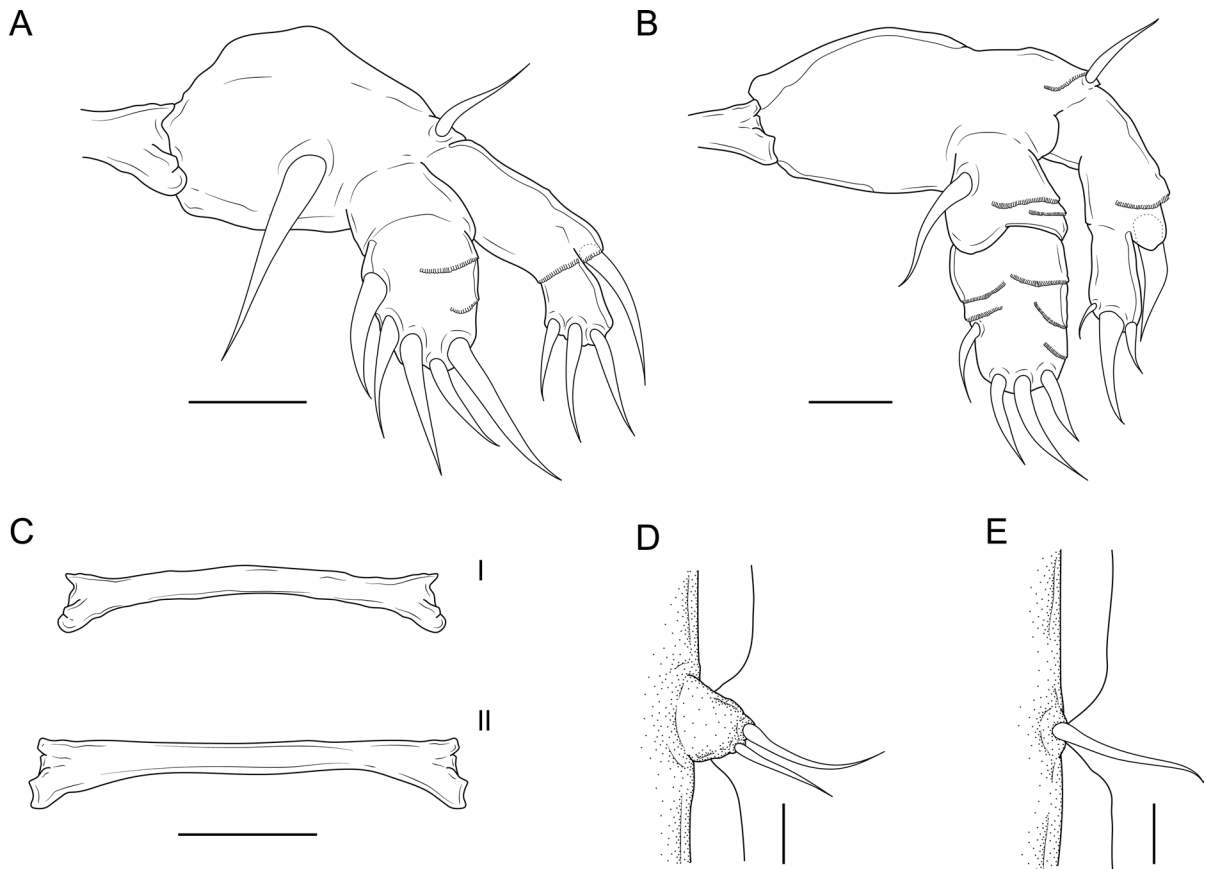


Fig. 3 *Hatschekia exigua* Pearse, 1951 (adult female). **A** leg 1, ventral; **B** leg 2, ventral; **C** intercoxal sclerites of legs 1 and 2, ventral; **D** leg 3, ventral; **E** leg 4, ventral. Scale bars: **A–B**= 20 μ m, **C**= 40 μ m, **D–E**= 20 μ m

\times 13–16 (15 ± 1.5), with six naked setae: five distal and one lateral.

Rostrum (Fig. 5A) formed by triangular plate with pair of blunt teeth anteriorly, and middle pointed process; pair of robust blunt rostral processes laterally to plate. Antennule (Fig. 5B) five segmented, 123–142 (132 ± 8.2) long; armature formula: 10, 4, 5, 1, 8 + 2 aesthetascs. Antenna (Fig. 5C) with three segments; last segment with proximolateral process and armed with two setae, terminal claw short and robust. Antenna 111–126 (121 ± 6.8) long; length of proximal, mid, and terminal segments 20–36 (29 ± 8.5), 52–58 (55 ± 2.5) and 29–45 (37 ± 6.6), respectively. Parabasal papilla (Fig. 5C) blunt, thumb-like, with annulated surface. Oral cone, mandible, maxillulae and maxilla as in female. Maxilliped absent.

Legs 1 and 2 biramous, joined by bar-like smooth intercoxal sclerite (Fig. 3D). Exopods represented by

two incompletely fused segments and two-segmented endopods. All setae on rami naked and thick. Armature formula as follow:

	Protopod	Exopod	Endopod
Leg 1	1-1	1-0; 3	0-0; 5
Leg 2	1-0	1-0; 4	0-1; 4

Leg 1 (Fig. 5D) 73–78 (75 ± 2.2) long, with coxa and basis fused to form protopod, retaining trace of suture. Protopod, exopod and endopod length 38–42 (41 ± 2), 31–40 (34 ± 4.1) and 23–26 (25 ± 1.4) long, respectively. Armature of leg 1 same as in female. Leg 2 (Fig. 5E) 82–94 (89 ± 5) long, with coxa and basis fused to form protopod, retaining trace of suture. Protopod, exopod and endopod 38–42 (41 ± 2), 31–40 (34 ± 4.1) and

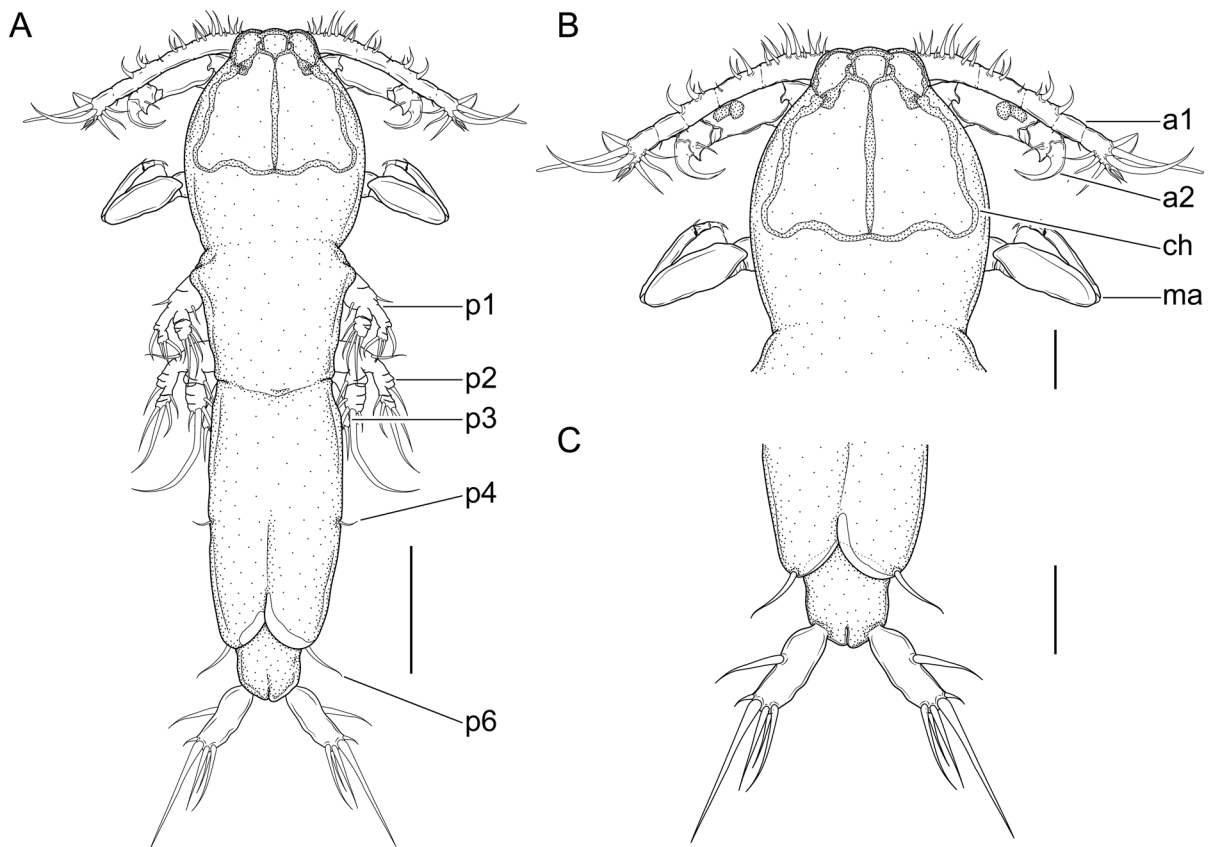


Fig. 4 *Hatschekia exigua* Pearse, 1951 (adult male). **A** habitus, dorsal, p1= leg 1, p2= leg 2, p3= leg 3, p4= leg 4, p6= leg 6; **B** detail of cephalothorax, dorsal, a1= antennule, a2=

antenna, ch= dorsal chitinous supporting frame, ma= maxilla; **C** posterior part of trunk, ventral. Scale bars: **A**= 50 μ m, **C–D**= 20 μ m

23–26 (25 ± 1.4) long, respectively. Protopods and rami of legs 1 and 2 ornamented with rows of blunt and fine spinules on anterior surface.

Leg 3 (Fig. 5G) represented by lateral lobe armed with five setae: two apical, two lateral and one near base. Leg 4 (Fig. 5H) represented by small naked seta located at posterior two thirds of trunk with slightly swollen base.

Discussion

The females analyzed in this work were identified as *H. exigua* because of the cephalothorax representing about one-fourth of total body length, with convex

lateral margins and expanded into rounded lateral lobes, the first exopod armed with one seta on the basal segment and three setae on the terminal segment, as well as with basal segment unarmed and terminal segment with five setae, the leg 3 reduced to two setae and the leg 4 reduced to single seta (Pearse, 1951). Moreover, considering that species of *Hatschekia* are highly host-specific at family level, the present specimens were collected from *Ho. adscensionis*, type host of *H. exigua*, and the fact that no other congener has been reported infesting Holocentridae fish, the specific identification of the present material is strongly supported (Kabata, 1979; Uyeno & Ali, 2013).

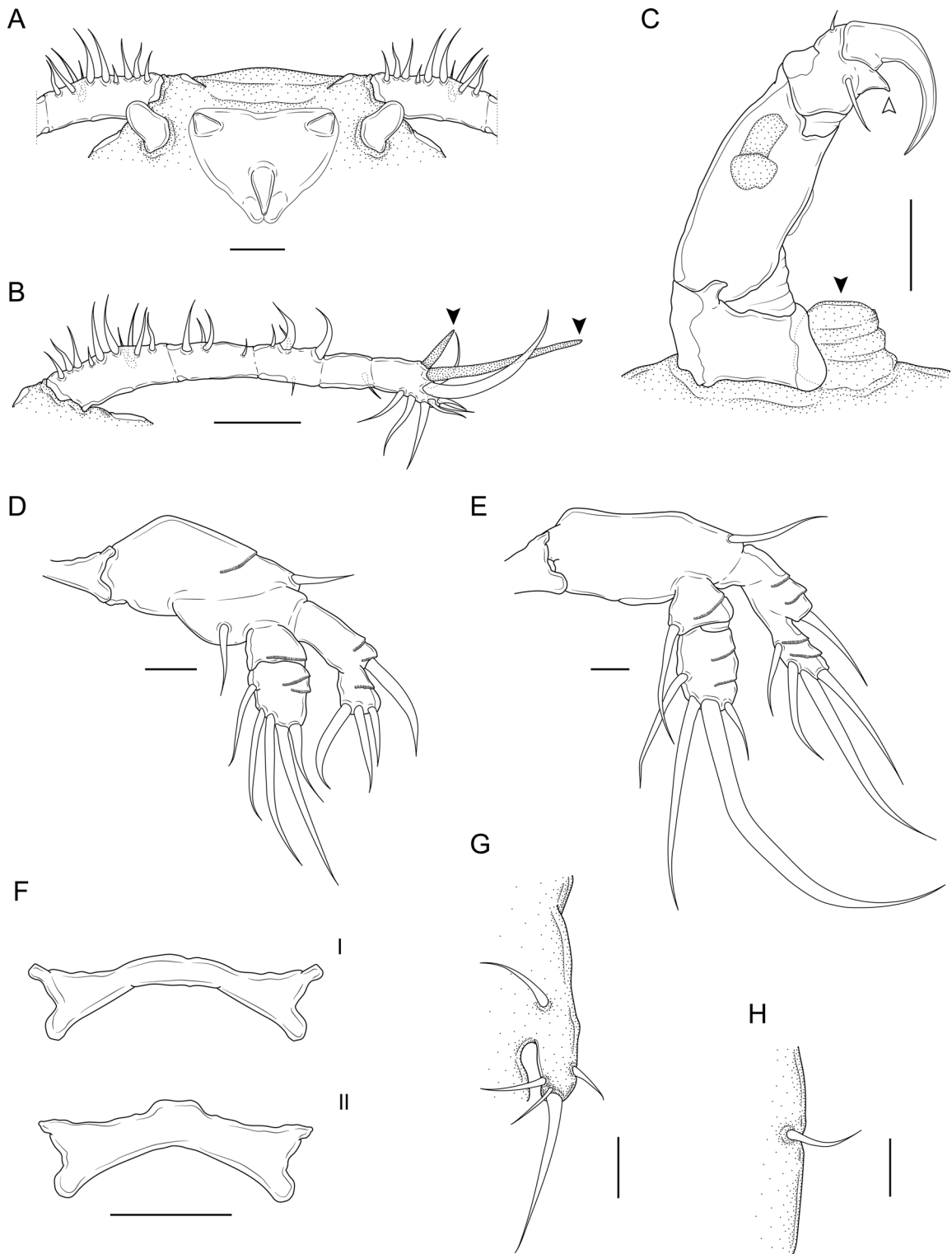


Fig. 5 *Hatschekia exigua* Pearse, 1951 (adult male). **A** detail of rostrum, ventral; **B** antennule, dorsal, arrows pointing to aesthetascs; **C** antenna, ventral, arrow pointing to parabasal

papilla; **D** leg 1, ventral; **E** leg 2, ventral; **F** intercoxal sclerites of legs 1 and 2, ventral; **G** leg 3, ventral; **H** leg 4, ventral. Scale bars: **A–B**= 30 μm , **C** = 20 μm , **C–H**= 10 μm

Examination of *H. exigua* type series (NMNH-88531) revealed the poor condition of material. It is represented by three copepods mounted on the same slide, in which the specimens are distorted and none of them show all the diagnostic features clearly. Based on the comparative analysis between the present specimens and those studied by Pearse (1951), it was possible to observe that the antennule of *H. exigua* was not 3-segmented, but at least 5-segmented. Further morphological comparisons were also compromised by the poorly detailed description of the type material (Pearse, 1951). As previously commented, Jones (1985) provided additional data and illustrations of the types of *H. exigua* but could not adequately redescribe the species. According to Jones (1985), the cephalothorax of *H. exigua* has lateral margins expanded into blunt lobes and the posterior margin of the trunk has small lobes on either side. Nevertheless, the cephalothorax of the present species has lateral blunt lobes, but not that prominent as illustrated by Jones (1985), and the posterior end of trunk is smooth (see Figs. 7L, M in Jones, 1985). These differences may be accounted by the mounting methodology and the poor conservation of the type material, which is distorted and compressed, compromising a detailed observation of body and appendages by Jones (1985). These new observations on the females, contribute to a better understanding *H. exigua* morphology, preventing further taxonomic confusion.

The small appendages of *Hatschekia* spp. are structures of great complexity, some apparently stable within conspecific individuals and others with intraspecific variability. These different levels of variability make several authors concern about which features should be used in the specific diagnosis and differentiation of species (Kabata, 1991; Uyeno & Nagasawa, 2009a; 2013; Paschoal et al., 2022). In this context, *H. exigua* presented intraspecific variability in the setation pattern of leg 2 when compared with the observations by Jones (1985), since the leg 2 was described with four and six apical setae on the terminal segments of exopod and endopod, respectively, while in the present study these segments were armed with three and four setae. The present specimens also differed from those described by Jones (1985) in the armature of the caudal ramus, while the author described at least four setae in this appendage, it was armed with six setae in the present material. Therefore, it is reasonable to consider that these differences

could be related to difficulties in the observation of the appendages or due to the loss of some setae by preservation issues (see paragraph above), mainly on the caudal ramus. However, the armature on the rami of legs 1 and 2 should be treated with caution because of its high intraspecific variability in the genus *Hatschekia* (Jones, 1985; Kabata, 1991; Uyeno & Nagasawa, 2013). In this sense, we reinforce that the general body plan and morphometric data should have preference in the specific diagnosis and differentiation, as well as the use of small appendages with great stability to support these analyses. At the time Pearse (1951) described *H. exigua*, there were only 37 valid congeneric species, and 75% (112 species) of the current richness of the genus was known posteriorly. Therefore, *Hatschekia* currently contains more than 140 valid species worldwide, 30 of which have a bar-like smooth intercoxal sclerite, and trunk lacking expansions, swellings, or processes as in the present females. Of these, only four have the same proportion of cephalothorax in relation to total body length as in *H. exigua*: *Hatschekia aulacocephalis* Izawa, 2016, *Hatschekia cirrhitchthysicola* Izawa, 2016, *Hatschekia euanus* Lee, Lee & Boxshall, 2013 and *Hatschekia hoplobrotulae* Izawa, 2015 (Lee et al., 2013; Izawa, 2015; 2016a). However, *H. exigua* can be distinguished from all these previously mentioned congeners because it has three setae on the last exopodal segment of leg 1 (vs. five setae in *H. aulacocephalis*; six in *H. cirrhitchthysicola*, *H. euanus* and *H. hoplobrotulae*) and five setae in the caudal rami (vs. six setae in the latter four species). *Hatschekia exigua* also differs from *H. aulacocephalis* based on the fourth segment of antennule lacking an enlarged ventral setae and the second segment of antenna unarmed (vs. enlarged seta present and second segment of antenna with proximoventral seta in the latter); from *H. cirrhitchthysicola* by rostral area lacking processes (vs. pointed processes in the latter); from *H. cirrhitchthysicola* and *H. euanus* by leg 3 reduced to two setae (vs. single seta in the latter two species); and from *H. hoplobrotulae* by leg 2 with three and four setae on the last exopodal and endopodal segment, respectively, and trunk 2.8 times longer than wide (vs. five setae on the last segment of both rami and trunk more than 3 times longer in the latter) (Lee et al., 2013; Izawa, 2015; 2016a).

Males of only 25 species of *Hatschekia* are currently known, indicating a scarce knowledge on such

representatives (Wilson, 1913; Pearse, 1947; Nuñez-Ruivo, 1954; Scram & Aspholm, 1997; Uyeno & Nagasawa 2009a; b; El-Rashidy & Boxshall, 2011; Uyeno & Nagasawa, 2012; Izawa, 2015; 2016a; b; c; 2018). The present males differ from all congeners because they possess leg 3 armed with five setae, feature that has never been reported in the genus. It should be mentioned that *H. exigua* shares the last exopodal segment of leg 1 armed with only three setae with two other congeners, *i.e.*, *Hatschekia curvata* Yamaguti & Yamasu, 1959 and *Hatschekia nemipteri* Izawa, 2016 (Izawa, 2016a). However, *H. exigua* can be clearly differentiated from these species by having five setae on the last endopodal segment of leg 1 (vs. two setae on the latter two species). It also can be distinguished from *H. curvata* by smooth intercoxal sclerites on legs 1 and 2 (vs. intercoxal sclerites with processes in the latter); and from *H. nemipteri* by a proximolateral process on the third segment of antenna (vs. absent in the latter) (Izawa, 2016a).

Since the establishment of *Hatschekia* in the 19th century, the differential diagnosis of species mainly relies on the morphology of females (Paschoal et al., 2022). Nevertheless, the morphological diversity of males, as presented here, was also important to support the specific diagnosis of *H. exigua*. Even though comparisons among males are less common in *Hatschekia*, it appears to be valuable for specific diagnosis and demonstrates that the body plan of males can be more diverse and useful than previously believed. Therefore, studies including males of *Hatschekia* should be better explored in future taxonomic studies, to contribute even more to the knowledge related to the genus and prevent systematics uncertainties.

Brazil has the longest coastline along the West Atlantic Ocean, extending for about 8,500 km, and supporting one of the richest ichthyofaunas of the world (Fernandez et al., 2019; Froese & Pauly, 2023). Despite being a hotspot of biodiversity for parasitic copepods, the regional diversity of hatschekiids appears to be underestimated due to uneven distribution of research effort, regarding the study of these organisms (Luque et al., 2013; Uyeno & Nagasawa, 2013; Paschoal et al., 2022). Such scarcity of studies becomes particularly evident in the State of Ceará, Northeastern Brazil, where only the fossil copepod *Kabatarina pattersoni* Cressey & Boxshall, 1989 † (Copepoda: Dichelesthiidae),

from *Cladocycclus gardneri* Agassiz † (Cladocyclidae), in the Araripe Plateau, has been reported (Cressey & Boxshall, 1989). In this sense, the present study represents not only the first report of *H. exigua* in Brazil but also the first hatschekiid copepod reported off the coast of Ceará. Therefore, the present results highlight the underestimated status of the richness associated with Hatschekiidae in Brazil, as well as with parasitic copepods in general, emphasizing the need for further investigations to improve the knowledge on their diversity and geographic distribution in the country.

Acknowledgments We wish to thank Dr. Chad Walter, Dr. Karen Osborn and Dr. Yolanda Villacampa from the Smithsonian National Museum of Natural History for kindly providing the type-material for examination. Also, thanks to the Instituto de Ciências do Mar – LABOMAR from the Universidade Federal do Ceará for providing facilities during fish analysis.

Author contributions JVC and CVF performed field collections; JVC and FP analyzed the copepods and prepared the illustrations; JVC and AJP prepared the main manuscript; FBP and FP supervised the study; all authors reviewed the manuscript & approved the final version.

Funding João V. Couto was supported by a student fellowship from Coordenação de Aperfeiçoamento de Pessoal do Ensino Superior (CAPES), Brazil. Felipe B. Pereira was supported by the Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG, Process no. APQ-01179-21). Fabiano Paschoal was supported by Fundação de Amparo à Pesquisa e ao Desenvolvimento Científico e Tecnológico do Maranhão (FAPEMA, Process no. 84516/2022), Brazil.

Data availability All data used analysis is fully available in the manuscript.

Declarations

Competing interests The authors declare no competing interests.

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval All applicable institutional, national and international guidelines for the care and use of animals were followed.

References

Boxshall, G. A., Halsey, S. H. (2004). *An introduction to copepod diversity*. The Ray Society.

- Bush, A. O., Lafferty, K. D., Lotz, J. M., & Shostak, A. W. (1997). Parasitology Meets Ecology on Its Own Terms: Margolis et al. Revisited. *The Journal of Parasitology*, 83(4), 575. <https://doi.org/10.2307/3284227>
- Cantatore, D. M. P., Braicovich, P. E., Alarcos, A. J., Lanfranchi, A. L., Rossin, M. A., Vales, D. G., & Timi, J. T. (2012). New records of parasitic copepods (Crustacea, Copepoda) from marine fishes in the Argentinean Sea. *Acta Parasitologica*, 57(1), 83–89. <https://doi.org/10.2478/s11686-012-0003-z>
- Cressey, R., & Boxshall, G. (1989). *Kabatarina pattersoni*, a Fossil Parasitic Copepod (Dichelethiidae) from a Lower Cretaceous Fish. *Micropaleontology*, 35(2), 150. <https://doi.org/10.2307/1485466>
- El-Rashidy, H. H., & Boxshall, G. A. (2011). Two new species of parasitic copepods (Crustacea) on two immigrant rabbitfishes (Family Siganidae) from the Red Sea. *Systematic Parasitology*, 79(3), 175–193. <https://doi.org/10.1007/s11230-011-9298-7>
- Fernandez, G. B., da Rocha, T. B., Barboza, E. G., Dillenburg, S. R., da Camara Rosa, M. L. C., Angulo, R. J., de Souza, M. C., de Oliveira, L. H. S., & Dominguez, J. M. L. (2019). Natural Landscapes Along Brazilian Coastline. In: Salgado, A., Santos, L., Paisani, J. (eds), *The Physical Geography of Brazil: environment, vegetation, and landscape* (1st ed., pp. 199–218). Springer. https://doi.org/10.1007/978-3-030-04333-9_10
- Figueiredo, J. L., & Menezes, N. A. (1980). *Manual de Peixes Marinhos do Sudeste do Brasil III. Teleostei (2)*. Museu de Zoologia, Universidade de São Paulo.
- Froese, R., & Pauly, D. (2023). *FishBase*. World Wide Web electronic publication. Retrieved October 05, 2023, from <http://www.fishbase.org>
- Humes, A. G., & Gooding, R. U. (1964). A Method for Studying the External Anatomy of Copepods. *Crustaceana*, 6(3), 238–240. <https://doi.org/10.1163/156854064x00650>
- Huys, R., & Boxshall, G. A. (1991). *Copepod evolution*. The Ray Society.
- Izawa, K. (2015). Some new and known species of Hatschekiidae (Copepoda, Siphonostomatoida) parasitic on Japanese actinopterygian fishes belonging to Ophidiiformes, Beryciformes and Scorpaeniformes, with discussion on the female genital system and the insemination mode in Siphonostomatoida*. *Crustaceana*, 88(3), 359–384. <https://doi.org/10.1163/15685403-00003409>
- Izawa, K. (2016a). Some new and known species of *Hatschekia* Poche, 1902 (Copepoda, Siphonostomatoida, Hatschekiidae) parasitic on the branchial lamellae of Japanese actinopterygian fishes belonging to Perciformes, with revision of the known species of the genus. *Crustaceana*, 89(2), 209–238. <https://doi.org/10.1163/15685403-00003503>
- Izawa, K. (2016b). Resurrection and redescription of *Hatschekia cylindrica* Shiino, 1957 (Copepoda, Siphonostomatoida, Hatschekiidae), with description of its developmental stages. *Crustaceana*, 89(3), 343–357. <https://doi.org/10.1163/15685403-00003527>
- Izawa, K. (2016c). Some new and known species of *Hatschekia* Poche, 1902 (Copepoda, Siphonostomatoida, Hatschekiidae) parasitic on the branchial lamellae of Japanese actinopterygian fishes belonging to Perciformes (2). *Crustaceana*, 89(6–7), 819–850. <https://doi.org/10.1163/15685403-00003549>
- Izawa, K. (2018). Redescription of five known species of *Hatschekia* Poche, 1902 (Copepoda, Siphonostomatoida, Hatschekiidae) parasitic on Japanese tetraodontiform fishes, with discussion on sexually dimorphic features in the genus. *Crustaceana*, 91(9), 1107–1132. <https://doi.org/10.1163/15685403-00003819>
- Jones, J. B. (1985). A revision of *Hatschekia* Poche, 1902 (Copepoda: Hatschekiidae), parasitic on marine fishes. *New Zealand Journal of Zoology*, 12(2), 213–271. <https://doi.org/10.1080/03014223.1985.10428280>
- Kabata, Z. (1979). *Parasitic Copepoda of British Fishes*. The Ray Society.
- Kabata, Z. (1991). Copepoda parasitic on Australian fishes, XIII: family Hatschekiidae. *Journal of Natural History*, 25(1), 91–121. <https://doi.org/10.1080/00222939100770081>
- Lee, S., Lee, W., & Boxshall, G. (2013). Seven new species of *Hatschekia* Poche, 1902 (Copepoda: Siphonostomatoida: Hatschekiidae) parasitic on fishes of New Caledonia, and a redescription of *H. cadenati* Nunes-Ruivo, 1954. *Zoosystema*, 35(3), 377–413. <https://doi.org/10.5252/z2013n3a3>
- Luque, J. L., & Tavares, L. E. R. (2007). Checklist of Copepoda associated with fishes from Brazil. *Zootaxa*, 1579(1), 1–39. <https://doi.org/10.11646/zootaxa.1579.1.1>
- Luque, J. L., Vieira, F. M., Takemoto, R. M., Pavanelli, G. C., & Eiras, J. C. (2013). Checklist of Crustacea parasitizing fishes from Brazil. *Check List*, 9(6), 1449. <https://doi.org/10.15560/9.6.1449>
- Núñez-Ruivo, L. P. (1954). Parasites de poissons de mer ouest-Africains récoltés par MJ Cadenat. III. Copépodes (2 note). Genres *Prohatschekia* n. gen. et *Hatschekia* Poche. *Bulletin de l'Institut Français d'Afrique Noire*, 16, 479–505.
- Paschoal, F., Couto, J. V., Pereira, F. B., & Luque, J. L. (2022). A New Species of Hatschekiid Copepod (Crustacea: Hatschekiidae) Parasitic on the Porkfish *Anisotremus virginicus* (Linnaeus, 1758) (Actinopterygii: Haemulidae), with Notes on Previously Known Species of *Hatschekia* Poche, 1902 Collected from Actinopterygians off Brazil. *Acta Parasitologica*, 67(3), 1126–1135. <https://doi.org/10.1007/s11686-022-00551-3>
- Pearse, A. S. (1947). Parasitic copepods from Beaufort, North Carolina. *Journal of the Elisha Mitchell Scientific Society*, 63(1), 1–16.
- Pearse, A. S. (1951). Parasitic Crustacea from Bimini, Bahamas. *Proceedings of the United States National Museum*, 101(3280), 341–372. <https://doi.org/10.5479/si.00963801.101-3280.341>
- Uyeno, D., & Ali, A. H. (2013). Parasitic copepods from two species of commercial fishes collected off Iraq, with description of *Hatschekia shari* n. sp. *Systematic Parasitology*, 86(3), 301–312. <https://doi.org/10.1007/s11230-013-9446-3>
- Uyeno, D., & Nagasawa, K. (2009a). Redescription of four species of *Hatschekia* (Copepoda: Siphonostomatoida: Hatschekiidae) parasitic on tetraodontiform fishes from Japan. *Zootaxa*, 2110(1), 1–21. <https://doi.org/10.11646/zootaxa.2110.1.1>

- Uyeno, D., & Nagasawa, K. (2009b). Three new species of *Hatschekia* Poche, 1902 (Copepoda: Siphonostomatoida: Hatschekiidae) parasitic on *Abalistes filamentosus* (Pisces: Tetraodontiformes: Balistidae) from off Okinawa, Japan. *Systematic Parasitology*, 74(3), 225–237. <https://doi.org/10.1007/s11230-009-9208-4>
- Uyeno, D., & Nagasawa, K. (2012). Two new species of the copepod *Hatschekia* Poche, 1902 (Siphonostomatoida: Hatschekiidae) from angelfishes (Pisces: Perciformes: Pomacanthidae) collected during the KUMEJIMA 2009 Expedition. *Zootaxa*, 3367(1), 49. <https://doi.org/10.11646/zootaxa.3367.1.5>
- Uyeno, D., & Nagasawa, K. (2013). The genus *Hatschekia* (Copepoda: Hatschekiidae) from pufferfishes (Tetraodontiformes: Tetraodontidae) off the Ryukyu Islands, Japan, with descriptions of four new species and a redescription of *H. pholas*. *Folia Parasitologica*, 60(1), 61–74. <https://doi.org/10.14411/fp.2013.008>
- Van der Laan, R., Fricke, R., & Eschmeyer, W. N. (2023). *Classification. Eschmeyer's Catalog of Fishes*. Retrieved October 05, 2023, from www.calacademy.org/scientists/catalog-of-fishes-classification
- Walter, T. C., & Boxshall, G. (2023). *Hatschekia* Poche, 1902. *World of Copepods database*. Retrieved October 05, 2023, from <http://marinespecies.org/copepoda/aphia.php?p=taxdetails&id=135591>.
- Wilson, C. B. (1913). Crustacean parasites of West Indian fishes and land crabs, with descriptions of new genera and species. *Proceedings of the United States National Museum*, 44(1950), 189–277. <https://doi.org/10.5479/si.00963801.44-1950.189>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.