Evaluation of clinical and laboratory parameters used in the identification of index cases for genetic screening of familial hypercholesterolemia in Brazil

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ABSTRACT

Background and aims: There is controversy on the accuracy of different diagnostic criteria for familial hypercholesterolemia (FH). The aim of this study is to assess the performance of different clinical criteria used to identify individuals for FH genetic cascade screening in Brazil.

Methods: All index cases (IC) registered in the Hipercol Brasil program between 2011 and 2016 were analyzed. Inclusion criteria were age ≥ 18 years and elevated LDL-cholesterol (LDL-C) levels, with a conclusive result in the genetic test, whether positive or negative. Initially, we tested the multivariable association between clinical and laboratory markers and the presence of an FH causing mutation. Then, we analyzed sensitivity, specificity, positive and negative predictive values for the LDL-C quartile distribution, LDL-C as a continuous variable, as well as the performance measures for the Dutch Lipid Clinic Network (DLCN) score to identify a mutation.

Results: Overall, 753 ICs were included and an FH causing mutation was found in 34% (n = 257) of the subjects. After multivariable analysis, LDL-C as a continuous variable, tendon xanthomas and corneal arcus were independently associated with the presence of FH mutations. LDL-C values ≥ 230 mg/dL (5.9 mmol/L) had the best tradeoff between sensitivity and specificity to diagnose a mutation. The DLCN score presented a better performance than LDL-C to identify a mutation, area under the ROC curve were 0.744 (95% CI: 0.704–0.784) and 0.730 (95% CI: 0.687–0.774), respectively, p = 0.014.

Conclusions: In our population, LDL-C ≥ 230 mg/dL is a feasible criterion to indicate ICs to genetic testing.

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1. Introduction

Familial hypercholesterolemia (FH) is an autosomal dominant genetic disorder caused by variants that alter LDL-cholesterol (LDL-C) catabolism, leading to elevated blood cholesterol levels and consequently increasing the risk of atherosclerotic cardiovascular disease (ASCVD) development. In addition, due to the long-term exposure to high LDL-C, individuals with FH may present clinical signs that indicate the presence of tendon xanthomas, corneal arcus (when present in individuals aged ≤ 45 years), and xanthelasmas [1].

Currently, there are about 1600 mutations described as causing FH, with 95% of them occurring in the gene coding for the LDL receptor (LDLR). The remaining 5% affect genes coding for apolipoprotein B (APOB) and proprotein convertase subtilisin/kexin type 9 (PCSK9) genes.
Heterozygous FH is estimated to affect up to 670,000 Brazilians [5]. The diagnosis and treatment of FH could change the prevalence scenario of ASCVD at early ages since LDL-C lowering treatment has been shown to reduce cardiovascular disease incidence and mortality [6,7].

Diagnosis of FH individuals may be cost-effectively performed through genetic cascade screening programs [7]. Cascade screening methods consist in the identification of an index case (IC), which, because of its high a priori probability of harboring a causal mutation, is then referred to genetic test and, in case of a positive result (i.e. an identified causal genetic mutation), all first-degree relatives are screened for the same mutation. Despite being the most cost-effective method, it depends on the assertive identification of an IC, which is itself based on clinical criteria [8]. In the literature, the most adopted clinical classifiers for FH are from the Simon Broome Register Group [9] and the Dutch Lipid Clinic Network (DLCN) [10]; however, there is no international consensus on which are the best clinical predictors to refer patients for genetic test. This is particularly relevant if one considers the different settings from which ICs may be referred (direct-consumer, primary care, secondary care) and the different availability of detailed laboratory and clinical information on them. Therefore, population-specific adjustments are frequently observed in the literature [11,12].

The accuracy of specific clinical or biochemical parameters may change from one setting to another and the overall cost-effectiveness of cascade screening programs depends on the balance between sensitivity and specificity testing for the IC. Therefore, in this study, we assessed the accuracy of different FH diagnostic clinical criteria in the Hipercol Brasil [5], a state of the art genetic cascade-screening program.

2. Materials and methods

This study was performed with subjects registered in an FH genetic cascade screening program in Brazil. Participants were recruited by doctors from the Heart Institute (InCor), University of São Paulo Medical School or from partner institutions. In addition, individuals, who contacted the program via website, were also selected by trained health professionals from the program. After inclusion criteria were met, participants were referred to genetic testing.

The program is conducted at the Laboratory of Genetics and Molecular Cardiology at the Heart Institute (InCor/HCFMUSP), University of São Paulo Medical School Hospital, São Paulo, Brazil. The study was approved by the Institutional Ethics Committee (CAPesq number 3757/12/013), and all subjects signed an informed consent form.

2.1. Study population and inclusion criteria

We included in this analysis all IC individuals registered in Hipercol Brasil between 2011 and 2016. The inclusion criteria for this study were: age ≥18 years and a conclusive result of the genetic test, whether positive or negative. The only inclusion criterion for being registered at Hipercol Brasil was to have previous documentation of an LDL-C ≥210 mg/dL. However, some individuals with LDL-C values ≤210 mg/dL were also included in the program during the selected period, when referred by local physicians due to other overwhelming clinical characteristics associated with FH.

2.2. Clinical and laboratory characteristics

All relevant clinical information was collected through a standardized questionnaire and physical exams followed by the collection of biological material as previously described [5]. Subjects were examined for the presence of xanthomas, xanthelasma or corneal arcus. Weight (kg) and height (m) were determined and the body mass index (BMI in kg/m²) was calculated. Laboratory exams were obtained from medical records or from previous exams brought by the participants. Values of fasting total cholesterol (TC), LDL-C, HDL-C, and triglycerides (TG) were collected. LDL-C values used were those available at baseline evaluation. When the subject was on statin use, we asked for the highest documented value before statin initiation. In case we could not retrieve that piece of information, the value obtained under statin treatment was used to calculate the scores. The DLNC score was calculated using information available at the baseline visit and, therefore, the presence of an FH causing mutation was not considered for diagnosis.

2.3. Genetic testing

Subjects were tested for six FH-related genes: LDLR, APOB, PCSK9, LDLRAP1, LIPA and APOE using state of the art molecular techniques. Target regions were considered as coding exons plus 10 bp of introns up- and downstream. The promoter region of LDLR was also screened. Templates were prepared on Ion One Touch System and sequenced in Ion Torrent PGM® platform, with 32 samples per run in a 316v2 Ion Chip. Raw FASTQ files were imported into CLC Genomics Workbench 9.5 (Qiagen) and analyzed using a custom pipeline. Minimum quality requirements for variant call were: base quality of PhredQ ≥20; target-region coverage ≥10x; frequency of variant allele ≥20% and bidirectional presence of the variant allele. After minor allele frequency filtering (MAF < 0.002) with control populations (NHBLI-ESP6500, ExAC and 1000Genomes), all potentially causal variants were consulted for previous description in ClinVar, Human Genome Mutation Database (HGMD® Professional - Qiagen), British Heart Foundation and Jojo Genetics databases. Pathogenicity attribution was performed according to the American College of Medical Genetics (ACMG) guideline [13].

For previously undescribed variants, functional impact prediction was performed with SIFT, PROVEAN and PolyPhen-2 and variants without previous description were considered as potentially pathogenic when pointed as damaging in at least two algorithms and if MAF ≤ 0.002. Individuals with negative results were also screened for large insertions and deletions via MLPA (MRC-Holland).

2.4. Statistical analysis

Initially, a descriptive analysis of the variables was carried out and results are presented according to the IC’s genetic results; mutation positive (M+) for those in whom a causative genetic variant was identified, and mutation negative (M−) for those without observed causative variants. For continuous variables, the mean and standard deviations were calculated. Categorical variables were described as frequencies. The differences between frequencies were compared using the Chi-square test. The differences between means were compared with Student t or analysis of variance (ANOVA) tests, if necessary. Logistic regression was used to test the independent association of clinical and laboratory variables with the presence of a mutation. The performance (sensitivity, specificity, positive and negative predictive values PPV and NPV, respectively) of different criteria to diagnose the presence of an FH causing mutation was tested as follows: first, the categorical distribution of LDL-C (25th, 50th and 75th percentiles) within the study population; second LDL-C levels as continuous variables and the best tradeoff between sensitivity and specificity was
determined by calculating the area under the receiver operation curve (ROC); third the DLCN score; we compared the discriminative power of different parameters to select ICs for genetic test. As expected, the higher the DLCN level, the greater the specificity and the lower the sensitivity for the presence of an FH causing mutation. DLCN levels >230 mg/dL (5.9 mmol/L) that coincided with the 50th percentile of the DLCN distribution of the IC population, had the best tradeoff between sensitivity and specificity to diagnose a mutation as shown in Supplemental Table 1. The DLCN score presented a better discrimination than LDL-C levels >230 mg/dL (5.9 mmol/L) to identify an FH causing mutation, area under the ROC curve respectively of 0.744 (95% CI: 0.704–0.784) and 0.730 (95% CI: 0.687–0.774), p = 0.01 (Supplemental Fig. 1 and Supplemental Table 1). The DLCN ≥ 230 mg/dL cutoff was identified as the best value in the age groups (≤40; 41–51; 52–59 year old), except for the elderly group (≥60 years), in which the best

3. Results

Seven hundred and fifty-three ICs were included in the study. Tables 1 and 2 describe clinical and laboratorial characteristics of included individuals. Overall, 34.1% (n = 257) of screened individuals had a positive mutation identified through genetic screening (M+). M+ individuals were significantly younger than M− ICs. The presence of characteristic clinical signs of FH such as xanthomas, xanthelasmas and corneal arcus was greater in M+ individuals. There were no differences between the groups regarding the presence of previous ASCVD or lipid lowering therapy. Overall, 40.9% of the individuals that were classified as “definitive” diagnostic in the DLCN clinical score had a causal variant identified (Table 1).

Table 2 shows baseline and on-treatment plasma lipids of the studied subjects. The average baseline and on-treatment LDL-C and TC values were higher in M+ individuals. On the other hand, M− individuals presented higher values for both baseline and on-treatment TG.

Table 3 shows that after multivariable logistic regression analysis, the presence of tendon xanthomas, corneal arcus and baseline LDL-C values was independently associated with the presence of an FH causative mutation. No independent association was found with family history of early ASCVD or elevated LDL-C levels. Table 4 and Fig. 1 show values for sensitivity, specificity, PPV and NPV of different parameters to select ICs for genetic test. As expected, the higher the LDL-C levels, the greater the specificity and the lower the sensitivity for the presence of an FH causing mutation. LDL-C levels >230 mg/dL (5.9 mmol/L), that coincided with the 50th percentile of the LDL-C distribution of the IC population, had the best tradeoff between sensitivity and specificity to diagnose a mutation as shown in Supplemental Table 1. The DLCN score presented a better discrimination than LDL-C ≥ 230 mg/dL (5.9 mmol/L) to identify an FH causing mutation, area under the ROC curve respectively of 0.744 (95% CI: 0.704–0.784) and 0.730 (95% CI: 0.687–0.774), p = 0.01 (Supplemental Fig. 1 and Supplemental Table 1). The LDL-C ≥ 230 mg/dL cutoff was identified as the best value in the age groups (≤40; 41–51; 52–59 year old), except for the elderly group (≥60 years), in which the best

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**Table 1**

<table>
<thead>
<tr>
<th>Mutation + (n = 257)</th>
<th>Mutation − (n = 496)</th>
<th>p value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Males (%)</td>
<td>36.7</td>
<td>216</td>
</tr>
<tr>
<td>Female (%)</td>
<td>36.7</td>
<td>216</td>
</tr>
<tr>
<td>Hypertension (%)</td>
<td>63.3</td>
<td>314</td>
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<tr>
<td>Diabetes (%)</td>
<td>12.7</td>
<td>63</td>
</tr>
<tr>
<td>Early coronary artery disease (%)</td>
<td>31.0</td>
<td>154</td>
</tr>
<tr>
<td>Acute myocardial infarction (%)</td>
<td>18.3</td>
<td>91</td>
</tr>
<tr>
<td>Family history of increased LDL-C levels (%)</td>
<td>48.0</td>
<td>238</td>
</tr>
<tr>
<td>Family history of early coronary artery disease (%)</td>
<td>30.4</td>
<td>78</td>
</tr>
<tr>
<td>Current pharmacological treatment (%)</td>
<td>52.0</td>
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<tr>
<td>BMI (kg/m²)</td>
<td>27.5</td>
<td>221</td>
</tr>
<tr>
<td>Smoking (%)</td>
<td>14.7</td>
<td>73</td>
</tr>
<tr>
<td>Tendon xanthoma (%)</td>
<td>10.1</td>
<td>50</td>
</tr>
<tr>
<td>Xanthelasmas (%)</td>
<td>12.5</td>
<td>32</td>
</tr>
<tr>
<td>Corneal arcus (%)</td>
<td>13.5</td>
<td>67</td>
</tr>
<tr>
<td>DLCN score (%)</td>
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<td></td>
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<tr>
<td>Definitive</td>
<td>10.1</td>
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</tr>
<tr>
<td>Probable</td>
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<tr>
<td>Possible</td>
<td>42.7</td>
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</tr>
<tr>
<td>Unlikely</td>
<td>12.1</td>
<td>60</td>
</tr>
</tbody>
</table>

* p < 0.05.

**Table 2**

<table>
<thead>
<tr>
<th>Mutation + (n = 257)</th>
<th>Mutation − (n = 496)</th>
<th>p value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baseline TC (mg/dL)</td>
<td>388 ± 84</td>
<td>134</td>
</tr>
<tr>
<td>On treatment TC (mg/dL)</td>
<td>340 ± 102</td>
<td>102</td>
</tr>
<tr>
<td>Baseline LDL-C (mg/dL)</td>
<td>307 ± 79</td>
<td>127</td>
</tr>
<tr>
<td>On treatment LDL-C (mg/dL)</td>
<td>265 ± 105</td>
<td>100</td>
</tr>
<tr>
<td>Baseline HDL-C (mg/dL)</td>
<td>48 ± 15</td>
<td>125</td>
</tr>
<tr>
<td>On treatment HDL-C (mg/dL)</td>
<td>46 ± 13</td>
<td>100</td>
</tr>
<tr>
<td>Baseline TG (mg/dL)</td>
<td>141 ± 63</td>
<td>127</td>
</tr>
<tr>
<td>On treatment TG (mg/dL)</td>
<td>144 ± 72</td>
<td>100</td>
</tr>
</tbody>
</table>

* p < 0.05.

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260

In the age group analysis, using LDL-C value as a sole parameter, the cutoff in the elderly group (the FH mutation presence in hypercholesterolemic individuals. However, as the aim of the program is to identify positives compared to other age groups. This result is understandable since cholesterol tends to increase with age, and older individuals might present high levels of LDL-C when entering the program, indicating an inadequate and ineffective therapy, regardless of a known genetic background. Although genetic testing is important for the cascade screening and for diagnostic confirmation, the adopted treatment and clinical management are based on the phenotype features of each individual, which may be highly variable [17–19].

The cascade necessarily begins with a genetic-positive IC, and the identification of a mutation is strongly associated with the clinical diagnosis [12]. Nonetheless, criteria to establish the most cost-effective cutoff for the selection of to-be-screened ICs are not widely agreed upon. Consequently, different programs worldwide have made adjustments to obtain a more effective screening performance for their populations [12,20]. Considering that defining the best cutoff of a diagnostic test procedure involves trade-offs between sensitivity and specificity, and that molecular testing is not able to capture the entire complexity of the clinical diagnosis (it is rather a tool for cascade screening), we aimed at analyzing the overall accuracy of the tested procedures in the high number of individuals undergoing a still costly procedure is the determinant to consider the strategy. In addition, one must consider the ease by which healthcare providers and even individuals from the general population would be able to self-identify or identify a family member as candidates for screening.

The DLCN score is recommended in different guidelines for FH diagnosis [4,11,21]. However, this score is strongly dependent on a reliable knowledge of family history, laboratory values in relatives, and the presence of clinical signs of cholesterol deposits in the skin and tendons. Indeed, in our study, cholesterol deposit signs were and tendons. Indeed, in our study, cholesterol deposit signs were considered the strategy. In addition, one must consider the ease by which healthcare providers and even individuals from the general population would be able to self-identify or identify a family member as candidates for screening.

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independently associated with mutation discovery, but they are becoming harder to find nowadays due to the use of lipid lowering therapy for hypercholesterolemia in the absence of an FH diagnosis [19]. In addition, some family information is frequently hard to obtain and usually inaccurate. Indeed, in our study, no independent association was found between family history of early ASCVD or elevated cholesterol levels with mutation finding. Therefore, we believe that the use of an LDL-C threshold could be an alternative to the DLCN score to indicate mutation screening, since previous evidence has suggested that very high blood cholesterol levels are associated with a greater chance of encountering a monogenic cause for the FH phenotype [22].

When comparing the discriminatory performance of DLNC scores with LDL-C values as a criterion for mutation identification, the former presented a better discriminatory performance. However, overall, the proposed LDL-C cutoff value of 230 mg/dL (5.9 mmol/L) was a reasonable alternative, with good discrimination capacity.

This study has limitations: first our cohort did not include individuals under 18 years and second, around 80% of the studied subjects were under statin treatment at the time of the evaluation, and even considering that we used the highest available LDL-C, with or without lipid lowering medications, we can’t totally rule out an effect of statin treatment on our results. However, this problem is common to the contemporary FH cohorts, where hypercholesterolemia is diagnosed before FH [23]. Finally, the cost-effectiveness of our findings must be tested in a greater number of individuals.

In conclusion, our data suggest that both the DLCN score and a sole criterion of an LDL-C \( \geq 230 \) mg/dL (5.9 mmol/L) seem to be adequate for molecular FH screening in Brazilian individuals. However, using only LDL \( \geq 230 \) mg/dL is enough to indicate an IC to genetic test, when obtaining some reliable information on all the clinical variables required in DLNC guidelines is often difficult.

**Conflict of interest**

RDS refers receiving honoraria for consulting or speaker activities in the last 3 years from Amgen, Aegerion, Astra Zeneca, Akcea, Boehringer-Ingelheim, Biolab, Cerenis, Genzyme, Kowa, Eli-Lilly, Merck, Pfizer, Sanofi/Regeneron, Torrent, Procaps and Unilever. All the other authors have no conflict of interest. We wish to confirm that there are no other known conflicts of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome.

**Financial support**

The funding of Sociedade Hospital Samaritano and Ministério da Saúde (PROADI-SUS; SIPAR: 25000.180.672/2011-81) and FAPESP (grant no 2013/17368-0) are gratefully acknowledged.

**Acknowledgments**

We thank the patients who participated in the cohort study, and the technical assistance of the Laboratory of Genetics and Molecular Cardiology group, Heart Institute (InCor) University of Sao Paulo Medical School Hospital.
Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.atherosclerosis.2017.06.917.

References


