



UNIVERSIDADE FEDERAL DO CEARÁ
PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM BIOTECNOLOGIA (RENORBIO-UFC)
BIOTECNOLOGIA EM SAÚDE

EDUARDO HENRIQUE CUNHA NEVES FILHO

**CARACTERIZAÇÃO MOLECULAR E AVALIAÇÃO PROGNÓSTICA DE
BIOMARCADORES EM LINFOMA DIFUSO DE GRANDES CÉLULAS B**

FORTALEZA

2023

EDUARDO HENRIQUE CUNHA NEVES FILHO

CARACTERIZAÇÃO MOLECULAR E AVALIAÇÃO PROGNÓSTICA DE
BIOMARCADORES EM LINFOMA DIFUSO DE GRANDES CÉLULAS B

Tese apresentada ao Programa de Pós-Graduação em Biotecnologia - RENORBIO da Universidade Federal do Ceará para obtenção do título de Doutor em Biotecnologia. Área de concentração: Biotecnologia em Saúde.

Orientador: Silvia Helena Barem Rabenhorst.

FORTALEZA

2023

Dados Internacionais de Catalogação na Publicação
Universidade Federal do Ceará
Sistema de Bibliotecas

Gerada automaticamente pelo módulo Catalog, mediante os dados fornecidos pelo(a) autor(a)

N423c Neves Filho, Eduardo Henrique Cunha.

Caracterização molecular e avaliação prognóstica de biomarcadores em Linfoma Difuso de Grandes Células B / Eduardo Henrique Cunha Neves Filho. – 2023.

85 f. : il. color.

Tese (doutorado) – Universidade Federal do Ceará, Pró-Reitoria de Pesquisa e Pós-Graduação, Programa de Pós-Graduação em Biotecnologia (Rede Nordeste de Biotecnologia), Fortaleza, 2023.

Orientação: Profa. Dra. Sílvia Helena Barem Rabenhorst.

1. Linfoma. 2. Biomarcadores. 3. MicroRNA. I. Título.

CDD 660.6

EDUARDO HENRIQUE CUNHA NEVES FILHO

CARACTERIZAÇÃO MOLECULAR E AVALIAÇÃO PROGNÓSTICA DE BIOMAR-
CADORES EM LINFOMA DIFUSO DE GRANDES CÉLULAS B

Tese apresentada ao Programa de Pós-Graduação em Biotecnologia - RENORBIO da Universidade Federal do Ceará para obtenção do título de Doutor em Biotecnologia. Área de concentração: Biotecnologia em Saúde.

Aprovada em: 17/04/2023.

BANCA EXAMINADORA

Profa Dra Silvia Helena Barem Rabenhorst (Orientadora)
Universidade Federal do Ceará (UFC)

Profa Dra Adriana Camargo Ferrasi
Universidade Estadual Paulista (UNESP)

Prof Dr Fábio Rocha Fernandes Távora
Universidade Federal do Ceará (UFC)

Prof Dr Fernando Barroso Duarte
Universidade Federal do Ceará (UFC)

Profa Dra Maria do Perpétuo Socorro Saldanha da Cunha
Instituto do Câncer do Ceará (ICC)

Dedico esse trabalho a todos os pacientes com câncer, em especial à minha mãe, Waleska, e à minha avó, Gláucia, vencedoras dessa luta, e à minha amiga e madrastra, Shirley, que, infelizmente, depois de grande batalha, não pode ver esta obra concluída, tendo sido motivo de inspiração a cada dia.

AGRADECIMENTOS

Ao Eterno, que nos concedeu o dom da ciência e que, através de seus mandamentos, nos impele a estudar e buscar conhecimento.

À minha orientadora, Profa Dra Silvia Helena Barem Rabenhorst, que me recebeu como aluno de iniciação científica no primeiro semestre da faculdade de medicina e que tem me mostrado a importância e o quão gratificante é a pesquisa ao longo de todos esses anos, estando presente em toda minha graduação, residência médica e doutorado. Muito obrigado por todos os ensinamentos, dedicação e compreensão durante todos esses anos! Sua dedicação e amor à pesquisa, ao ensino e à ciência são inspiradores.

Aos professores participantes da Banca Examinadora, pelo tempo, pelas valiosas colaborações e sugestões.

Aos meus preceptores da Residência Médica em Anatomia Patológica, hoje em dia colegas de especialidade e amigos, em especial ao Dr Carlos Gustavo Hirth e à Dra Maria do Perpétuo Socorro Saldanha da Cunha (Dra Patty) por todos os ensinamentos e estímulo à pesquisa. Vocês foram fundamentais na minha formação como médico patologista e como pesquisador.

À Unidade de Hemolinfopatologia do Ospedale Sant'Orsola da Universidade de Bolonha, em especial à Dra Elena Sabattini e ao Dr. Claudio Agostinelli, pelos ensinamentos e pelo compartilhamento de experiências e orientações no início deste trabalho.

Aos colegas pesquisadores do LABGEM, em especial aos “meus” alunos de iniciação científica, Igor Frederico e Stella Zancheta, pelo auxílio e dedicação na realização desse trabalho.

Aos técnicos em histologia e em imuno-histoquímica do Instituto do Câncer do Ceará, especialmente Manoela, Veralúcia e Shirley, e do Hospital Infantil Albert

Sabin, Daniele, Bryan e Marianne, por toda ajuda com a preparação do material a ser analisado.

Às minhas amigas e colegas de profissão Débora Barros, Josianne Nascimento, Gislane Rocha, Tamises Melo e Ingrid Frederico pela compreensão e paciência ao longo desse trabalho. Obrigado por terem me coberto no trabalho e por terem emprestado os ouvidos sempre que necessário.

Ao Instituto do Câncer do Ceará pela estrutura concedida.

Ao Laboratório Pathus e à dra Luciana Rocha por terem cedido a hibridizadora necessária para a realização dos experimentos.

Ao meu marido, Vicente Linhares, meu companheiro em tudo, por ter me acompanhado, sempre estimulando e sendo compreensivo, escutando minhas angústias. Você tornou essa caminhada muito mais fácil.

Agradeço à minha família e aos meus amigos. Aos meus pais, Eduardo e Waleska, e avós, Neves, Gláucia e Chico, por todos os ensinamentos e apoio ao longo da vida. Aos meus irmãos, Leonardo, Giovanna e Marcelo por todo companheirismo.

Em especial, agradeço à vovó Ceça, minha maior base, que não pôde acompanhar de perto a realização deste trabalho, mas sempre esteve presente como fonte de inspiração e sempre me estimulou a estudar, a aperfeiçoar-me e a buscar incansavelmente meus sonhos e objetivos.

RESUMO

O Linfoma Difuso de Grandes Células B (LDGCB) é o subtipo mais comum entre os linfomas, apresentando comportamento clínico agressivo e resposta terapêutica heterogênea. Propôs-se, assim, a subdivisão do LDGCB em dois subtipos principais a depender da célula de origem da neoplasia, o subtipo centro germinativo (CG) e não-centro germinativo (nCG), apresentando vias patogênicas distintas e aparente evolução clínica diferente. Adicionalmente, biomarcadores como a expressão das oncoproteínas MYC e BCL2 e a presença de alterações cromossômicas envolvendo os genes *MYC*, *BCL2* e *BCL6* têm demonstrado importância na estratificação prognóstica nos LDGCB. Alterações genéticas no gene *TP53*, importante supressor tumoral, e no gene *EZH2*, que codifica uma subunidade do complexo *PRC2* (*polycomb repressive complex 2*) atuando como uma metiltransferase, parecem estar associadas à patogênese dos LDGCB, embora o significado clínico dessas alterações seja incerto. Por outro lado, diferentes padrões de expressão de microRNAs, como miR-125b e miR-155b, são descritos por influenciar o comportamento biológico desses linfomas, entretanto, a associação desses miRs com os genes descritos é pouco relatada. Outro aspecto relevante nestes tumores é a presença do vírus de Epstein-Barr (EBV), que parece também determinar diferentes padrões prognósticos a depender das células de origem. Assim, o objetivo deste trabalho foi avaliar a importância patogenética e prognóstica das translocações de *MYC*, *BCL2* e *BCL6*, da expressão proteica de BCL2, MYC, TP53 e EZH2, do perfil de expressão de miR-155b e miR-125b e da presença de EBV, considerando a estratificação dos LDGCB pela célula de origem. O estudo dos rearranjos cromossômicos foi realizado por Hibridização *in situ* com fluorescência (FISH), das expressões proteicas por imuno-histoquímica e da expressão de miRNAs e presença EBV por Hibridização *in situ* com estratégias diferenciadas. Cento e trinta e nove casos fizeram parte desse estudo, sendo o subtipo CG discretamente mais frequente que o nCG (53,9% e 46,1%, respectivamente). Em uma primeira abordagem, EZH2 foi significativamente associado à expressão de MYC e TP53 ($p=0.0002$ e $p=0.0000$, respectivamente) e a elevado índice de proliferação celular ($Ki-67>70\%$, $p=0.0082$). O subtipo nCG apresentou as associações observadas na amostra geral, enquanto apenas a expressão de TP53 foi associada à de EZH2 no grupo CG. Nas análises relacionadas ao miRs, foi observado que a detecção de miR-125b correlacionava-se positivamente com o índice de Ki-67 ($p=0.035$) nos nCG. Considerando o subtipo CG, além da associação da positividade de miR-125b com $Ki-67>70\%$ ($p=0.043$), a porcentagem de células positivas foi também correlacionada tanto à expressão de MYC quanto à dupla expressão de MYC/BCL2 ($p=0.047$ e $p=0.049$, respectivamente). Analisando os aspectos clínicos, nos pacientes com LDGCB nCG, a porcentagem e a intensidade da marcação de miR-155b, assim como o

escore de *Allred*, foram correlacionados à progressão da doença ($p= 0.038$, $p=0.057$ e $p= 0.039$, respectivamente). Na análise multivariada, o fenótipo CG demonstrou-se como fator prognóstico independente, associado com maior sobrevida geral ($p=0.007$). Considerando o grupo nCG, embora não significante, as expressões de TP53, miR-125b e miR-155b demonstra uma tendência estatística a funcionar como potenciais biomarcadores de prognóstico nesses tumores. Levando em consideração a presença de EBV, este foi positivo em 7,9% dos pacientes. Apesar da baixa frequência, foi observado que a maioria dos casos EBV(+) apresentavam alto índice de proliferação (72,7%), independentemente do subgrupo. Interessantemente, miR-125b foi observado em praticamente todos esses casos, enquanto miR-155b em apenas dois. Embora a expressão de BCL2 tenha sido frequente nos casos EBV(+), a presença de translocação envolvendo BCL2 foi vista em apenas dois casos. Similarmente, a expressão e a translocação de MYC também foram raramente observadas, enquanto a expressão de TP53 foi relativamente comum (45,4%). Em conclusão, as associações significativas entre EZH2, MYC e TP53, em diferentes padrões de acordo com a célula de origem, indicam potenciais vias patogênicas nesses subtipos. A expressão de miR-125b está relacionada a MYC no subtipo nCG. Adicionalmente, TP53, miR125b e miR-155b parecem representar fatores prognósticos potenciais no subgrupo nCG.

Palavras-chaves: linfoma; biomarcadores; microRNA; EBV

ABSTRACT

Diffuse large B-cell lymphoma (DLBCL) is the most common lymphoma in adults, often with aggressive behavior and representing a clinically and genetically heterogeneous disease. Hence, it has been proposed to classify DLBCL in two subsets by the cell-of-origin into germinal center (GC) and non-germinal center (nGC) subtypes, with apparent distinct pathogenic pathways and differences with respect to clinical course. Additional biomarkers, such as MYC and BCL2 expression and chromosomal translocations involving MYC, BCL2 and BCL6, have been given attention as DLBCL prognostic stratification. Alterations involving TP53, an important tumor suppressor gene, and EZH2, which codifies a polycomb repressive complex 2 (PRC2) subunit and acts as a methyltransferase, seem to be associated to DLBCL pathogenesis, but their clinical impact is not well understood. On the other hand, different microRNA expression patterns, such as miR-125b and miR-155b, are described as key influencers on DLBCL biological behavior, however their association with the aforementioned genes are little reported. Another relevant aspect regarding these tumors is the presence of Epstein-Barr Virus (EBV), which also seem to imply on distinct prognostic patterns depending on the cell-of-origin classification. The aim of this study was to evaluate the pathogenetic and prognostic importance of MYC, BCL2 and BCL6 translocations, BCL2, MYC, TP53 and EZH2 protein expression, miR-125b and miR-155b expression profile and the presence of EBV in a series of DLBCL considering the cell-of-origin classification. Chromosomal rearrangements were carried out by fluorescent in situ hybridization (FISH), protein expressions by immunohistochemistry and microRNA detection, as well as the presence of EBV, by in situ hybridization, with special strategies. Among 139 DBCL cases, the GC subtype was slightly more frequent than the nGC (53.9% and 46.1%, respectively). In a first approach, EZH2 positivity was associated to MYC and TP53 expression ($p=0.0002$ and $p=0.0000$, respectively) and to high proliferative index ($Ki-67>70\%$, $p=0.0082$). The nGC DLBCL presented the associations observed in the general sample; however, only TP53 immunodetection showed associations to EZH2 expression in the GC subtype. When we analyzed the microRNAs, miR-125b detection was positively correlated to the Ki-67 index ($p=0.035$) in the nGC. Considering the GC subgroup, the percentage of miR-125b positive cells was also correlated to both MYC and MYC/BCL2 double expression ($p=0.047$ and $p=0.049$, respectively). When it comes to the clinical aspects, in the nGC patients, miR-155b percentage and intensity, as well as Allred score, were positively correlated to disease progression ($p=0.038$, $p=0.057$ and $p=0.039$, respectively). In a multivariate analysis, GC phenotype was a significant independent factor associated with higher overall survival ($p=0.007$) and, considering the nCG group, although not significant, the expression of TP53, miR-125b and miR-155b showed statistics trends to be regarded as potentials prognostic biomarkers in

these tumors. Taking in consideration EBV, 7.9% of cases were EBV(+)-DLBCL. Although the number of EBV (+) cases is not high, most of the tumors (72.7%) had a high proliferative index (Ki-67>70%) independently on being GC or nGC subtype. miR-125b was detected in almost all of these cases, and conversely, only two cases were miR155b positive. It is noteworthy that BCL2 expression was observed in most cases, although *BCL2* translocation was a rare event, being detected in only two cases. Similarly, MYC expression and MYC translocation were rarely observed, and TP53, on the other hand, was commonly expressed (45.4%). In conclusion, the significant associations among EZH2, MYC and TP53, with different patterns depending on cell-of-origin classification, indicate potential pathogenetic pathways in these subtypes. miR-125b expression is related to MYC in nGC-DLBCL. In addition, TP53, miR-125b and miR-155b seem to represent potential prognostic factors in the nGC subtype.

Key words: lymphoma; biomarkers; microRNA; EBV

SUMÁRIO

1	INTRODUÇÃO.....	11
1.1	Etiologia.....	12
1.2	Características clínicas.....	12
1.3	Diagnóstico.....	13
1.4	Biomarcadores prognósticos.....	16
1.4.1	<i>Rede MYC e TP53.....</i>	16
1.4.2	<i>MicroRNAs.....</i>	17
1.4.3	<i>Vírus de Epstein-Barr (EBV)</i>	19
2	HIPÓTESE.....	20
3	OBJETIVO GERAL.....	20
4	OBJETIVOS ESPECÍFICOS.....	20
5	METODOLOGIA.....	21
5.1	Expressão de MYC, BCL2, TP53 e EZH.....	22
5.2	Translocações em BCL2, BCL6 e MYC.....	24
5.3	Determinação da expressão de miR-125b e miR-155.....	25
5.4	Identificação da presença de EBV.....	27
5.5	Análise estatística.....	28
6	RESULTADOS.....	28
6.1	População de estudo.....	28
7	CAPÍTULO 1.....	31
8	CAPÍTULO 2.....	40
9	CAPÍTULO 3.....	69
10	CONCLUSÕES.....	79
	REFERÊNCIAS.....	84

1 INTRODUÇÃO

Os linfomas não-Hodgkin (LNH) são malignidades histologicamente e molecularmente heterogêneas originadas dos linfócitos. Dentre estes tumores, o subtipo mais comum é o Linfoma Difuso de Grandes Células B (LDGCB), perfazendo aproximadamente 31% dos diagnósticos de LNH, segundo o estudo multicêntrico (THE NON-HODGKIN'S LYMPHOMA CLASSIFICATION PROJECT, 1997). Estimativas mundiais de 2020 mostram que ocorreram, aproximadamente, 540 mil casos novos de linfoma não-Hodgkin, o equivalente a 3% do total dos casos novos estimados. Destes, 304 mil casos novos ocorreram em homens, com um risco estimado de 7,3 por 100 mil, e, nas mulheres, houve 240 mil casos novos, com um risco estimado de 5,2 por 100 mil. As maiores taxas de incidência encontram-se na Austrália, na Nova Zelândia, na América do Norte e nos países do Norte da Europa, em ambos os sexos (GLOBOCAN, 2020). O LDGCB acomete discretamente mais homens que mulheres (relação 1,2:1), com média de idade ao diagnóstico de 64 anos, embora qualquer idade possa ser acometida (JAFFE *et al*, 2017). No Brasil, os linfomas LNH correspondem à oitava neoplasia mais frequente nos homens e à nona neoplasia mais frequente em mulheres, com aproximadamente 12 mil novos casos diagnosticados por ano. Por razões ainda desconhecidas, a incidência desses linfomas dobrou nos últimos 25 anos, principalmente entre pacientes acima de 60 anos. A mortalidade por esta doença é alta, sendo de, aproximadamente, 39% nos cinco primeiros anos do diagnóstico (INCA, 2020).

O LDGCB apresenta comportamento clínico agressivo e, embora uma boa parcela dos pacientes diagnosticados com LDGCB possa ser curada com o esquema quimioterápico padrão baseado em rituximab, ciclofosfamida, doxorrubicina, vincristina e prednisona (R-CHOP), a resposta terapêutica pode ser imprevisível, com muitos pacientes apresentando desfecho desfavorável (LENZ *et al*, 2008). Na prática clínica, o Índice Internacional Prognóstico (IPI) tem sido usado largamente como a ferramenta primária para predizer prognóstico nos pacientes com LNH agressivos e com resposta negativa aos tratamentos, sendo baseado em cinco critérios: idade, estadiamento, níveis sérios de Lactato Desidrogenase (LDH), Performance Status (PS) e número de sítios de acometimento extranodal (LARRABEITI-ETXEBARRIA *et al ET AL*, 2019). Embora seja um sistema reprodutível e facilmente aplicado para estratificação clínica, o IPI não considera as

características biológicas do tumor necessárias para abordagem individualizada dos pacientes (CAO *et al*, 2016).

1.1 Etiologia

A maior parte dos pacientes com LDGCB não apresenta fatores de risco definidos. Raros casos estão relacionados a síndromes de imunodeficiências primárias ou adquiridas, transplante de órgãos ou uso de medicações, como metotrexato e fludarabina. Adicionalmente, uma minoria de casos extranodais pode estar associada à inflamação ou irritação crônica (JAFFE *et al*, 2017).

A maioria dos LDGCB desenvolve-se *de novo*, no entanto, alguns casos são transformações de um linfoma de baixo grau de base que progrediu, como o linfoma folicular, leucemia linfocítica crônica/linfoma linfocítico de pequenas células, linfoma da zona marginal, linfoma linfoplasmocítico ou linfoma de Hodgkin com predominância linfocitária nodular. Raramente, alguns casos de LDGCB podem ainda ocorrer sincronicamente ou metacronicamente com linfoma de Hodgkin clássico (FREEDMAN, 2019).

1.2 Características clínicas

Pacientes com LDGCB frequentemente apresentam uma massa sintomática de crescimento rápido, geralmente um aumento nodal no pescoço ou abdome ou, no caso de linfoma mediastinal primário de grandes células B, no mediastino, mas podem apresentar uma massa em qualquer parte do corpo. O local mais comum de doença primária extranodal é o trato gastrointestinal, especialmente o estômago, mas a doença pode surgir em praticamente qualquer tecido, incluindo testículo, osso, tireoide, glândulas salivares, pele, fígado, mama, suprarrenais, rins, cavidade nasal, anexos oculares, seios paranasais, colo uterino, vagina e sistema nervoso central (FREEDMAN; FREIDBERG; ASTER, 2022).

Sintomas sistêmicos "B" (ou seja, febre, perda de peso, sudorese noturna intensos) são observados em, aproximadamente, 30% dos pacientes, estando normalmente associados à doença agressiva, e a LDH está elevada em mais da metade deles (FREEDMAN; ASTER, 2020). Aproximadamente 60% dos casos são diagnosticados em estágios avançados (III/IV), cujo estadiamento é realizado

baseando-se no sistema de *Ann Arbor*, que leva em consideração o número de cadeias linfáticas e a localização como principais critérios classificatórios. Apesar do tamanho do tumor não determinar alteração no estadiamento, massas iguais ou maiores a 10cm avaliadas por tomografia computadorizada são chamadas de massa “bulk” e parecem estar relacionadas a pior prognóstico (AJCC, 2018). A determinação da *Performance Status (PS)* da *Eastern Cooperative Oncology Group (ECOG)* é outro aspecto clínico relevante, que descreve o nível de funcionalidade do paciente em termos de habilidades de autocuidado, atividade diária e integridade física (Tabela 1) (OKEN *et al*, 1982).

LDGCB é ainda uma doença definidora de AIDS. Quando comparado com o linfoma na população HIV-negativa, o linfoma na população HIV-positiva é caracterizado por sintomas B mais frequentes, doença extranodal, envolvimento de locais incomuns (como cavidades corporais, reto, partes moles) e estágio avançado ao diagnóstico (FREEDMAN, 2019).

Tabela 1 – Descrição dos graus de PS da ECOG

Grau da PS - ECOG	Descrição
1	Assintomático.
2	Sintomático, mas capaz de realizar todas suas atividades normalmente.
3	Sintomático, restrito ao leito <50% do dia.
4	Sintomático, restrito ao leito >50% do dia.
5	Morto

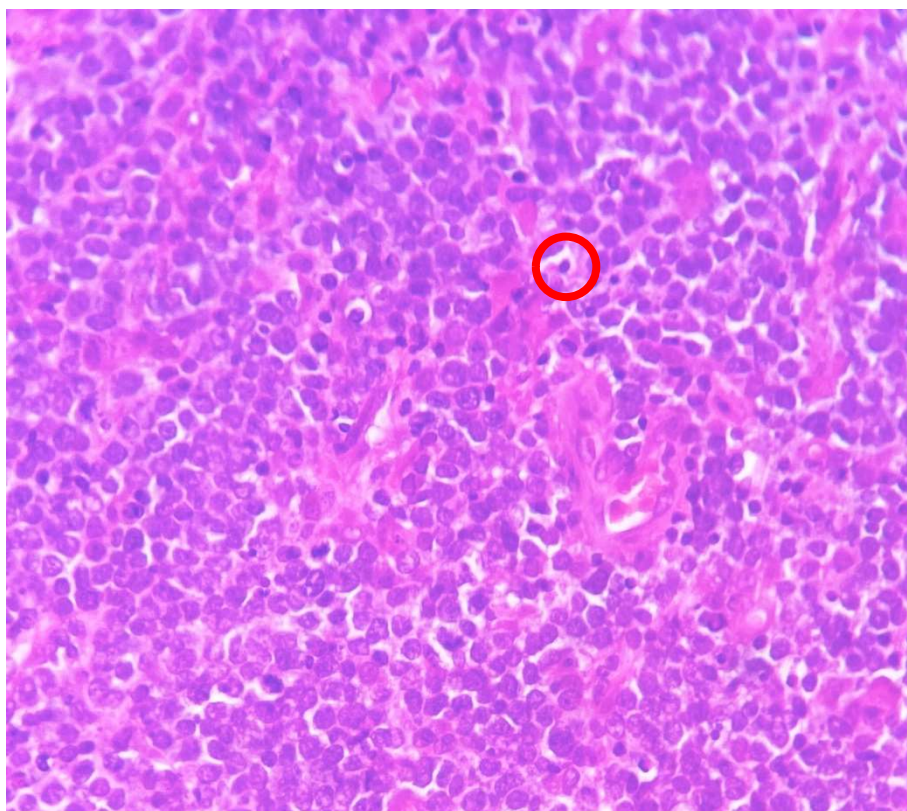
Fonte: elaborada pelo autor.

1.3 Diagnóstico

Os LDGCB são heterogêneos dos pontos de vista clínico, morfológico e biológico (NOGAI; DÖRKEN; LENZ, 2011). Essa entidade é histopatologicamente definida como uma proliferação linfoide atípica difusa formada por células de

tamanho igual ao de um núcleo de histiócito ou maior que o tamanho do núcleo de dois linfócitos normais (Figura 1). Essa definição ampla abrange, consequentemente, um grupo de diversos padrões morfológicos (JAFFE *et al*, 2017). O diagnóstico patológico do LDGCB é, assim, baseado na morfologia e na imunofenotipagem. A imunoexpressão de marcadores pan-B, como CD20 e CD79a, é suficiente para estabelecer o diagnóstico em muitos casos, mas um conjunto muito mais amplo de marcadores imuno-histoquímicos pode ser necessário em casos com características morfológicas atípicas.

Figura 1 – Corte histológico corado em hematoxilina-eosina (HE) representando caso de LDGCB (40x). Note a comparação do tamanho das células neoplásicas com o linfócito normal circulado em vermelho.

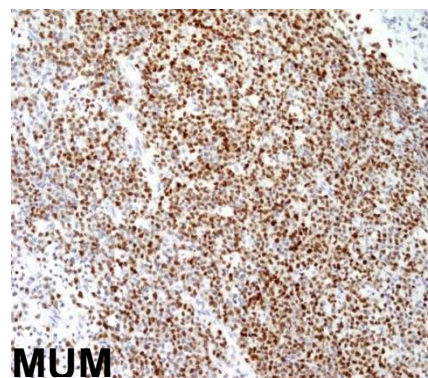
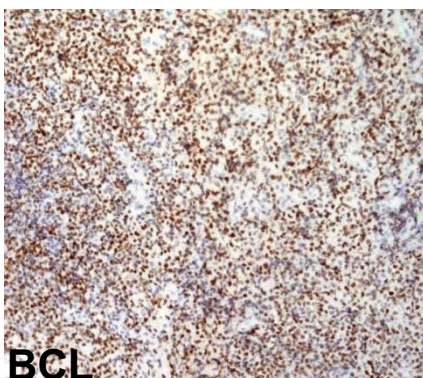
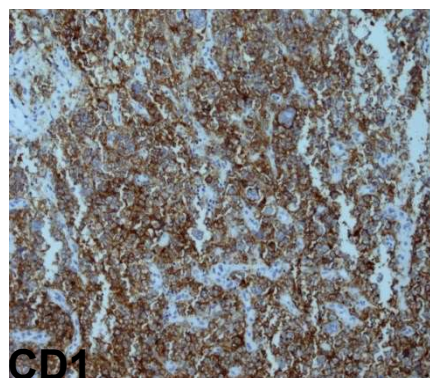
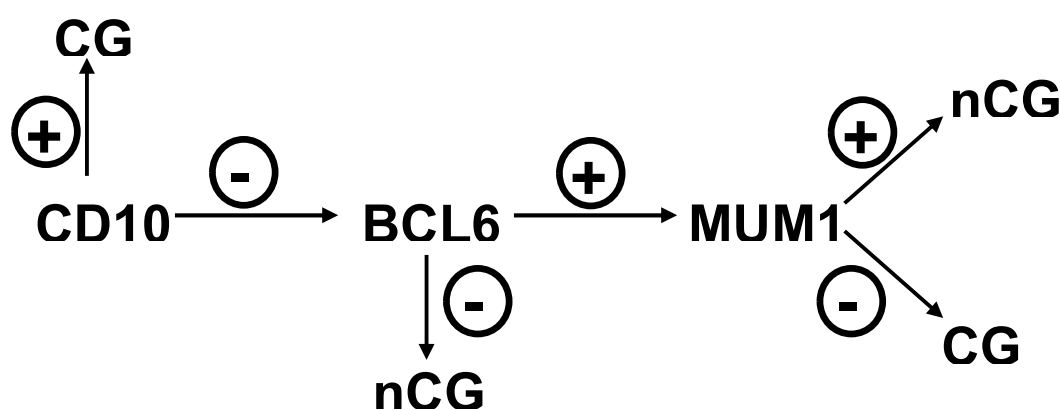


Fonte: elaborada pelo autor.

Nos últimos anos, estudos de perfis de expressão gênica foram realizados (ALIZADEH *et al*, 2000; CAO *et al*, 2016; LENZ *et al*, 2008) e dividiram os LDGCB em três subgrupos, de acordo com a célula de origem: os LDGCB de origem em centro germinativo (GCB), LDGCB de origem em células ativadas (ABC) e Linfoma B difuso Primário do Mediastino (LBPM). Essas subdivisões sugerem que a célula

originária do tumor deriva de células B em diferentes estágios de maturação e apresentam comportamento clínico significativamente diferente (NOGAI; DÖRKEN; LENZ, 2011), justificando o alto índice de mortes decorrentes dessa neoplasia apesar do tratamento, motivando estudos para a compreensão da patogênese dos LDGCB e para a identificação de marcadores preditivos dentro desses subgrupos. A aplicação de técnicas de sequenciamento genético para traçar esses perfis de expressão gênica, entretanto, está restrita a grandes centros, tornando esse método de classificação pouco aplicável na prática diagnóstica. Nesse sentido, protocolos de perfis de expressão proteica, avaliados por imuno-histoquímica, têm sido desenvolvidos como alternativa. Dentre eles, um dos mais usados é o algoritmo de Hans, que se baseia no perfil de expressão de CD10, BCL6 e MUM1 (Figura 2) e subdivide o LDGCB em tipo centro germinativo (CG) e não-centro germinativo (nCG), demonstrando boa correlação com os subtipos moleculares GCB e ABC, respectivamente. O valor prognóstico da utilização desses algoritmos, entretanto, ainda é contraditório (JAFFE *et al*, 2017).

Figura 2 – Esquematisação do algoritmo de Hans



Fonte: elaborada pelo autor.

1. 4 Biomarcadores prognósticos

1.4.1 Rede MYC e TP53

Um avanço significativo na compreensão da patogênese dos LDGCB foi obtido em casos nos quais os rearranjos do gene *MYC* estão presentes. *MYC* é um fator de transcrição que atua em várias vias celulares, incluindo a proliferação celular, sendo rearranjado entre 5% a 15% dos casos de LDGCB. Frequentemente, a translocação envolvendo *MYC* é associada a outras translocações, sendo a mais frequente com a de *BCL2*, gene chave na regulação da apoptose e, em menor frequência, com *BCL6*, fator de transcrição atuante no processo de maturação linfocitária em centro germinativo. Linfomas com recorrentes quebras cromossômicas ativando múltiplos oncogenes, um dos quais o *MYC*, são referidos como linfomas “*double-hit*” ou “*triple-hit*”, incluídos na atualização de 2016 da classificação da Organização Mundial da Saúde (OMS) das doenças hematológicas (JAFFE *et al*, 2017; SWERDLOW *et al*, 2016).

A expressão imuno-histoquímica de *MYC* também tem sido explorada nos LDGCB, sendo detectada entre 30 e 50% dos casos, com a concomitância da expressão com *BCL2* encontrada entre 20 e 35% dos casos. Entretanto, a maioria desses tumores não possui alterações cromossômicas de *MYC* ou de *BCL2*, sendo chamados, então, de “linfomas duplo-expressores”, apresentando melhor comportamento do que os linfomas “*double-hit*”, porém, pior sobrevida em comparação ao LDGCB padrão. Essas observações sugerem que a dupla expressão de *MYC* e de *BCL2* deve ser considerada indicador prognóstico nos LDGCB, embora não defina nova categoria diagnóstica (JOHNSON; SLACK; SAVAGE, 2012; SWERDLOW, 2014, 2016). A definição dessas alterações, entretanto, é ainda insuficiente para explicar a heterogeneidade clínica da doença, de modo que a avaliação de outros marcadores de promissor valor prognóstico e terapêutico é fundamental.

Interessantemente, observa-se que os mecanismos regulatórios envolvendo *MYC* incluem repressão transcricional relacionada a complexos proteicos nos quais a proteína EZH2 é subunidade enzimática. EZH2 é uma metiltransferase que leva à trimetilação da lisina 27 da histona 3 (H3K27me3), uma modificação pós-tradução que culmina com a repressão dos complexos proteicos associados às vias de *MYC*. Adicionalmente, a expressão de EZH2 é uma característica comum de várias

neoplasias hematológicas, mas, em LDGCB, os mecanismos pelos quais essa expressão é regulada e suas associações com características clínico-patológicas e com a resposta terapêutica ao R-CHOP ainda são mal compreendidos (TIAN *et al*, 2016; TIAN; DORFMAN, 2019).

Estudos recentes envolvendo sequenciamento de última geração identificaram diversas mutações somáticas comuns em todos os grupos de LDGCB, mas também perfis de alterações genéticas diferentemente representados nos subtipos GCB e ABC. Entre as anomalias comumente observadas em ambos os subtipos, destacam-se as alterações em *TP53* (PASQUALUCCI; DALLA-FAVERA, 2015; SWERDLOW, 2016). Mutações no gene *TP53* e imunorreatividade para a proteína TP53 ocorrem entre 22% e 40% dos casos de LDGCB, respectivamente, e não há uma correlação estrita entre os dois fenômenos. O papel de *TP53* na gênese dos LDGCB, entretanto, é pouco compreendido (JAFFE *et al*, 2017; PASQUALUCCI; DALLA-FAVERA, 2015; SWERDLOW, 2016).

O supressor tumoral *TP53* age regulando diversos genes-alvo, coordenando as principais defesas celulares contra o crescimento tumoral e promove reparo do DNA, apoptose, controle do ciclo celular e senescência. Assim, mutações em *TP53* levam à desregulação do ciclo celular, à instabilidade genômica e à perda do controle da proliferação celular. Interessantemente, mutações em *TP53* que alteram a função da proteína TP53 têm sido descritas em mais de 50% dos tipos de câncer (TOLEDO; WAHL, 2006; VENTURA *et al*, 2007; XUE *et al*, 2007). Nesse sentido, uma coorte multicêntrica envolvendo 506 pacientes demonstrou que mutações na região codificante de *TP53* estão relacionadas à pior sobrevida em pacientes com LDGCB tratados com esquema R-CHOP tanto do subtipo GCB quanto ABC, sugerindo que a avaliação do *status* de *TP53* pode ser útil para estratificar pacientes em tratamento com R-CHOP em grupos prognósticos distintos (XU-MONETTE *et al*, 2012).

1.4.2 MicroRNAs

Entre os biomarcadores estudados na última década com potencial prognóstico em LDGCB, incluem-se os microRNAs (miRNA). Essas moléculas são sequências curtas de 20 a 24 nucleotídeos não-codificantes que podem controlar a expressão gênica principalmente a nível pós-transcrição. Assim, o miRNA tem um

papel crucial em diversos processos biológicos como a diferenciação celular e a homeostase, o que pode se estender para a iniciação e progressão tumoral e resposta ao tratamento. A desregulação dos miRNA está presente em muitos cânceres, incluindo os LDGCB, podendo agir como oncogene ou supressora tumoral dependendo do tipo celular ou do microambiente (JØRGENSEN *et al*, 2015; NI *et al*, 2016).

Em 2009, Li e colaboradores, utilizando gene chip de microRNA como método para analisar a expressão tecidual de 59 pacientes com LDGCB e de 26 tipos de linhagens celulares de LDGCB, identificaram 63 microRNAs envolvidos no desenvolvimento e na progressão desse linfoma. O miR-155 foi encontrado significativamente superexpresso em LDGCB, onde pacientes LDGCB ABC possuem maior expressão se comparados aos GCB. O papel central de miR-155 na patogênese e na agressividade dos LDGCB tem sido relacionado à ativação das vias de *MYC*, já que a expressão de antagonistas dessa via é diretamente inibida por miR-155 (LAWRIE, 2013). Adicionalmente, estudos apontam a regulação da expressão do gene *TP53* por miRs. Dentre os estudados, o miR-125b parece inibir diretamente a ação de *TP53* quando superexpresso em LDGCB (KUMAR *et al*, 2011; LAWRIE *et al*, 2009). Outro estudo nesse sentido, associou o miR-125b à patogênese de neoplasias hematológicas em modelos de camundongos (ENOMOTO *et al*, 2011) (Figura 3).

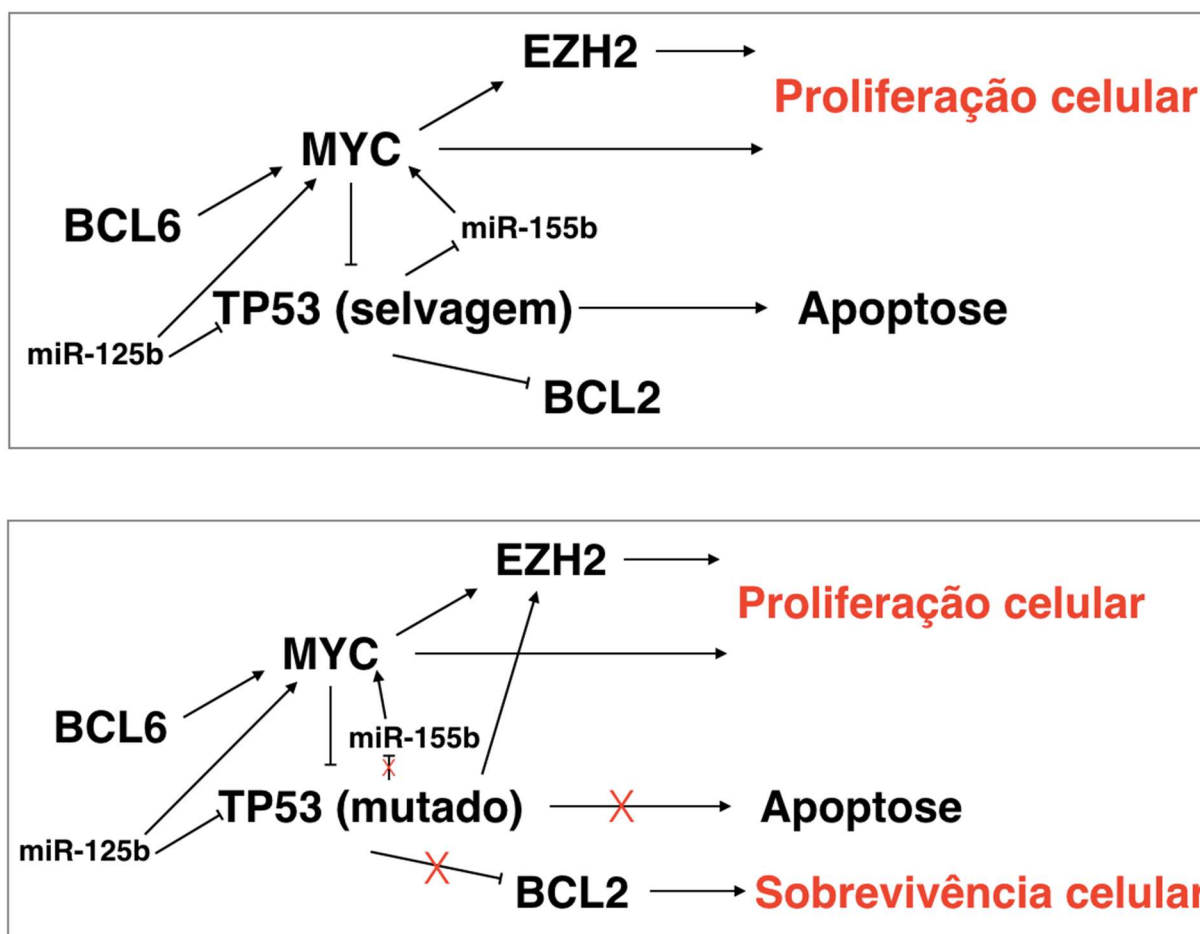


Figura 3 - Esquema representativo da interação dos biomarcadores citados.

Fonte: elaborada pelo autor.

1.4.3 Vírus de Epstein-Barr (EBV)

O vírus de Epstein-Barr (EBV) foi descrito há, aproximadamente, 50 anos em LNH e tem sido demonstrado que 90% da população adulta é portadora do vírus na circulação sanguínea de forma assintomática. Em LDGCB, a presença de EBV está fortemente associada a pacientes imunossuprimidos (JAFTE *et al*, 2017). Embora descrita como baixa em pacientes imunocompetentes, a prevalência de EBV afeta principalmente idosos e é variável geograficamente, com estudos europeus apontando uma positividade de aproximadamente 2% e estudos latino-americanos com taxas próximas às de estudos asiáticos (9%) (HOFSCHEIER *et al*, 2011; HOELLER *et al*, 2009; UNER *et al*, 2011; PARK *et al*, 2007). Interessantemente, a

positividade de EBV tem sido associada à deterioração clínica mais rápida, à resposta terapêutica pobre e à menor sobrevida global e sobrevida livre de doença nesses pacientes (PARK *et al*, 2007). Estratificando os subtipos de LDGCB de acordo com a célula de origem, um estudo coreano apontou ainda uma pior sobrevida global nos pacientes com o subtipo ABC quando associados à positividade para EBV (PARK *et al*, 2007). Estudo recente em pacientes imunossuprimidos pós-transplante demonstrou, inclusive, que o perfil mutacional de *TP53* em LDGCB varia de acordo com a presença ou não de EBV, o que aponta a necessidade de um melhor entendimento da influência viral na patogênese desses linfomas (COURVILLE *et al*, 2016).

Diante do exposto, fica clara a importância de se estudar as alterações em LDGCB de acordo com os subtipos. Frente à necessidade de marcadores prognósticos, o objetivo deste estudo foi investigar a expressão de *EZH2* e o perfil de expressão de *miR-155* e *miR-125b*, além de suas associações com as apresentações imunofenóticas, incluindo a expressão de *MYC*, *BCL2* e *TP53* e os rearranjos cromossômicos envolvendo *MYC*, *BCL2* e *BCL6*, características clínico-patológicas e resposta terapêutica a R-CHOP em uma série de LDGCB.

2 HIPÓTESE

As translocações de *MYC*, *BCL2* e *BCL6*, a expressão proteica de *TP53*, *EZH2*, *BCL2* e *MYC*, o perfil de expressão dos miRNAs *miR-155b* e *miR-125b* e a presença de EBV determinam padrões prognósticos diferenciados nos diferentes tipos moleculares de LDGCB.

3 OBJETIVO GERAL

Avaliar a importância prognóstica das translocações de *MYC*, *BCL2* e *BCL6*, da expressão proteica de *TP53*, *EZH2*, *BCL2* e *MYC*, do perfil de expressão de *miR-155b* e *miR-125b* e da presença de EBV nos diversos subtipos moleculares de LDGCB.

4 OBJETIVOS ESPECÍFICOS

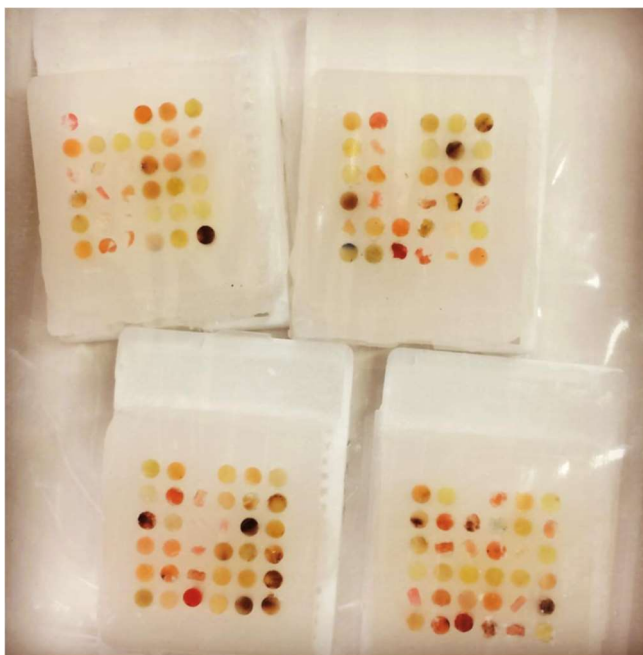
1. Associar a expressão proteica de MYC, BCL2 e EZH2 nos casos de LDGCB de acordo com a célula de origem e com a presença das translocações envolvendo *MYC*, *BCL2* e *BCL6*;
2. Analisar a expressão proteica de TP53 de acordo com os dados clínicos e histopatológicos;
3. Traçar o perfil de expressão de miR-125b e miR-155 nos casos de LDGCB;
4. Determinar a frequência da infecção por EBV nos casos de LDGCB, correlacionando os dados clínico-patológicos com os dados encontrados;
5. Avaliar a importância prognóstica e o potencial terapêutico dos dados encontrados.

5 METODOLOGIA

Trata-se de um estudo longitudinal, retrospectivo, não-intervencionista, cuja população de estudo constou inicialmente de 263 pacientes acompanhados do Instituto do Câncer do Ceará (ICC) com diagnóstico de LDGCB entre os anos de 2011 e 2017, com média de seguimento de 130 meses, aprovado no comitê de ética e pesquisa local sob CAAE 66308017.8.0000.5528. Foram excluídos do estudo os pacientes cujo material anátomo-patológico não estivesse arquivado no Serviço de Patologia do ICC ou cuja amostragem fosse exígua, totalizando 139 casos disponíveis para estudo. Cortes histológicos do espécime cirúrgico corados em hematoxilina-eosina (HE) foram utilizados para avaliação histopatológica, sendo aplicada a classificação revisada da OMS de 2016 e estabelecido o estadiamento clínico de *Ann Arbor* (SWERDLOW, 2014, 2016). Adicionalmente, foi aplicado o algoritmo definido por Hans para distinção entre os subtipos moleculares de acordo com a célula de origem (JAFFE *et al*, 2017). Para avaliação dos marcadores estudados, foram construídos blocos de microarranjo de tecido (TMA, *tissue microarray*), no qual áreas representativas de cada caso foram selecionadas microscopicamente com um *punch* de 2,0mm usando o sistema *Manual Tissue Microarrayer Quick-Ray™ system* (UNITMA Co., Bokjeong-ro, Korea), de acordo com orientações do fabricante (Figura 4). A maioria dos pacientes foi submetida a pelo menos quatro ciclos de R-CHOP ou a protocolos R-CHOP-símiles (R-

miniCHOP e R-COP). A resposta primária ao tratamento (RC, resposta completa; RP, resposta parcial; DE, doença estável e PD, progressão de doença) e o seguimento dos pacientes foram avaliados de acordo com os critérios recomendados (CHESON, 2007).

Figura 4 – Foto representativa dos blocos de TMA construídos para realização das análises de biomarcadores



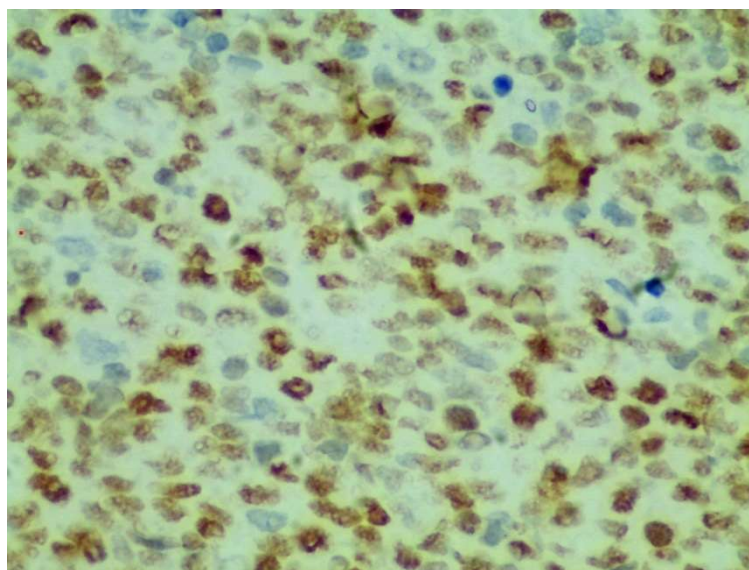
Fonte: elaborado pelo autor.

5.1 Expressão de MYC, BCL2, TP53 e EZH2

A expressão desses marcadores foi avaliada centralmente por imunohistoquímica. Essa técnica foi realizada em cortes histológicos de 2µm dos blocos de TMA usando EZH2 (*Clone 11, dilution 1:50, Cell MarqueTM, Darmstadt, Germany*), MYC (*Clone Y69, dilution 1:50, Abcam, Shanghai, China*), TP53 (*DO-7, ready-for-use, DAKOTM, Glostrup, Denmark*) e BCL2 (*Clone 124, ready-for-use, DAKOTM*). Após desparafinização e recuperação de epítomos utilizando *PT LinkTM DAKO*, os cortes foram incubados através do *AutostainerLink48TM DAKO* e visualizados utilizando polímero *Envision FlexTM DAKO*. Controles positivos e negativos foram utilizados para atestar a fidelidade das reações.

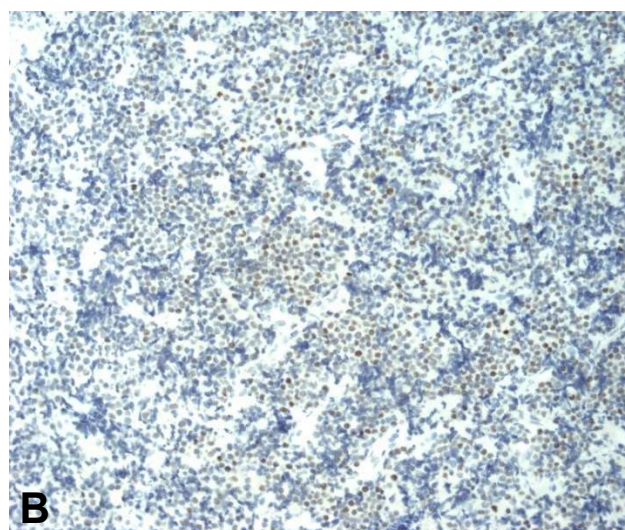
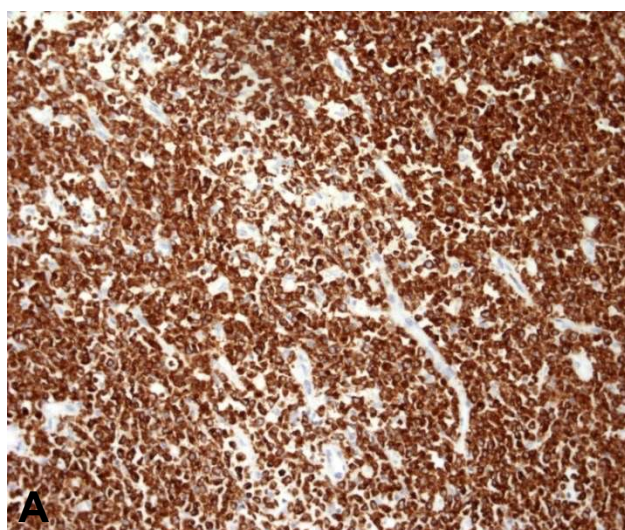
Os valores de positividade foram os seguintes: MYC $\geq 40\%$; BCL2 e TP53 $\geq 50\%$; e EZH2 $\geq 70\%$ (Figuras 5 e 6), como previamente descrito (LIU, 2017; XUMONETTE, 2012). Quando positivas, a intensidade de marcação de TP53 e EZH2 foram classificadas em fraca, moderada e forte (Figura 7) (FEDCHENKO; REIFENRATH, 2014).

Figura 4 – Corte histológico representativo de caso positivo para a expressão de EZH2 (40x)



Fonte: elaborada pelo autor.

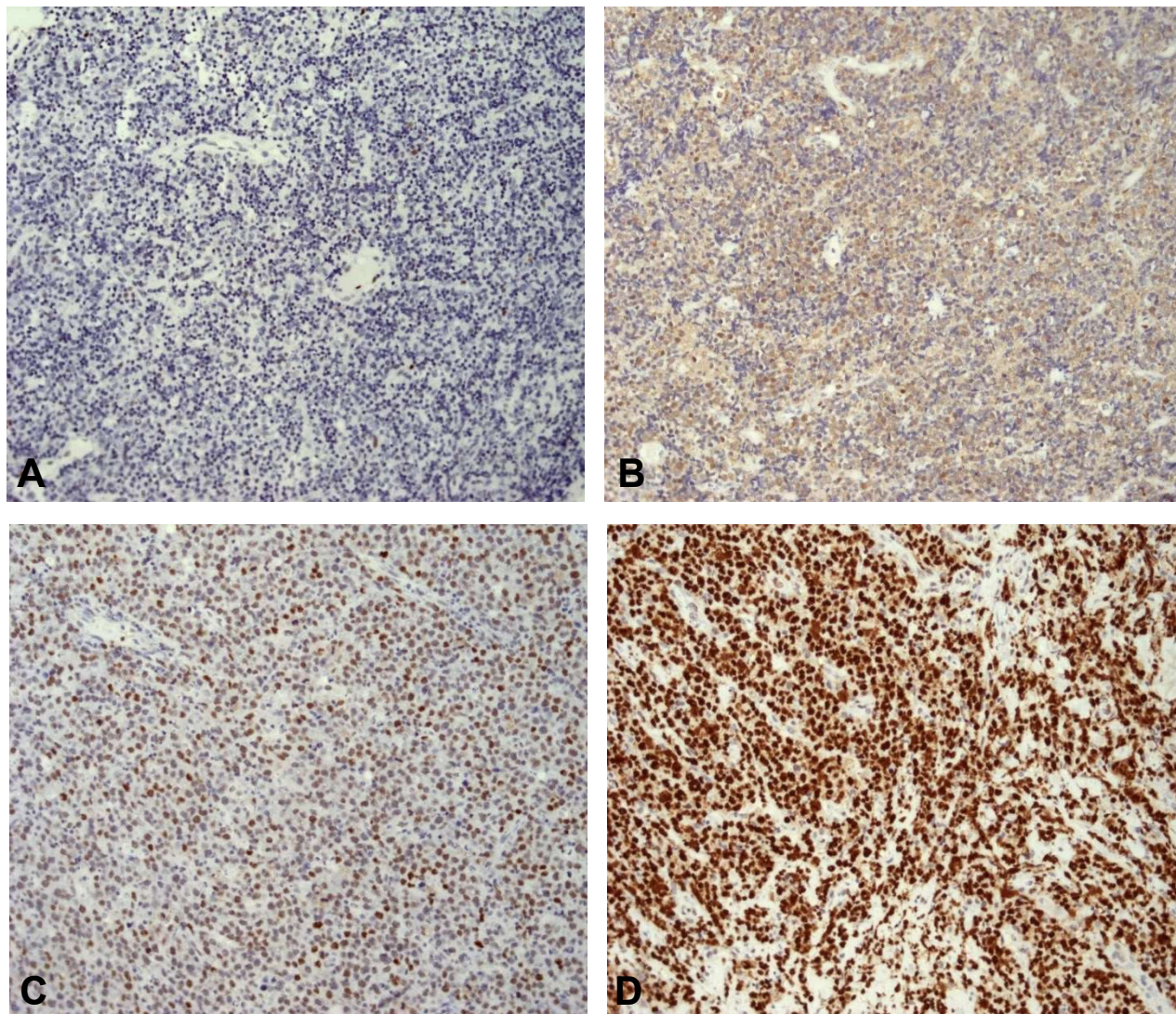
Figura 5 – Corte histológico representativo de caso positivo para a expressão de (A)



BCL2 e (B) MYC (20x)

Fonte: elaborada pelo autor.

Figura 7 – Corte histológico representativo de casos com marcação (A) negativa, (B)



fraca, (C) moderada e (D) forte para a expressão de TP53 (20x)

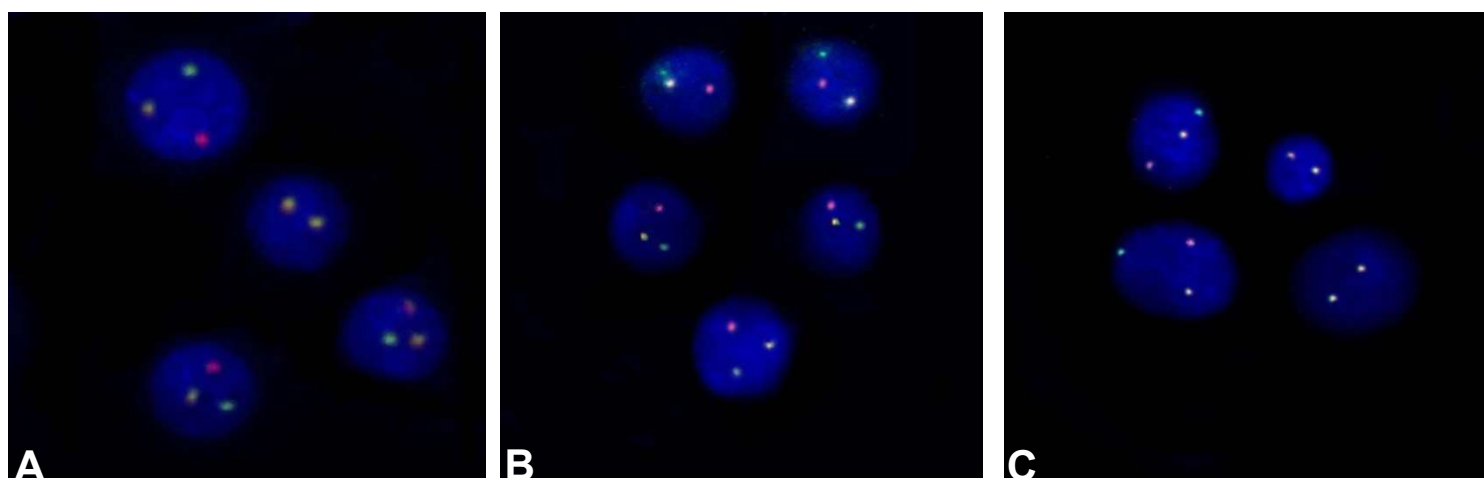
Fonte: elaborada pelo autor.

5.2 Translocações em *BCL2*, *BCL6* e *MYC*

As translocações envolvendo *MYC*, *BCL2* e *BCL6* foram identificadas por Hibridização *in situ* Fluorescente (FISH). Essa técnica foi realizada em cortes de 2,0µm dos blocos de TMA usando sondas de dupla coloração do tipo *break-apart* (*MYC*/8q24 e *BCL6*/3q27) e *dual-color fusion* (*BCL2*/18q21) (*Abbott Laboratories*, Des Plaines, IL) como previamente descrito (*HORN et al*, 2013). As células foram hibridizadas com sondas de *MYC*, *BCL2* e *BCL6* e os núcleos foram contracorados

com 4',6-diamidino-2-fenilindol *antifade*. A fluorescência foi detectada utilizando-se de microscópio de fluorescência *Olympus BX41* (*Olympus*TM, Japão) com filtros de excitação para 4',6-diamidino-2-fenilindol (260 nm) e rodamina (570 nm). Para cada caso, até 200 núcleos em interfase foram analisados através do sistema *ASI image analysis* (*Applied Spectral Imaging*TM, Israel) (Figura 8).

Figura 8 – Corte histológico com fluorescência representativo de casos positivos para translocação em (A) MYC, (B) BCL2 e (C) BCL6



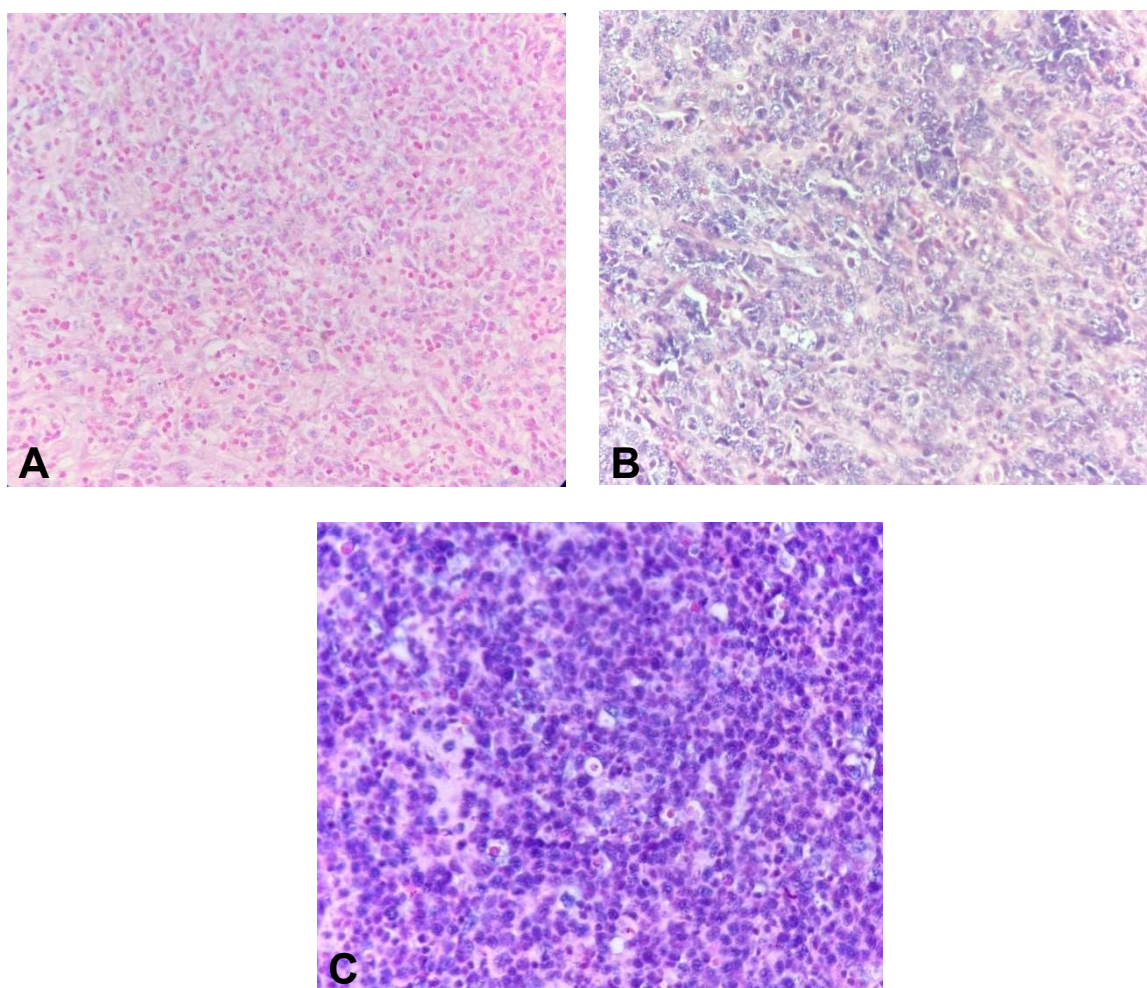
Fonte: elaborada pelo autor.

5.3 Determinação da expressão de miR-125b e miR-155

A expressão dos miRNAs estudados foram avaliadas por Hibridização *in situ*, utilizando-se diferentes sondas de detecção de miRNA de ácido nucleico bloqueado com mercúrio (LNA) (*Exiqon*, Vedbaek, Dinamarca). Essa metodologia é pouco utilizada na pesquisa de expressão de miRNAs, porém é facilmente aplicável à rotina de um laboratório de anatomia patológica e a tecnologia das sondas de LNA garante especificidade de detecção (FAN, 2012). Resumidamente, cortes de 2,0µm dos blocos de TMA foram submetidos a tratamento com proteinase K (15mg/mL) por 10 minutos e posteriormente à hibridização a 53°C durante 90 minutos com sondas hsa-miR-125b-5p (40nM em um tampão ISH livre de formamida), hsa-miR-155-5b (60nM em um tampão ISH livre de formamida), sonda U6 (1nM, como controle positivo) e sonda “scramble miRNA” (pronta para uso, como controle negativo) marcadas com digoxigenina (DIG). Posteriormente, realizaram-se lavagens rigorosas com tampões de SSC a 53°C e à temperatura ambiente. Todos os cortes

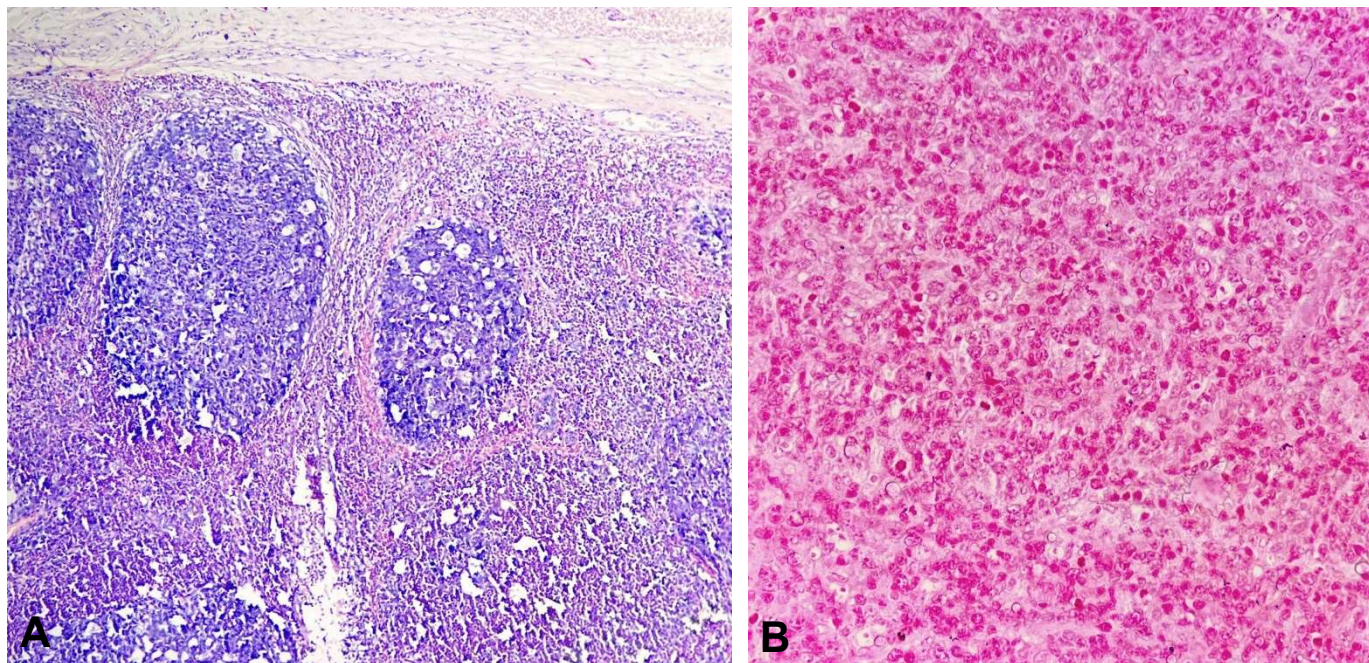
foram incubados com anticorpos anti-DIG (Roche, Alemanha) e submetidos ao método da fosfatase alcalina para revelação com NBT-BCIP e contracoloração com *Fast-Red*, conforme descrito previamente (MUNCH-PETERSEN *et al*, 2015). Consideraram-se positivos os casos com pelo menos 30% das células marcadas de azul em contraste com a contracoloração avermelhada e a intensidade da marcação foi classificada como fraca, moderada ou intensa (Figura 9). Tecido linfoide não neoplásico (tonsila) de quatro pacientes diferentes foi incubado com sondas U6 e “*scramble* miRNA” e utilizado como controle positivo e negativo, respectivamente, das reações (Figura 10). Adicionalmente, as sondas hsa-miR-125b-5p e hsa-miR-155-5b também foram testadas nestes casos a título de comparação com a marcação em amostra neoplásica. Avaliou-se, ainda, o escore de *Allred* para cada caso, como previamente descrito (SARIKOC *et al*, 2013).

Figura 9 – Corte histológico representativo de casos com marcação (A) fraca, (B) moderada e (C) forte para a expressão de miR-125b (40x)



Fonte: elaborada pelo autor.

Figura 10 – Corte histológico representativo de tonsila incubada com (A) sonda U6 (controle positivo) e (B) “scramble RNA” (controle negativo) (20x)

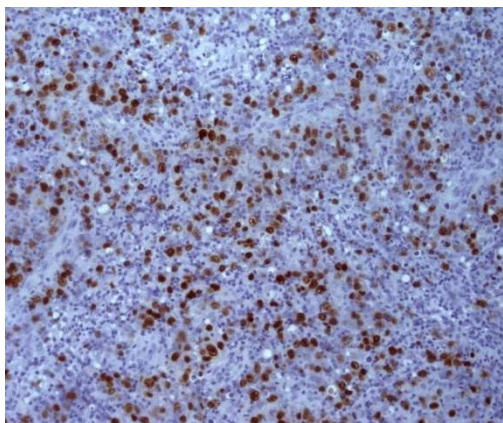


Fonte: elaborada pelo autor.

5.4 Identificação da presença de EBV

A presença de EBV foi avaliada através da técnica de Hibridização *in situ* do RNA usando sondas biotiniladas de 30bp (5'-AGACACCGTCCTCACCACCCGGGACTTGTA-3') complementares ao RNA EBER1, de forma a demonstrar o transcrito de EBV em grande quantidade nos núcleos de células latentemente infectadas, conforme previamente descrito (Figura 11) (FERRASI *et al*, 2010).

Figura 11 – Corte histológico representativo de caso positivo para a expressão de EBER1 (20x)



Fonte: elaborada pelo autor.

5.5 Análise estatística

A análise dos dados foi avaliada através do software *SPSS statistics* 17.0 (*SPSS Inc.*, Chicago, IL, USA). Diferenças estatisticamente significativas foram analisadas pelo teste do χ^2 e pelo teste exato de Fisher. Curvas de sobrevida foram analisadas pelo método de Kaplan-Meier e a análise de regressão de Cox foi utilizada para avaliação do valor prognóstico. Adicionalmente, os dados não-paramétricos foram analisados pelo coeficiente de correlação de Spearman. A significância estatística foi dada por $p < 0,05$.

6 RESULTADOS

6.1 População de estudo

Cento e trinta e nove casos fizeram parte da análise, sendo 52,5% do sexo masculino (73/139) com idade mediana de 61,0 anos, variando de 21 a 98 anos. A maioria dos pacientes apresentou doença nodal (64,0%, 89/139) e, entre aqueles com acometimento extranodal, o trato gastrointestinal foi o sítio mais frequente (34%, 17/50). Levando em consideração a classificação de Hans para a célula de origem, o subtipo centro germinativo foi mais frequente que o não-centro germinativo (53,9%, 75/139 e 46,1%, 64/139, respectivamente). Adicionalmente, EBV foi detectado em onze pacientes (7,9%). A distribuição de características clínicas na população de estudo está resumida na TABELA 2.

Tabela 2 – Distribuição das características clínicas de acordo com o subtipo.

Idade	0,142		
> 60	34 (45,3%)	37 (57,8%)	
≤ 60	41 (54,7%)	27 (42,2%)	
Estágio (Ann Arbor)	0,807		
I-II	35 (59,3%)	43 (61,4%)	
III-IV	24 (40,7%)	27 (38,6%)	
Gênero	0,583		
Feminino	34 (45,3%)	32 (50,0%)	
Masculino	41 (54,7%)	32 (50,0%)	
Performance status (ECOG)	0,270		
0-1	31 (64,6%)	24 (53,3%)	
≥ 2	17 (35,4%)	21 (46,7%)	
LDH	0,887		
Normal	24 (38,7%)	22 (40,0%)	
Aumentado	38 (61,3%)	33 (60,0%)	
Acometimento extranodal	0,707		
> 1	7 (9,3%)	7 (11,3%)	
≤ 1	68 (90,7%)	55 (88,7%)	
Sintomas B			
Presentes	29 (42,6%)	21 (38,2%)	0,616
Ausentes	39 (57,4%)	34 (61,8%)	

Massa <i>Bulky</i>			0,309
Presente	43 (61,4%)	31 (52,5%)	
Ausente	27 (38,6%)	28 (47,5%)	

Fonte: Elaborada pelo autor.

A avaliação da frequência e distribuição dos marcadores e as análises de sobrevida encontram-se nos capítulos 1, 2 e 3.

7 CAPÍTULO 1

Artigo: EZH2 expression is dependent on MYC and TP53 regulation in diffuse large B-cell lymphoma

Autores: Eduardo Henrique Neves Filho, Carlos Gustavo Hirth, Igor Allen Frederico, Rommel Mario Burbano, Thiago Carneiro, Silvia Helena Rabenhorst

Ano de publicação: 2020.

Revista: APMIS (Journal of Pathology, Microbiology and Immunology)

EZH2 expression is dependent on MYC and TP53 regulation in diffuse large B-cell lymphoma

EDUARDO HENRIQUE NEVES FILHO,¹ CARLOS GUSTAVO HIRTH,² IGOR ALLEN
FREDERICO,¹ ROMMEL MARIO BURBANO,³ THIAGO CARNEIRO³ and SILVIA HELENA
RABENHORST¹

¹LABGEM, Departamento de Patologia e Medicina Legal, Universidade Federal Do Ceará, Fortaleza, Brazil;

²Serviço de Patologia, Instituto do Câncer do Ceará, Fortaleza, Brazil; and ³Hospital Ophir Loyola, Belém, Brazil

Neves Filho EH, Hirth CG, Frederico IA, Burbano RM, Carneiro T, Rabenhorst SH. EZH2 expression is dependent on MYC and TP53 regulation in diffuse large B-cell lymphoma. *APMIS* 2020; 128: 308–315.

EZH2 is an important epigenetic regulator, but its role in diffuse large B-cell lymphoma (DLBCL) pathogenesis and its relationship with MYC, BCL2, and TP53 expression, chromosomal rearrangements, and clinical features are still poorly understood. So, we investigated EZH2 expression and its associations with the immunophenotypic presentations, including MYC, BCL2, and TP53 expression, *MYC*, *BCL2*, and *BCL6* translocation status, clinicopathological features, and therapeutic response to R-CHOP in a series of 139 DLBCL cases. EZH2 positivity was associated with MYC and TP53 expression ($p = 0.0002$ and $p = 0.0000$, respectively) and to high proliferative index (Ki67>70%, $p = 0.0082$). No associations were found among EZH2 expression and chromosomal translocation status. The non-germinal center (nGC) DLBCL presented most of associations observed in the general sample; however, only TP53 immunodetection showed associations with EZH2 expression in the germinal center (GC) DLBCL. EZH2 expression had no impact on therapeutic efficacy in R-CHOP-treated patients. In conclusion, EZH2 seems to be upregulated by MYC, to rely on *TP53* alterations, and is associated with high proliferative tumors in DLBCL, which might be dependent on GC or nGC subclassifications. Furthermore, it is not a therapeutic efficacy marker to R-CHOP in our series.

Key words: Diffuse large B-cell lymphoma; EZH2; MYC; TP53; R-CHOP.

Eduardo H. Neves Filho, Laboratório de Genética Molecular (LABGEM), Departamento de Patologia e Medicina Legal, Faculdade de Medicina, Universidade Federal do Ceará, Rua Cel. Nunes de Melo, 1315 Campus do Poranga-bussu CEP 60.183-630, Fortaleza/CE, Brasil. e-mail: edu0689@yahoo.com.br

Diffuse large B-cell lymphoma (DLBCL) is the most common non-Hodgkin lymphoma in adults, accounting for up to 35% of B-cell malignancies, representing a clinically and genetically heterogeneous disease (1). In recent years, gene expression profiling (GEP) has identified at least two distinct subsets of DLBCL with clear phenotypic differences with respect to clinical characteristics and disease course, namely germinal center (GC) versus activated B cell (ABC) (2). Subsequently, immunohistochemical signatures were developed to capture the distinction between these subgroups and to be used as surrogates to GEP in the daily practice, notably Hans algorithm which classifies DLBCL by the cell of origin into germinal center

(GC-DLBCL) and non-germinal center (nGC-DLBCL) subtypes (3, 4). This classification, nevertheless, is insufficient to address DLBCL complexity, suggesting that a variety of molecular pathways with distinct interactions may be involved in DLBCL. Understanding of the molecular mechanisms that contribute to this clinical heterogeneity is relevant to identify patients with more aggressive behavior and an initial step for design alternative therapeutic strategies.

Alterations in *MYC* gene and protein expression in DLBCL define a subset with particularly aggressive behaviors (5, 6). These alterations have been found frequently associated with other oncogenic abnormalities, such as *BCL2* or *BCL6* gene translocations and protein overexpression, which are associated with poor performance status, rapid

evolution, poor response to current therapies, and a median survival of less than 1 year (5, 7).

The regulatory mechanisms involving *MYC* include transcriptional repression due to polycomb repression complexes 1 and 2 (PRC1 and PRC2, respectively) both in *MYC* autoregulation and in general *MYC*-mediated repression (8). EZH2, an enzymatic subunit of the PRC2, has been reported to play a significant role in DLBCL tumorigenesis (9), functioning as a methyltransferase that leads to trimethylation of lysine 27 of histone H3 (H3K27me3), a post-translational modification that leads to transcriptional repression of PRC2 target genes (10). The mutation or overexpression of *EZH2* has been identified in several types of cancers and has been shown to correlate with metastatic progression and poor survival (11). EZH2 expression is a common feature of several hematological neoplasms, but in DLBCL, the mechanisms by which EZH2 expression is regulated and its associations with clinicopathological characteristics and therapeutic response to the standard chemotherapy protocol based on rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone (R-CHOP) are still poorly understood (10, 12, 13). Furthermore, alterations in the tumor suppressor gene *TP53*, an important mediator of cell-cycle arrest, DNA repair, and apoptosis, are an independent marker of poor prognosis in patients with DLBCL and appear to influence EZH2 regulation by poorly understood mechanisms (14, 15).

Hence, the aim of this study was to investigate EZH2 expression and its associations with the immunophenotypic presentations, including *MYC*, *BCL2*, and *TP53* expression and *MYC*, *BCL2*, and *BCL6* translocation status, clinicopathological features, and therapeutic response to R-CHOP in a series of DLBCL.

MATERIALS AND METHODS

Case selections

Samples of 139 lymphoma cases were obtained from the files of the Department of Pathology, Instituto do Câncer do Ceará (Cancer Institute of Ceará), Fortaleza-Brazil, from 2012 to 2017, with the institutional internal review board's approval. The pathological diagnoses were established according to the criteria of the 2016 World Health Organization Classification, based on morphological and immunohistochemical (IHC) findings. Diagnostic slides from all cases were reviewed by two hematopathologists (EHCNF and CGH). Determination of the germinal center (GC) or non-GC phenotype was based on the Hans algorithm (16). Most patients ($n = 90$) underwent at least 4 cycles of R-CHOP or R-CHOP-like regimens. Treatment responses (CR, complete response; PR, partial

response; SD, stable disease; and PD, progressive disease) and follow-up were evaluated according to the recommended criteria (17).

Tissue microarray preparation, immunohistochemistry, and analyses

The representative tumor areas in each case were selected by microscopic examination to make a tissue microarray (TMA) with 2.0 mm punch using Manual Tissue Microarrayer Quick-Ray™ system (Unitma Co., Bokjeong-ro, Korea), according to the manufacturer's instructions. IHC was performed on 2- μ m tissue microarray sections using EZH2 (Clone 11, dilution 1:50, Cell Marque™, Darmstadt, Germany), *MYC* (Clone Y69, dilution 1:50, Abcam, Shanghai, China), *TP53* (DO-7, ready-for-use, DAKO™, Glostrup, Denmark), *BCL2* (Clone 124, ready-for-use, DAKO™), *CD10* (Clone 56C6, ready-for-use, DAKO™), *BCL6* (Clone PG-B6P, ready-for-use, DAKO™), *MUM1* (Clone Mum1P, ready-for-use, DAKO™), and Ki-67 (Clone MIB-1, ready-for-use, DAKO™). All immunoreactions were stained using a fully automated Auto Stainer (DAKO™), in which antigen retrieval was processed through the PTLINK (DAKO™) system with the FLEX Plus (DAKO™) amplification system according to the manufacturer's instructions. The cutoff values for positivity were as follows: *CD10*, *BCL6*, and *MUM1*, $\geq 30\%$; *MYC*, $\geq 40\%$; *BCL2* and *TP53*, $\geq 50\%$; and Ki-67 and EZH2, $\geq 70\%$, as described previously (Fig. 1) (13, 14). When positive, both *TP53* and EZH2 stain intensity were classified as weak, moderate, and intense (18). All samples were independently evaluated by two experienced hematopathologists (EHCNF and CGH) who were blinded to the clinical information. Conflicting results were properly discussed, and a consensus was achieved for all cases.

Fluorescence in situ hybridization

FISH was performed on 2- μ m tissue microarray sections using dual-color break-apart probes (*MYC*/8q24 and *BCL6*/3q27) and dual-color fusion translocation probes (*BCL2*/18q21) (Abbott Laboratories, Des Plaines, IL) as previously described (7). Cells were hybridized with *MYC*, *BCL2*, and *BCL6* probes, and nuclei were counterstained with 4',6-diamidino-2-phenylindole antifade. Fluorescence was detected using a Olympus BX41 fluorescence microscope (Olympus™, Japan) with excitation filters for 4',6-diamidino-2-phenylindole (260 nm) and rhodamine (570 nm). For each case, up to 200 interphase nuclei were analyzed using an ASI image analysis system (Applied Spectral Imaging™, Israel) (Fig. 2). The spots in nuclei were scored as previously described (7).

Statistical analyses

Analyses were carried out using the statistics software SPSS version 17.0 (Chicago, IL, USA). Statistically significant differences were evaluated by the chi-squared test. Correlations were analyzed by Spearman's rank correlation coefficient. The results were considered statistically significant when p values were < 0.05 .

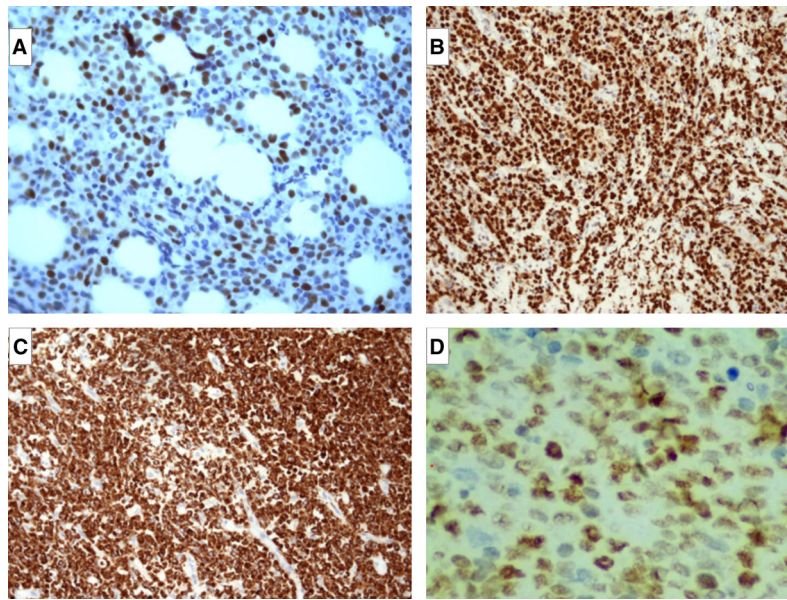


Fig. 1. Immunohistochemical staining of representative cases of (A) MYC, (B) TP53, (C) BCL2, and (D) EZH2 [Colour figure can be viewed at wileyonlinelibrary.com]

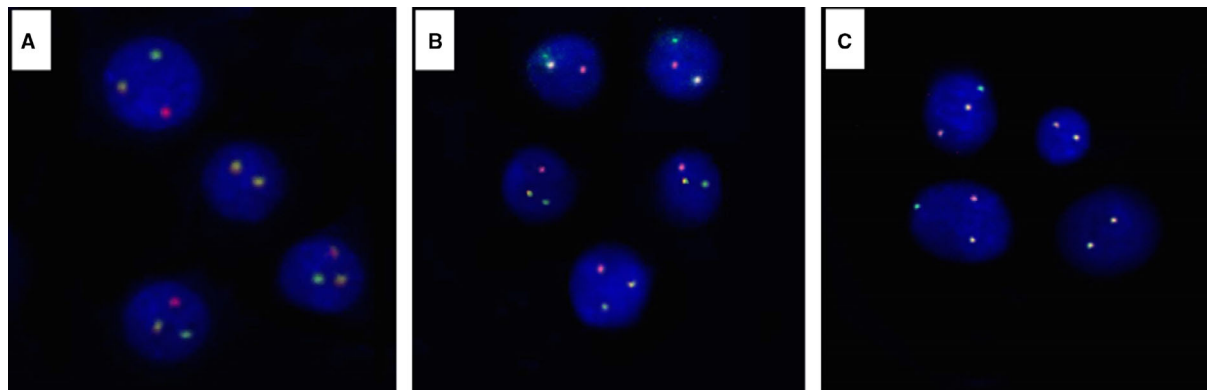


Fig. 2. FISH representative cases of rearranged (A) MYC, (B) BCL2, and (C) BCL6 [Colour figure can be viewed at wileyonlinelibrary.com]

RESULTS

Study population

Among the 139 analyzed cases, 52.5% (73/139) were males. The median age was 61.0 years, ranging from 21 to 98. Most of the patients had nodal disease (55.4%, 77/139), and among those with extranodal lymphoma, the gastrointestinal tract was the most common site (25.8%, 16/62). When the cell-of-origin classification according to Hans algorithm was applied, the germinal center subtype (GC-DLBCL) was slightly more frequent (53.9%, 75/139) than the non-germinal center subtype (nGC-DLBCL) (46.1%, 64/139).

EZH2, MYC, BCL2, and TP53 expression and MYC, BCL2, and BCL6 translocation status

All samples were successfully available for immunohistochemistry reactions of MYC, BCL2, and TP53, and 137 samples were available for EZH2 evaluation. BCL2 was the most frequently expressed (67.6%, 94/139), followed by TP53 (32.3%, 45/139), EZH2 (28.5%, 39/137), and MYC (10.0%, 14/139). Nine cases (6.4%) harbored both MYC and BCL2 immunoreexpression.

EZH2 positivity was significantly associated with MYC and TP53 expression ($p = 0.0002$ and 0.000 , respectively). Moreover, when the EZH2 immunostaining intensity was considered, a positive

Table 1. Relationship among EZH2, MYC, BCL2, and TP53 expression according to Hans algorithm

	GC-DLBCL			nGC-DLBCL		
	EZH2 +	EZH2 –	p	EZH2 +	EZH2 –	p
MYC +	2 (11.1%)	2 (3.6%)	0.2	8 (38.1%)	1 (2.4%)	0.0004
MYC –	16 (88.9%)	54 (96.4%)		13 (61.9%)	41 (97.6%)	
BCL2 +	12 (66.7%)	39 (69.6%)	1	13 (61.9%)	30 (71.4%)	0.6
BCL2 –	6 (33.3%)	17 (30.4%)		8 (38.1%)	12 (28.6%)	
TP53 +	11 (61.1%)	11 (19.6%)	0.0002	12 (57.1%)	9 (21.4%)	0.01
TP53 –	7 (38.9%)	45 (80.4%)		9 (42.9%)	33 (78.6%)	
MYC/BCL2 +	2 (11.1%)	1 (1.8%)	0.1	5 (23.8%)	1 (2.4%)	0.01
MYC/BCL2 –	16 (88.9%)	55 (98.2%)		16 (76.2%)	41 (97.6%)	
MYC/TP53 +	1 (55.6%)	1 (1.8%)	0.4	6 (28.6%)	0 (0%)	0.0008
MYC/TP53 –	17 (94.4%)	55 (98.2%)		15 (71.4%)	42 (100%)	
BCL2/TP53 +	7 (38.9%)	9 (16.1%)	0.08	9 (42.9%)	8 (19.0%)	0.08
BCL2/TP53 –	11 (61.1%)	47 (83.9%)		12 (57.1%)	34 (81.0%)	
Ki67 ≥ 70%	12 (66.7%)	32 (57.1%)	0.6	21 (100%)	26 (61.9%)	0.0006
Ki67 < 70%	6 (33.3%)	24 (42.9%)		0 (0%)	16 (38.1%)	
MYCt +	3 (16.7%)	15 (26.8%)	0.5	7 (33.3%)	5 (11.9%)	0.08
MYCt –	15 (83.3%)	41 (73.2%)		14 (66.7%)	37 (88.1%)	
BCL2t +	3 (16.7%)	7 (12.5%)	0.6	5 (23.8%)	8 (19.0%)	0.9
BCL2t –	15 (83.3%)	49 (87.5%)		16 (76.2%)	34 (81.0%)	
BCL6t +	1 (55.6%)	8 (14.3%)	0.4	1 (4.8%)	0 (0%)	0.3
BCL6t –	17 (94.4%)	48 (85.7%)		20 (95.2%)	42 (100%)	
Double-hit +	1 (55.6%)	7 (12.5%)	0.6	3 (14.3%)	3 (7.1%)	0.4
Double-hit –	17 (94.4%)	49 (87.5%)		19 (85.7%)	39 (92.9%)	

correlation with MYC expression ($r = 0.460$; $p = 0.000$) and with TP53 expression and stain intensity was found ($r = 0.472$; $p = 0.000$ and $r = 0.446$; $p = 0.000$, respectively). In addition, a significant relationship between EZH2 positivity and MYC+/TP53+ double expressors was observed ($p = 0.0006$). Although no relationship between EZH2 and BCL2 was found, 77.8% (7/9) of MYC+/BCL2+ and 87.5% (7/8) of TP53+/BCL2+ cases were statistically associated with EZH2 expression and, also, correlated to EZH2 intensity ($r = 0.396$; $p = 0.001$ and $r = 0.246$; $p = 0.05$, respectively).

Taking in account the proliferation index evaluated by Ki67 expression, EZH2 positivity was related to Ki67>70% ($p = 0.0082$). Additionally, a positive correlation between Ki67 index and EZH2 expression and stain intensity showed up ($r = 0.369$; $p = 0.003$ and $r = 0.394$; $p = 0.001$, respectively). MYC and TP53 were also associated with Ki67>70% ($p = 0.003$ and 0.002 , respectively). Worth noting, MYC+/TP53+, MYC+/BCL2+, and TP53+/BCL2+ were consistently related to Ki67>70% ($p = 0.023$, 0.023 , and 0.012 , respectively).

Considering the translocation status, MYC was the most frequently rearranged gene (21.6%, 30/139), followed by BCL2 (16.5%, 23/139) and BCL6 (7.2%, 10/139). Double-hit translocations were observed in 7.9% (11/139) considering MYC/BCL2 and in 2.2% (3/139) considering MYC/BCL6. No triple-hit alterations were observed in our data. No

relationship between EZH2 expression and the presence of chromosomal translocations involving any studied gene was found.

Cell-of-origin classification and EZH2 analyses

Although no associations were observed considering EZH2 expression and DLBCL subtypes according to Hans algorithm, EZH2 immunostaining was significantly associated either with the lack of CD10 expression ($p = 0.019$) or, conversely, with MUM1 positivity ($p = 0.039$). Noteworthy, MYC was statistically more frequent in the nGC-DLBCL group ($p = 0.042$) and, similarly to EZH2, tended to be associated with the CD10 negativity ($p = 0.070$) and with MUM1 positivity ($p = 0.054$).

The data were also analyzed taking in consideration the Hans algorithm subtypes. The nGC-DLBCL cases had similar results to the general sample, as shown in Table 1. On the other hand, in the CG-DLBCL group, EZH2 expression was only associated with TP53 expression (Table 1) and correlated to TP53 stain intensity ($r = 0.440$; $p = 0.000$). No relationship between EZH2 and proliferation index in the CG-DLBCL cases was found.

Although in the general sample no relationship was observed between EZH2 expression and the presence of chromosomal rearrangements, when we considered the nGC-DLBCL cases, an association between EZH2 expression and MYC rearrangement tended to show up (Table 1).

Table 2. Distribution of EZH2 expression according to clinical response to R-CHOP therapy

	EZH2 +	EZH2–	p
cR	14 (58.4%)	43 (67.2%)	0.6
pR	6 (25.0%)	8 (12.5%)	0.2
SD	2 (8.3%)	10 (15.6%)	0.4
P	2 (8.3%)	3 (4.7%)	0.6
cR + pR	20 (83.3%)	51 (79.7%)	1
SD + P	4 (16.7%)	13 (20.3%)	1

EZH2 expression and clinicopathological associations

No association was found between EZH2 expression and LDH levels, ECOG status, presence of B symptoms, Bulky mass, or disease staging in the general sample. Among the nGC-DLBCL cases, EZH2 tended to be associated with advanced ECOG status (2–4) ($p = 0.05$).

The expression of EZH2 was also evaluated according to the primary response to R-CHOP-based therapy. We were able to analyze this parameter in 90 patients, which is summarized in Table 2. No associations between therapeutic response and EZH2 expression in either the general sample or the Hans subtype groups were found in the analyzed series.

DISCUSSION

Alterations involving *MYC*, *BCL2*, and *BCL6* are well-established prognostic and therapeutic markers in DLBCL, with currently some routine applications (5). In addition, *TP53* status has been pointed as an additional prognostic feature (14). However, these alterations are insufficient to address DLBCL complex molecular landscape and to fully explain its prognostic heterogeneity. So that, EZH2 appears to be a potential biomarker, as it has been shown to be a common DLBCL feature and it seems to be a key part of *MYC* and *TP53* regulatory network (8, 19). In this study, we observed that EZH2 expression is a common event in a DLBCL series, being expressed in about 28% of the cases, similar to a Korean series (20), and we demonstrated a relationship among EZH2, *MYC*, and *TP53*, corroborating with proposals of some authors for EZH2 regulation (15, 21, 22). This is, to our knowledge, the first epidemiological evaluation of EZH2 expression in DLBCL in the Brazilian population.

Overexpression of *MYC* is a common event in lymphoma, including DLBCL. However, it does not appear to be a frequent feature of Brazilian DLBCL patients, as highlighted in our results (10.0%) and in Oliveira *et al.*'s (15.7%) (23). In this study, *MYC* overexpression was significantly associated with EZH2 expression and staining intensity.

There are very few studies associating EZH2 and *MYC*, highlighting Tian *et al.*'s (10, 12). In their first evaluation, they found that EZH2 expression was an important feature of aggressive B-cell neoplasms and its expression was associated with *MYC* overexpression in Burkitt and double-hit lymphomas, but not in DLBCL (12). They proposed that EZH2 is involved in different regulation cascades according to its different types and confirmed the previous cited findings in a subsequent study (10). However, they had a lower number of cases than our series, which may lead to an underestimation of possible associations.

The results of the present study also corroborate Liu *et al.*'s, in which they evaluated EZH2 and *MYC* expression in gastrointestinal DLBCL (13) and showed that EZH2 tends to be overexpressed in a high number of tumor cells, reflecting its role as an oncoprotein and its importance in tumor development.

According to our results, among the 13 *MYC*+ cases, 10 showed EZH2 overexpression, what evidences that when *MYC* is detected, EZH2 tends to be expressed. Indeed, the regulations involving *MYC* and EZH2 are complex. Sander *et al.* suggested that EZH2 regulation by *MYC* might depend on microRNA modulation in lymphoma (24). Sun *et al.*, studying ovarian cancer, also suggested that *MYC* promotes EZH2 expression through miRNA regulation and might play an important role in cell survival (25). These and our results suggest that *MYC* overexpression plays a function in upregulating EZH2 expression.

In addition to *MYC*, *TP53* positivity was significantly associated with EZH2 expression, either considering immunodetection or immunointensity. These findings are corroborated by several studies, which have analyzed *TP53* expression and *TP53* mutational status in solid tumors (15, 26, 27). Likewise, Tang *et al.*, in a *in vitro* study on cell lines, proposed a novel *TP53*-mediated cell-cycle arrest pathway through *TP53*-dependent EZH2 repression (19). Although *TP53* genetic alterations have not been evaluated in our study, *TP53* immunoexpression has been widely correlated to *TP53* mutations (15, 28, 29), including in DLBCL (14), as mutated *TP53* leads to protein stability in comparison with the wild-type form, which has a shorter half-life. So that, our findings suggest that *TP53* impaired transcriptional activity may have a role in EZH2 upregulation in DLBCL.

Furthermore, in this series of DLBCL, EZH2 expression correlated with proliferation rate assessed by Ki-67 staining, which is typically associated with aggressive tumors in B-cell non-Hodgkin lymphoma, as has been previously reported in a

range of human cancers and different hematological neoplasias (12, 23, 30).

In our series of DLBCL, no relationship between EZH2 and BCL2 expression was found, which is in accordance with the literature (13, 30). The fact that MYC+/BCL2+ and TP53+/BCL2+ cases were statistically associated with EZH2 expression was probably due to the interactions with MYC and TP53. MYC, BCL2, and BCL6 translocation status individually or combined (double-hits) was also not associated with EZH2 expression. Although Liu et al. (13) have not evaluated BCL6 rearrangements, they found similar results in gastrointestinal DLBCL for MYC and BCL2 translocations.

No association between EZH2 expression and cell-of-origin group was found in our data. This is consistent with recent studies reporting that abundant EZH2 expression was common in DLBCL regardless of cell of origin (13, 31). Nevertheless, when the analyses were carried out considering each marker involved in Hans algorithm, we found that EZH2 expression was statistically more frequent in CD10-negative and in MUM1-positive cases, which would suggest a non-germinal center-like phenotype.

Considering the cell-of-origin classification and the relationship of EZH2 expression and the other markers, only TP53 immunodetection and stain intensity showed associations with EZH2 expression in the GC-DLBCL. Although TP53 mutations might result in distinctive molecular programs harbored by GC and nGC-DLBCL (14), according to our results, TP53 impairment estimated by TP53 overexpression seems to upregulate EZH2 expression regardless of DLBCL subtype. On the other hand, EZH2 upregulation by MYC expression seems to be a restricted feature to the nGC-DLBCL cases, as in the GC-DLBCL group no relationship between MYC and EZH2 expression was observed. Also, EZH2 expression tended to be associated with MYC translocation in the nGC-DLBCL. This may possibly be explained by a direct correlation of MYC translocation and MYC expression in this group. These results suggest that, considering these genes, EZH2 upregulation is dependent on MYC overexpression rather than on MYC rearrangements.

In the present study, no association between EZH2 expression and well-known prognostic factor including poor performance status, elevated LDH, B symptoms, and disease staging was noted. Considering the nGC-DLBCL group, however, a poor performance status defined by ECOG scores ≥ 2 showed a borderline association with EZH2 expression, which is consistent with Liu et al. study (13).

Other authors studying trimethylated H3K27 (H3K27me3) expression, a direct consequence of EZH2 upregulation in many cancers, found that its overexpression is also associated with an ECOG status 2-4 (10, 30, 32). Furthermore, Oh et al. (30) suggest that H3k27me3 expression is associated with poorer survival specially in the nGC subtype, what might explain that, in our cases, a EZH2-associated poor performance status in nCG-DLBCL is due to H3k27me3 effect. To identify these possible mechanisms, however, further studies should follow.

EZH2 expression, in this study, had no impact on therapeutic efficacy in R-CHOP-treated patients. Oh et al. (30) also failed on finding EZH2 expression as a therapeutic factor. However, the current literature regarding these aspects is limited and controversial. As a matter of fact, Deng et al. (33, 34), in China, found that patients with EZH2 low expression usually responded well to R-CHOP regimen in the short-term or long-term survival in comparison with those of patients with higher expression, and Lee et al. (32), in Korea, found that high-level expression of EZH2 was associated with superior overall survival compared to low expression. Our study may be limited as it is from a single institute. Further study, especially multicenter-based studies, should be followed for validating the present results.

In conclusion, EZH2 seems to be upregulated by MYC expression, to rely on TP53 alterations, and is associated with high proliferative tumors in DLBCL, which might be dependent on GC or nGC subclassifications. In addition, although EZH2 was associated with poor performance status in nGC-DLBCL, it is not a therapeutic efficacy marker to R-CHOP in our series.

This work was supported by the Fundação Cearense de Apoio ao Desenvolvimento Científico e Tecnológico (FUNCAP) [PPSUS 2017 grant number 3925618/2017].

REFERENCES

1. Chapuy B, Stewart C, Dunford AJ, Kim J, Kamburov A, Redd RA, et al. Molecular subtypes of diffuse large B cell lymphoma are associated with distinct pathogenic mechanisms and outcomes. *Nat Med* 2018;24:679–90.
2. Karmali R, Gordon LI. Molecular subtyping in diffuse large B cell lymphoma: closer to an approach of precision therapy. *Curr Treat Options Oncol* 2017;18:11.
3. Carbone A, Gloghini A, Kwong YL, Younes A. Diffuse large B cell lymphoma: using pathologic and molecular biomarkers to define subgroups for novel therapy. *Ann Hematol* 2014;93:1263–77.

4. Boltežar L, Prevodnik VK, Perme MP, Gašljević G, Novaković B. Comparison of the algorithms classifying the ABC and GCB subtypes in diffuse large B-cell lymphoma. *Oncol Lett* 2018;15:6903–12.
5. Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, et al. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood* 2016;127:2375–90.
6. Karube K, Campo E. MYC alterations in diffuse large B-cell lymphomas. *Semin Hematol* 2015;52:97–106.
7. Horn H, Ziepert M, Becher C, Barth TFE, Bernd H-W, Feller AC, et al. MYC status in concert with BCL2 and BCL6 expression predicts outcome in diffuse large B-cell lymphoma. *Blood* 2013;121:2253–63.
8. Cole MD. MYC association with cancer risk and a new model of MYC-mediated repression. *Cold Spring Harb Perspect Med* 2014;4:a014316.
9. Velichutina I, Shaknovich R, Geng H, Johnson NA, Gascoyne RD, Melnick AM, et al. EZH2-mediated epigenetic silencing in germinal center B cells contributes to proliferation and lymphomagenesis. *Blood* 2010;116:5247–55.
10. Tian X, Xu J, Dorfman DM. Utility of combined EZH2, p-ERK1/2, p-STAT, and MYC expression in the differential diagnosis of EZH2-positive Hodgkin Lymphomas and related large B-cell lymphomas. *Am J Surg Pathol* 2019;43:102–9.
11. Bisserier M, Wajapeyee N. Mechanisms of resistance to EZH2 inhibitors in diffuse large B-cell lymphomas. *Blood* 2018;131:2125–2137.
12. Tian X, Pelton A, Shahsafaei A, Dorfman DM. Differential expression of enhancer of zeste homolog 2 (EZH2) protein in small cell and aggressive B-cell non-Hodgkin lymphomas and differential regulation of EZH2 expression by p-ERK1/2 and MYC in aggressive B-cell lymphomas. *Mod Pathol* 2016;29:1050–7.
13. Liu Y, Yu K, Li M, Zeng K, Wei J, Li X, et al. EZH2 overexpression in primary gastrointestinal diffuse large B-cell lymphoma and its association with the clinicopathological features. *Hum Pathol* 2017;64:213–21.
14. Xu-Monette ZY, Wu L, Visco C, Tai YC, Tzankov A, Liu WM, et al. Mutational profile and prognostic significance of TP53 in diffuse large B-cell lymphoma patients treated with R-CHOP: report from an International DLBCL Rituximab-CHOP Consortium Program Study. *Blood* 2012;120:3986–96.
15. Shioyama S, Yoshida S, Soga D, Motohashi H, Shin-tani S. Aberrant expression of EZH2 is associated with pathological findings and P53 alteration. *Anticancer Res* 2013;33:4309–17.
16. Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delabie J, Ott G, et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. *Blood* 2004;103:275–82.
17. Cheson BD. The International Harmonization Project for response criteria in lymphoma clinical trials. *Hematol Oncol Clin North Am* 2007;21:841–54.
18. Fedchenko N, Reifensath J. Different approaches for interpretation and reporting of immunohistochemistry analysis results in the bone tissue - a review. *Diagn Pathol* 2014;9:221.
19. Tang X, Milyavsky M, Shats I, Erez N, Goldfinger N, Rotter V. Activated p53 suppresses the histone methyltransferase EZH2 gene. *Oncogene* 2004;23:5759–69.
20. Oh EJ, Kim SH, Long YWI, Ko YH, Yoon SO. Non-coding RNA HOTAIR expression in diffuse large B-cell lymphoma. In relation to polycomb repressive complex pathway proteins and H3K27 trimethylation. *J Pathol Transl Med* 2016;50:369–76.
21. Schmitz R, Wright GW, Huang DW, Johnson CA, Phelan JD, Wang JQ, et al. Genetics and pathogenesis of diffuse large B-cell lymphoma. *N Engl J Med* 2018;378:1396–407.
22. Chen Q, Zheng PS, Yang WT. EZH2-mediated repression of GSK-3 β and TP53 promotes Wnt/ β -catenin signaling-dependent cell expansion in cervical carcinoma. *Oncotarget* 2016;7:36115–29.
23. Oliveira CC, Maciel-Guerra H, Kucko L, Hiram EJ, Brilhante AD, Quevedo FC, et al. Double-hit lymphomas: clinical, morphological, immunohistochemical and cytogenetic study in a series of Brazilian patients with high-grade non-Hodgkin lymphoma. *Diagn Pathol* 2017;12:3.
24. Sander S, Bullinger L, Klapproth K, Fiedler K, Kestler HA, Barth TF, et al. MYC stimulates EZH2 expression by repression of its negative regulator miR-26a. *Blood* 2008;112:4202–12.
25. Sun J, Cai X, Yung MM, Zhou W, Li J, Zhang Y, et al. miR-137 mediates the functional link between c-Myc and EZH2 that regulates cisplatin resistance in ovarian cancer. *Oncogene* 2019;38:564–80.
26. Pietersen AM, Horlings HM, Hauptmann M, Langerød A, Ajouaou A, Cornelissen-Steijger P, et al. EZH2 and BMI1 inversely correlate with prognosis and TP53 mutation in breast cancer. *Breast Cancer Res* 2008;10:R109.
27. Yamada A, Fujii S, Daiko H, Nishimura M, Chiba T, Ochiai A. Aberrant expression of EZH2 is associated with a poor outcome and P53 alteration in squamous cell carcinoma of the esophagus. *Int J Oncol* 2011;38:345–53.
28. Freed-Pastor WA, Prives C. Mutant p53: one name, many proteins. *Genes Dev* 2012;26:1268–86.
29. Frazier MW, He X, Wang J, Gu Z, Cleveland JL, Zambetti GP. Activation of c-myc gene expression by tumor-derived p53 mutants requires a discrete C-terminal domain. *Mol Cell Biol* 1998;18:3735–43.
30. Oh EJ, Yang WI, Cheong JW, Choi SE, Yoon SO. Diffuse large B-cell lymphoma with histone H3 trimethylation at lysine 27: another poor prognostic phenotype independent of c-Myc/Bcl2 coexpression. *Hum Pathol* 2014;45:2043–50.
31. Zhou Z, Gao J, Popovic R, Wolniak K, Parimi V, Winter JN, et al. Strong expression of EZH2 and accumulation of trimethylated H3K27 in diffuse large B-cell lymphoma independent of cell of origin and EZH2 codon 641 mutation. *Leuk Lymphoma* 2015;56:2895–901.
32. Lee HJ, Shin DH, Kim KB, Shin N, Park WY, Lee JH, et al. Polycomb protein EZH2 expression in diffuse large B-cell lymphoma is associated with better prognosis in patients treated with rituximab, cyclophosphamide, doxorubicin, vincristine and prednisone. *Leuk Lymphoma* 2014;55:2056–63.

33. Deng YJ, Chen G, Zhu WF, Rui HB, Hong HL, Chen JM. Expression and prognostic significance of H3K27 trimethylation protein in DLBCL. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 2016;24:1379–85.
34. Deng YJ, Chen G, Zhu WF, Chen XH, Hong HL, Huang LM, et al. Significance of H3K27me3 and EZH2 in predicting the therapeutic efficacy of diffuse large B-cell lymphoma. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 2018;26:159–65.

CAPÍTULO 2

Artigo: Prognostic impact of miR-125b and miR-155b and their relationship with MYC and TP53 in diffuse large B-cell lymphoma: cell-of-origin classification matters.

Autores: Eduardo Henrique Neves Filho, Stella Barbanti Zancheta, Paulo Góberlânio de Barros Sousa, Rommel Mario Burbano, Silvia Helena Rabenhorst

Ano de aceite: 2023.

Revista: Journal of Clinical and Experimental Hematopathology.

PROGNOSTIC IMPACT OF MIR-125B AND MIR-155B AND THEIR RELATIONSHIP WITH *MYC* AND *TP53* IN DIFFUSE LARGE B-CELL LYMPHOMA: CELL-OF-ORIGIN CLASSIFICATION MATTERS.

Running title: miR-125b and miR155b detection in diffuse large B-cell lymphoma

EDUARDO HENRIQUE CUNHA NEVES FILHO¹, STELLA BARBANTI ZANCHETA¹, PAULO GOBERLÂNIO DE BARROS SILVA², ROMMEL MARIO RODRÍGUEZ BURBANO³, SILVIA HELENA BAREM RABENHORST¹.

¹LABGEM - Departamento de Patologia e Medicina Legal - Universidade Federal Do Ceará - Fortaleza/Brazil.

¹UNICHRISTUS - Fortaleza/Brazil.

³Hospital Ophir Loyola - Belém/Brazil.

Declaration of interests: none.

Acknowledgement: This work was supported by the Fundação Cearense de Apoio ao Desenvolvimento Científico e Tecnológico (FUNCAP) [PPSUS 2017].

Corresponding author:

Dr Eduardo H C Neves Filho, MD
Laboratório de Genética Molecular (LABGEM).
Departamento de Patologia e Medicina Legal - Faculdade de Medicina
Universidade Federal do Ceará
Rua Cel. Nunes de Melo, 1315
Campus do Porangabussu CEP 60.183-630
Fortaleza/CE
Brasil
edu0689@yahoo.com.br
Phone: +55 85 981810904

ABSTRACT

Tumoral microRNAs, such as miR-125b and miR-155b, are important gene expression regulators with complex pathogenetic mechanisms. However, their role in DLBCL, especially when cell-of-origin classification is considered, are still to be elucidated. In a series of 139 DLBCL cases considering germinal center (GC) *versus* nonGC subtypes, we investigated miR-125b and miR-155b expression by *in situ* hybridization and their association with some immunophenotypic presentations, including MYC, BCL2 and TP53 expression, *MYC*, *BCL2* and *BCL6* translocation status, as well as clinicopathological features and outcomes. miR-125b detection was positively correlated to the Ki-67 index ($p=0.035$) in the nGC. Considering the GC subgroup, the percentage of miR-125b positive cells was also correlated to either MYC and MYC/BCL2 double expression ($p=0.047$ and $p=0.049$, respectively). When it comes to nGC patients, miR-155b percentage and intensity, as well as Allred score, were positively correlated to disease progression ($p=0.038$, $p=0.057$ and $p=0.039$, respectively). In a multivariate analysis, GC phenotype was a significant independent factor associated with higher OS ($p=0.007$) and, considering the nGC group, although not significant, the expression of TP53, miR-125b and miR-155b seems to be potential prognostic biomarkers in these tumors. This study demonstrated different pathways based on cell-of-origin classification and highlighted different clinical outcomes. miR-125b, miR-155b and TP53 expression may also represent potential prognostic factors in nGC-DLBCL.

KEY WORDS: Diffuse Large B-Cell Lymphoma, miR-125b, miR-155b, MYC, TP53, R-CHOP.

1. INTRODUCTION

Diffuse large B-cell lymphoma (DLBCL) is the most common non-Hodgkin lymphoma in adults, accounting for up to 35% of B-cell malignancies, representing a biologically heterogeneous disease with unpredictable clinical outcomes. Despite the standard chemotherapy protocol based on rituximab, cyclophosphamide, doxorubicin, vincristine and prednisone (R-CHOP), 20% to 30% of patients still experience relapse or refractory disease [1]. DLBCL has been defined by genetic expression profiling (GEP) in mainly two distinct subsets named germinal center (GC) and activated B cell (ABC) with notable phenotypic differences with respect to clinical characteristics and disease course, but still with very limited application in clinical practice [2]. Subsequently, immunohistochemical protocols were developed to be used as surrogates to GEP. Notably, Hans algorithm, which classifies DLBCL in the cell - of - origin into germinal center (GC-DLBCL) and in non-germinal center (nGC-DLBCL) subtypes, showing a good correlation with GEP defined entities and potential prognostic impact [3]. This classification, nevertheless, is insufficient to address DLBCL complexity, suggesting that a variety of molecular pathways with distinct interactions may be involved in DLBCL.

Several biomarkers have been shown to predict the outcome of DLBCL, but only a few have clinical applicability, reinforcing the necessity to identify their pathways. In a previous publication [4], changes in the interaction of *MYC*, *BCL2*, *BCL6*, and *TP53* may imply in loss of cell proliferation and apoptotic control, impaired DNA-repair mechanisms and B-cell maturation disturbance. Furthermore, these intricate interactions seem to vary upon GC versus nGC subgroups, with different molecular pathways between both subtypes with variable clinical outcomes, suggesting that GC and nGC phenotypes and their characteristic biological profile should be considered as two different entities.

In the last few years, changes in noncoding RNAs have been shown relevant in many cancers, addressing the importance of epigenetic alterations and aberrant expression. In this context, microRNAs (miRs) came up as a group of biomarkers with potential prognostic impact on DLBCL, as an endogenous gene expression regulator [5, 6]. For instance, miR-125b became relevant in cancer research, being often associated to cell proliferation, apoptosis inhibition and loss of tumor suppres-

sive functions [7]. Although miR-125b is aberrantly expressed in a variety of tumors and has either oncogene and tumor suppressor activity, the expression, function, and clinical relevance of miR-125b in both DLBCL GC and nGC subgroups are still unclear. In addition, miR-155b has been demonstrated to have a key role in hematopoiesis and immune response, acting as an oncomiR in many malignancies, especially in lymphoma [8]. Indeed, some studies have pointed miR-155b as a marker of malignancy in B-cell lymphoproliferative disorders, and even as a potential tool to differentiate ABC from GCB-DLBCL, but its prognostic role is still controversial [9, 10].

The aim of this study was to investigate the association of miR-125b and miR-155b with MYC, BCL2 and TP53 expression, *MYC*, *BCL2* and *BCL6* translocation status along with clinicopathological features, therapeutic response to R-CHOP and clinical outcomes in DLBCL, focusing in the differences between GC and nGC subtypes.

2. MATERIALS AND METHODS

2.1 Case selections

Samples of 139 lymphoma cases were obtained from the files of the Department of Pathology, Instituto do Câncer do Ceará (Cancer Institute of Ceará), Fortaleza-Brazil, from 2012-2017, with the institutional internal review board's approval. The pathological diagnoses were established according to the criteria of the 2022 World Health Organization classification, based on morphological and immunohistochemical (IHC) findings. Diagnostic slides from all cases were reviewed by two hematopathologists. Determination of the germinal center (GC) or non-GC phenotype was based on the Hans' algorithm [3]. Most patients (n = 92) underwent at least 4 cycles of R-CHOP or R-CHOP-like regimens and only these cases were considered in univariate and multivariate analyses. All other patients (n = 47) were submitted to heterogeneous treatment regimens and were not considered in such analyses. Treatment responses (CR, complete response; PR, partial response; SD, stable disease and PD, progressive disease) and follow-up were evaluated according to the recommended criteria [11].

2.2 Tissue microarray preparation and immunohistochemistry analyses

The representative tumor areas in each case were selected by microscopic examination to make a tissue microarray (TMA) with a 2.0 mm punch using Manual Tissue Microarrayer Quick-Ray™ system (UNITMA Co., Bokjeong-ro, Korea), according to the manufacturer's instructions. IHC was performed on 2-μm tissue microarray sections using MYC (Clone Y69, dilution 1:50, Abcam, Shanghai, China), TP53 (DO-7, ready-for-use, DAKO™, Glostrup, Denmark), BCL2 (Clone 124, ready-for-use, DAKO™), CD10 (Clone 56C6, ready-for-use, DAKO™), BCL6 (Clone PG-B6P, ready-for-use, DAKO™), MUM1 (Clone Mum1P, ready-for-use, DAKO™) and Ki-67 (Clone MIB-1, ready-for-use, DAKO™). Immunoreactions were stained with a fully automated Auto Stainer (DAKO™), in which antigen retrieval was processed through PTLINK (DAKO™) with FLEX Plus (DAKO™) amplification system according to the manufacturer's instructions. The cutoff values for positivity were considered as follows: CD10, BCL6, and MUM1, $\geq 30\%$; MYC, $\geq 40\%$; BCL2 and TP53, $\geq 50\%$; and Ki-67, $\geq 70\%$, as described previously [4]. When positive, TP53 staining intensity was classified as weak, moderate and intense [12]. All samples were independently evaluated by two experienced hematopathologists who were blinded to the clinical information. Conflicting results were properly discussed and a consensus was achieved for all cases.

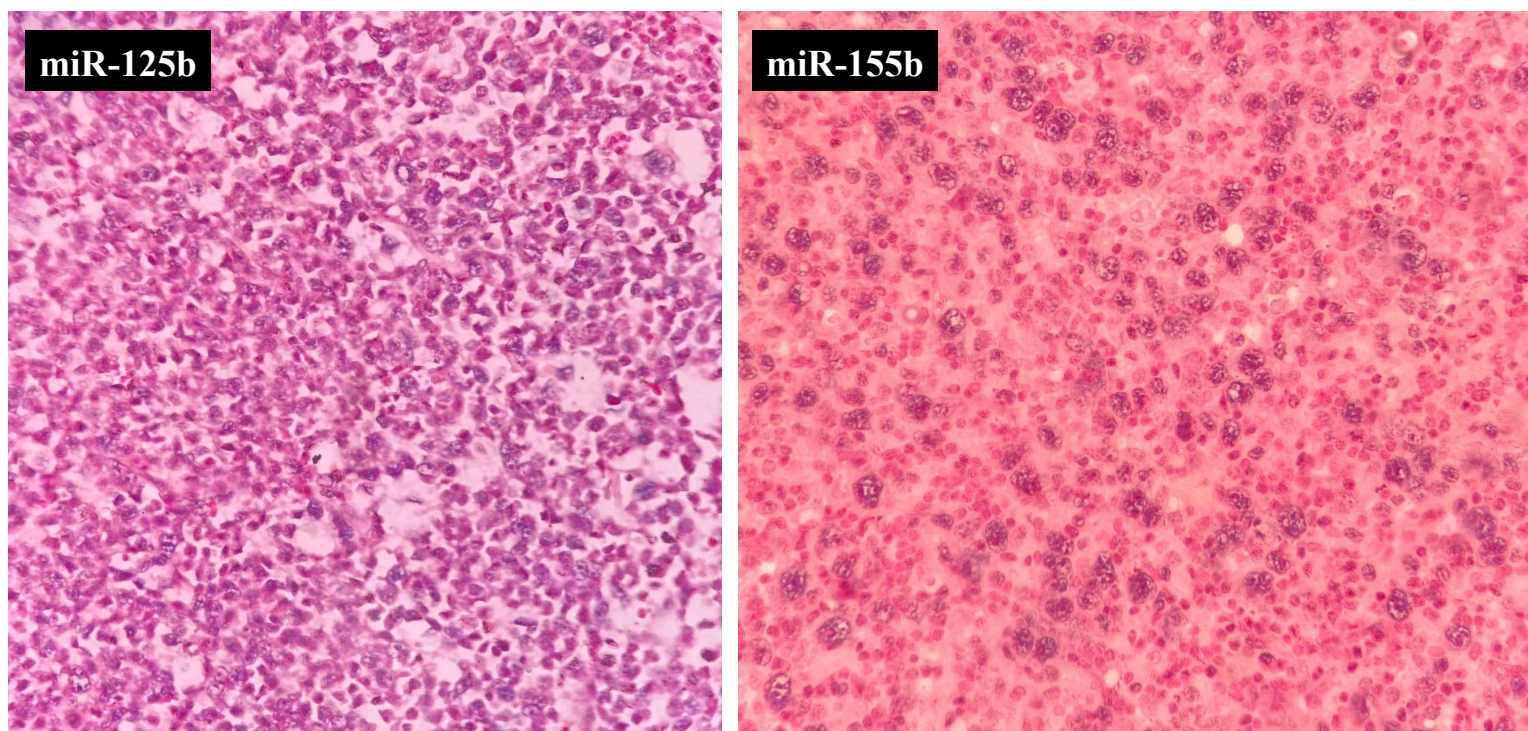
2.3 Fluorescence in-situ hybridization

FISH was performed on 2-μm tissue microarray sections using dual-color break-apart probes (MYC/8q24 and BCL6/3q27) and dual-color fusion translocation probes (BCL2/18q21 and IGH/MYC) (Abbott Laboratories, Des Plaines, IL) as previously described [13]. Cells were hybridized with MYC, BCL2 and BCL6 probes and nuclei were counterstained with 4',6-diamidino-2-phenylindole antifade. Fluorescence was detected with a Olympus BX41 fluorescence microscope (Olympus™, Japan) with excitation filters for 4',6-diamidino-2-phenylindole (260 nm), and rhodamine (570 nm). For each case, up to 200 interphase nuclei were analyzed using an ASI image analysis system (Applied Spectral Imaging™, Israel). The spots in nuclei were scored as previously described [13].

2.4 miRNA in-situ hybridization

For miR-125b and miR-155b we performed ISH using specific LNA probes (hsa-miR-125b-5p miRCURY and hsa-miR-155b-5p miRCURY LNA miRNA detection probes, respectively, at 40nM for both probes (Exiqon™, Vedbæk, Denmark) as previously described [14]. Briefly, 2-µm tissue microarray sections were submitted to a proteinase-K treatment at 15 mg/mL for 10 minutes and hybridization at 53°C for 90 minutes with the aforementioned DIG-labeled locked nucleic acid probes. Stringent washes were performed at 53°C and room temperature for both probes using SSC buffers. All slides were incubated with a DIG-blocking reagent (Roche, Mannheim, Germany), followed by alkaline phosphatase-conjugated anti-DIG (Roche), and NBT-BCIP substrate for 1 hour to elicit the blue ISH signal. All sections were counterstained with nuclear fast red (Vector Labs., Burlingame, CA) and mounted. Non-neoplastic lymphoid tissue (tonsil) incubated with U6 snRNA probe and a scramble probe were used as positive and negative controls, respectively. The cutoff for positivity was $\geq 30\%$ of blue-stained neoplastic cells in contrast to the red colored background as previously described [14] (Figure 1). When positive, the stain intensity was classified as weak, moderate and intense and the Allred score was calculated accordingly [15].

Figure 1 - ISH representative case of positive miR-125b and miR-155b, highlighting



blue-stained neoplastic cells in contrast to the red colored inflammatory background

2.5 Statistical analyses

Data was analyzed with SPSS version 17.0 (Chicago, IL, USA). Statistically significant differences were evaluated by the chi-square or Fisher exact test. Correlations between miRNA continuous variables such as positive cells percentage, Allred score and staining intensity and clinicopathological features were analyzed by Spearman's rank correlation coefficient. Overall survival (OS) was calculated from the date of diagnosis until whether death from any cause or the last follow-up appointment date. Survival rates were plotted in Kaplan-Meier curves and compared by the log-rank test. Univariate and multivariate analyses were performed using the Cox proportional hazard regression model. A $p < 0.05$ was considered significant.

3. RESULTS

3.1 Study population

Among the 139 cases, 52.5% (73/139) were males. The median age was 61.0 years, ranging from 21 to 98. Most of the patients had nodal disease (64.0%, 89/139) and, among those with extranodal lymphoma, the gastrointestinal tract was the most common site affected (34%, 17/50). When the cell-of-origin classification according to Hans' algorithm was applied, the germinal center subtype (GC-DLBCL) was slightly more frequent (53.9%, 75/139) than the non-germinal center subtype (nGC-DLBCL) (46.1%, 64/139).

3.2 *MYC*, *BCL2* and *TP53* expression, *MYC*, *BCL2* and *BCL6* translocations status and miR-125b and miR-155b detection

3.2.1 *Distribution by cell-of-origin classification*

The data of the biomarker's distribution according to cell-of-origin classification are summarized in Table 1. In the nGC group, *MYC* expression was significantly more frequent ($p=0.044$), along with *MYC/TP53* double expression ($p=0.048$). Interestingly, *BCL2* expression was associated to the presence of *BCL2* chromosomal rearrangement only in this subset of cases ($p=0.005$). Conversely, *BCL6* translocation ($p=0.018$) was almost exclusively present in the GC subgroup. Association of *MYC* immunoexpression with *MYC* translocations were observed in both subgroups ($p<0.001$ for the nGC and $p=0.014$ for the GC).

Among the 139 samples, 135 cases were successfully available for microRNA ISH detection, being miR-125b detected in 58.5% (79/135) and miR-155b in only 24.4% (33/135). No statistical difference was observed in the cell-of-origin subgroups for neither miRs detection. In the nGC patients, miR-125b detection was positively correlated to the Ki-67 index ($r=0.267$; $p=0.035$). Considering the GC subgroup, besides a miR-125b association to Ki-67>70% ($p=0.043$) (supplementary table), the percentage of miR-125b positive cells was also correlated to either *MYC* and *MYC/BCL2* double expression ($r=0.234$; $p=0.047$ and $r=0.234$; $p=0.049$, respectively) when we analyzed by Spearman's rank coefficient. No associations were observed between miR-125b and the studied chromosomal rearrangements, nor between miR-155b and any of these parameters. Supplementary table with biomarkers distribution and clinical features according to miR-125b and miR-155b positive *versus* negative status is available.

In a previous study, we described different association patterns among EZH2, MYC and TP53 depending on the cell-of-origin classification, as being an EZH2/MYC/TP53 pathway a nGC feature and, conversely, an EZH2/TP53 pathway a GC characteristic [4]. Despite the small number of studied cases, we analyzed miR-125b and miR-155b distribution according to these different pathways. In the EZH2/MYC/TP53 patients, positivity for miR-125b was observed in 66.6% (4/6) of the cases, while for miR-155b in 33.3% (2/6). In the EZH2/TP53 patients (GC subgroup), 45.4% (5/11) were positive for miR-125b and 27.3% (3/11) for miR-155b expression. Considering the MYC/TP53 cases, miR-125b was detected in 75.0% (6/8) and miR-155b in only 25.0% (2/8). No statistically significant associations were found among any of these groups.

Table 1 - GC versus nGC classification and biomarkers detection

	GC (53.9%, 75/139)	nGC (46.1%, 64/139)	<i>p</i>
Age (median)	58.3 (21-98)	61.7 (22-90)	0.142
Nodal disease	50 (66.7%)	39 (61.0%)	0.483
MYC expression	4 (5.3%)	10 (15.6%)	0.044*
BCL2 expression	51 (68.0%)	43 (67.2%)	0.919
BCL6 expression	69 (92.0%)	31 (48.4%)	0
TP53 expression	23 (30.7%)	22 (34.4%)	0.641
Ki-67 index > 70%	45 (60.0%)	48 (75.0%)	0.061

	GC (53.9%, 75/139)	nGC (46.1%, 64/139)	p
MYC/BCL2 expres- sion	3 (4.0%)	6 (9.4%)	0.967
MYC/TP53 expres- sion	2 (2.7%)	7 (11.1%)	0.048*
MYC translocation	18 (24.0%)	12 (18.7%)	0.453
IgH/MYC translocation	4 (5.3%)	1 (1.6%)	0.373
BCL2 translocation	10 (13.3%)	13 (20.3%)	0.270
BCL6 translocation	9 (12.0%)	1 (1.6%)	0.018*
Double translocation	8 (10.7%)	6 (9.4%)	0.801
miR-125b detection	39 (54.2%)	40 (63.5%)	0.243
miR-155b detection	15 (20.5%)	17 (27.0%)	0.402

3.2.2 *Clinicopathological associations*

LDH levels, ECOG status, presence of B symptoms, Bulky mass, extranodal disease and staging were evaluated as clinical features in association to the studied biomarkers. In the GC subgroup, miR-155b detection was significantly associated to the absence of B symptoms ($p= 0.006$) (supplementary table). In fact, both the percentage and the intensity of miR-155b staining as well as Allred score were negatively correlated to the presence of B symptoms ($r= -0.377$; $p= 0.002$, $r= -0.333$; $p= 0.0060$ and $r= -0.358$; $p= 0.003$, respectively) when we analyzed by Spearman's rank coefficient. Similar results were observed for the ECOG status regarding to the same

staining parameters for miR-155b using Spearman's rank coefficient ($r = -0.404$; $p = 0.021$, $r = -0.406$; $p = 0.006$, $r = -0.384$; $p = 0.009$ and $r = -0.394$; $p = 0.007$, respectively).

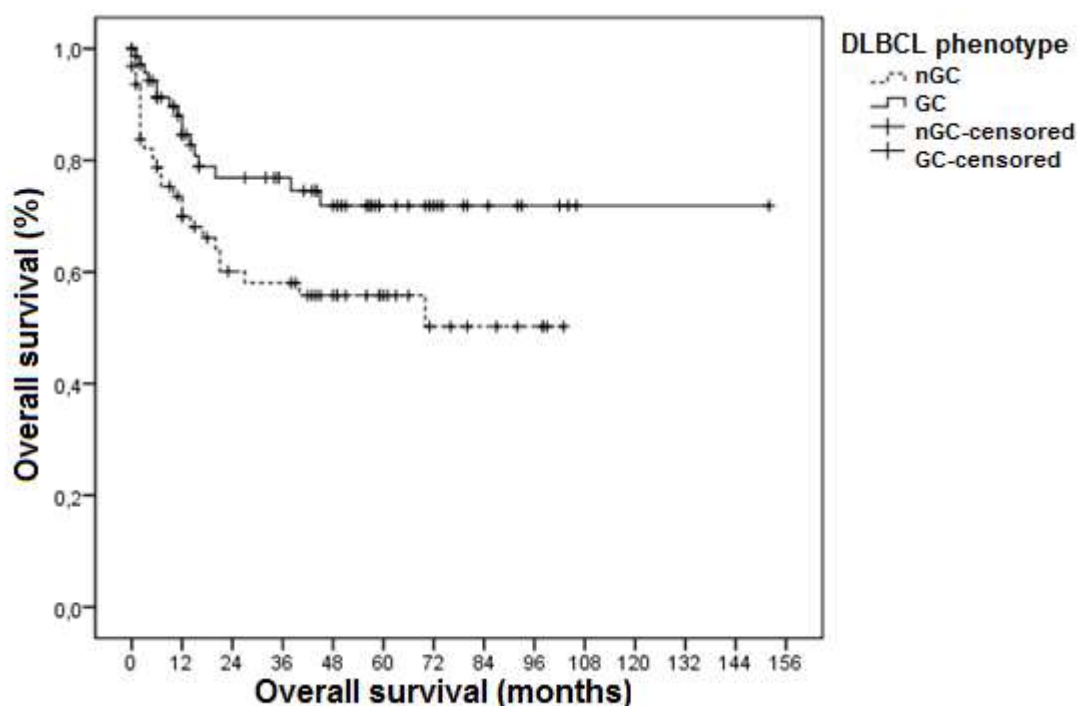
In the nGC subgroup, miR-125b detection was significantly associated to nodal disease ($p = 0.009$) by the chi-square test, and miR-125b percentage and Allred score were also correlated to nodal disease ($r = 0.303$; $p = 0.016$ and $r = 0.266$; $p = 0.035$, respectively) when we applied the Spearman's rank coefficient. TP53 expression showed association with B symptoms ($p = 0.031$) and advanced ECOG status (2-4) ($p = 0.044$), while BCL2 was associated to the presence of bulky mass. Neither MYC expression nor the presence of any of chromosomic rearrangements were statistically associated to these features. Regarding the presenting age of patients, BCL2 expression was consistently higher in those older than 60, either in the GC and nGC subgroups ($p = 0.015$ and $p = 0.026$, respectively). Supplementary table with clinical features according to miR-125b and miR-155b positive *versus* negative status is also provided.

3.2.3 *Primary response to R-CHOP and prognostic analyses*

Biomarkers detection was also evaluated according to the primary response to R-CHOP based therapy, which we were able to analyze in 92 patients. In nGC patients, miR-155b percentage and intensity, as well as Allred score, were positively correlated to progressive disease (PD) ($r = 0.317$; $p = 0.038$, $r = 0.293$; $p = 0.057$ and $r = 0.317$; $p = 0.039$, respectively). Additionally, association between BCL2 expression with either stable disease (SD) and stable disease plus progressive disease (SD+PD) was observed ($p = 0.018$ and $p = 0.016$, respectively).

Kaplan-Meier survival curves for OS rates for both GC and nGC subtypes are outlined in Figure 2. OS rates of the GC and nGC patients were 78.1% and 58.7%, respectively, with nGC-DLBCL having poorer OS rates compared with GC-DLBCL ($p = 0.023$).

Figure 2 - Survival curves for OS rates with respect to the GC (n=49) and nGC (n=43) status



3.2.4 Univariate analyses of prognostic factors for OS

Univariate Cox proportional hazard regression analysis was used to assess the prognostic factors for OS, as described in table 2. In the general sample, the GC phenotype showed a significant positive prognostic impact. Considering cell-of-origin classification, in the nGC-DLBCL group we found that age, Ann Arbor stage, extranodal involvement, LDH level, ECOG performance status and bulky mass were significant negative prognostic factors for OS in our patients cohort. However, in the GC subgroup, only extranodal involvement, stage and the presence of B symptoms were negatively significant. In addition, MYC expression, MYC(+)/TP53(+) double-expression, and MYC(+)/BCL2(+)/TP53(+) triple-expression were consistently associated to poorer OS rates in GC-DLBCL. No significant OS rates comparing miR-125b and miR-155b detection was observed in any subgroup.

Table 2 - Prognostic factors that affect OS (univariate analysis)

Risk Factor	GC		nGC	
	RR (95% CI)	<i>p</i>	RR (95% CI)	<i>p</i>

Age (>60 vs ≤60)	78.24 (60.05-96.42)	0.906	85.39 (71.51-99.28)	<0.001
Stage (I-II vs III-IV)	90.18 (78.91-101.45)	0.048	72.86 (57.49-88.22)	0.039
Extranodal disease (>1 vs ≤1)	32.43 (7.40-57.46)	0.010	11.07 (2.02-20.12)	0.001
LDH level (elevated vs normal)	109.21 (83.79-134.63)	0.160	44.48 (27.98-60.98)	0.028
ECOG PS (≥2 vs <2)	60.92 (34.08-87.77)	0.052	34.89 (14.93-54.85)	0.003
MYC (positive vs negative)	21.00 (3.37-38.63)	0.037	54.06 (23.88-84.24)	0.577
BCL2 (positive vs negative)	123.52 (105.61-141.43)	0.108	45.30 (31.38-59.22)	0.003
TP53 (positive vs negative)	78.01 (54.61-101.41)	0.883	56.86 (35.83-77.89)	0.501
MYC/BCL2 (positive vs negative)	44.60 (15.15-74.05)	0.334	28.70 (4.15-53.25)	0.422
MYC/TP53 (positive vs negative)	11.00 (11.00-11.00)	0.013	61.57 (26.07-97.08)	0.990
MYC/BCL2/TP53 (positive vs negative)	11.00 (11.00-11.00)	0.013	15.50 (0.00-31.52)	0.090

MYC translocation (positive vs negative)	73.82 (50.46-97.19)	0.310	45.85 (29.78-61.91)	0.560
BCL2 translocation (positive vs negative)	45.50 (21.37-69.63)	0.303	34.63(17.84-51.43)	0.531
BCL6 translocation (positive vs negative)	47.80 (29.61-65.99)	0.854	0	0.439
Double-hit translocation (positive vs negative)	52.43 (29.48-75.37)	0.592	36.17 (12.97-59.36)	0.742
miR-125b (positive vs negative)	107.36 (82.68-132.04)	0.472	66.94 (51.94-81.95)	0.341
miR-155b (positive vs negative)	86.13 (75.02-97.24)	0.156	50.41 (33.98-66.85)	0.850

3.2.5 Multivariate analyses of prognostic factors for OS

The clinicopathological features and biomarkers presence were also included in a multivariate analysis of survival, as described in table 3. In this analysis, GC phenotype was a significant independent factor associated to higher OS (HR= 0.14; CI 95% 0.04 - 0.6; $p=0.007$). Considering the nGC group, Ann Arbor stage, extranodal involvement and ECOG PS were consistently independent factors related to lower OS. Interestingly, although not significant, the expression of TP53, miR-125b and miR-155b seems to be potential prognostic biomarkers in these tumors. No significant OS rates comparing any variables was observed in the GC subset.

Table 3 - Prognostic factors that affect OS (multivariate analysis)

Risk Factor	GC		nGC	
	HR (95% CI)	<i>p</i>	HR (95% CI)	<i>p</i>
Age (>60 vs ≤60)	12.10 (0.27-53.27)	0.197	1.08 (0.95-1.23)	0.235
Stage (I-II vs III-IV)	1.45 (0.00-1.51)	0.219	4.67 (1.11-19.71)	0.036
Extranodal disease (>1 vs ≤1)	6.41 (0.00-1.03)	0.191	4.13 (1.91-8.92)	0.028
LDH level (elevated vs normal)	4.67 (0.00-8.27)	0.180	0.48 (0.04-5.65)	0.561
ECOG PS (>1 vs ≤2)	1.00 (0.00-22.15)	0.219	13.55 (2.22-82.80)	0.005
MYC (positive vs negative)	1.51 (0.00-15.70)	0.915	3.60 (0.00-1.55)	0.900
BCL2 (positive vs negative)	1.00 (0.00-10.99)	0.175	16.78 (0.45-622.19)	0.126
TP53 (positive vs negative)	17.80 (0.00-28.29)	0.790	2.61 (0.90-7.60)	0.078
MYC translocation (positive vs negative)	2.30 (0.00-1.61)	0.527	1.00 (0.00-3.94)	0.906
BCL2 translocation (positive vs negative)	9.10 (0.00-3.65)	0.866	0.05 (0.00-2.72)	0.144

BCL6 translocation (positive vs negative)	6.16 (0.00-3.04)	0.755	0.00 (0.00-9.62)	0.965
Double-hit translocation (positive vs negative)	1.00 (0.00-10.98)	0.797	2.85 (0.14-5.96)	0.147
miR-125b (positive vs negative)	1.13 (0.78-1.62)	0.525	0.95 (0.91-1.00)	0.065
miR-155b (positive vs negative)	0.84 (0.43-1.66)	0.620	1.03 (1.00-1.07)	0.084

4. DISCUSSION

The DLBCL categorization in GC and nGC subgroups has clinical relevance as it implies in different carcinogenic pathways and potential prognostic impact. In order to better understand the interaction of some DLBCL most studied biomarkers and their impact on DLBCL outcome, we focused our analyses on distinguishing the main features of GC from nGC phenotypes.

4.1 Differential detection of the studied biomarkers according to cell-of-origin phenotype and their interactions

Our results showed that MYC expression was significantly associated to the nGC group, corroborating with Wight et al. findings in which MYC overexpression was a nGC feature [16]. In nGC DLBCL patients, the mechanism described in which MYC is overexpressed is mainly BCR and NF- κ B induced, while in the GC subtype they are caused by the translocation of MYC [17]. However, in this study MYC chromosomal translocation was nearly equivalent in both subtypes, with MYC expression consistently associated to its translocation, suggesting that MYC translocation, in our series, is a molecular mechanism that leads to MYC expression regardless the cell-of-origin. In addition, among the 30 cases with translocated MYC, only 14 were positive

for MYC expression, which highlights that *MYC* translocation itself is not always sufficient to result in its own overexpression. Copie-Bergman *et al* investigated *MYC* translocation partners (immunoglobulin loci - Ig vs. non-Ig) in a large series of DLBCL and found that high MYC protein expression was indeed significantly more frequent in the MYC-Ig subgroup compared to MYC-non-Ig DLBCL [18]. So that, we next analyzed among the 30 *MYC* rearranged patients which cases carried IgH/*MYC* translocation using a specific probe. Interestingly, only one out of all five IgH/*MYC* positive cases showed MYC expression. Studying Burkitt lymphoma, Wilda *et al.* analyzed the influence of breakpoint locations within the *MYC* gene on MYC transcript levels and demonstrated that the highest MYC mRNA levels occur when MYC expression is directly triggered via its cryptic P3 promoter within the first intron, which may explain why although our cases show MYC rearrangements, even IgH/*MYC* translocations, and not necessarily result on MYC expression [19]. Although we have not studied MYC translocation partners in this study, the presence of a great number of non-Ig partners may explain our observations. Interestingly, despite the small differences in the *BCL2* translocation frequency between GC-DLBCL (13%) and nGC-DLBCL (20%), only nGC tumors demonstrated to have a relationship between this translocation and *BCL2* expression. As well as MYC expression, *BCL2* expression was not caused by translocation in most cases, suggesting that other mechanisms might be involved. We have also found that the MYC/*TP53* double expression was more frequent in the nGC subtype. Several studies have shown that MYC overexpression *per se* is insufficient to promote neoplastic growth as it induces apoptosis via *TP53*-related pathway [20, 21]. However, as *TP53* expression is highly associated to mutated *TP53*, its expression may enhance adverse effects of MYC overexpression on DLBCL as demonstrated by some authors [21, 22, 23], indicating that MYC/*TP53* double expression is also a nGC feature.

It has been shown that *BCL6* is frequently affected by chromosomal translocations, occurring more frequently in nGC than in GC DLBCL and leading to a deregulated *BCL6* expression by preventing its silencing at the conclusion of the germinal center response [24]. Conversely, In our series, *BCL6* translocation was almost exclusively observed in the GC subgroup. That is interesting as in this subgroup *BCL6* expression was supposed to be positive as a result of Hans algorithm itself, which may suggest that in our cases, chromosomal translocation accounted for such overexpression. *BCL6* translocation involves several non-Ig partners, whose expression

may be regulated by some BCL6 regulatory regions, and the expression of some of these partner genes may influence the biology of the DLBCL [25]. We speculate that in our cases differential translocation partners may lead to BCL6 overexpression without activating a complete post germinal center genetic machinery, keeping the lymphoma cell-of-origin with a GC phenotype.

When it comes to gene expression, miRNA demonstrated to have an important role in DLBCL pathogenesis. miR-125b and miR-155b have been described to be crucial in regulating important pathogenetic pathways, especially involving *MYC* and *TP53*, which may lead to cell proliferation, disruption of apoptosis and DNA repair mechanisms [6, 10]. In this study, although neither miR-125b nor miR155b were related to any DLBCL subtype, it is noteworthy that miR-125b was associated to high proliferation index independently on cell-of-origin subgroups. The ectopic expression of miR-125b in B-cell lymphoma results in an enhanced NF- κ B activity, thereby facilitating cell proliferation and cancer progression in malignant B-cell lymphoma [5, 7]. Likewise, Kim et al demonstrated that miR-125b constitutively activates NF- κ B pathway, regardless of the molecular subtype, which may promote tumor cell proliferation and regulate apoptosis [26]. In fact, for both, GC and nGC DLBCL, miR-125b was positively correlated to Ki-67. As a GC characteristic, miR-125b was associated to either *MYC* or *MYC/BCL2* expression. As a matter of fact, Manfè et al. suggest a mechanism in cancer cells in which miR-125b targets the *MYC* antagonist *MAD4* [27]. In addition, miR-125b is likely to up-regulate Wnt/ β -catenin activity by down-regulating β -catenin degradation targets, thereby allowing more downstream targets such *MYC* to be expressed [5].

4.2 Clinical impact of the studied biomarkers according to cell-of-origin phenotype

We also analyzed clinical features and their associations with the studied biomarkers considering the cell-of-origin classification. It is interesting to note that miR-155b is associated to the absence of B symptoms and to a lower ECOG status in our GC subgroup. In contrast, a Turkish study found that higher levels of miRNA-155 in patients were associated to the presence of B symptoms, involvement of extranodal sites, and high ECOG performance scores in DLBCL [28]. In a mantle-cell lymphoma study, however, He et al. found no associations between miR-155b and these parameters at all [29]. Both studies did not consider GC *versus* nGC classification on

their analyses, and they used RT-PCR to evaluate miR-155b expression rather than ISH, which may explain our conflicting findings. Considering the nGC subgroup, we found that mi-125b expression was consistently associated to nodal disease, as opposed to Zhan et al., who found no association between miR-125b expression and clinical parameters [30].

With regards to nGC-DLBCL, we found that TP53 expression was associated to B-symptoms and advanced ECOG status. That is in accordance with a Russian cohort of *de novo* DLBCL patients, whose B-symptoms, splenomegaly and bone marrow involvement were TP53 mutation correlated [31]. However, in this study, the authors have not stratified their patients by cell-of-origin classification. BCL2 expression, on the other hand, seems to be associated to higher disease onset age regardless of DLBCL molecular subtype. Again, this does not seem to be a general observation, as Iqbal et al., also considering cell-of-origin classification, has not found such result [32].

Next, we evaluated clinical outcomes of each biomarker according to molecular subtype. As presumed, the GC phenotype showed an independent positive prognostic impact as highlighted by the multivariate analysis. Most DLBCL recapitulate the differentiation and maturation mechanisms observed in the two main subtypes previously defined by GEP: the GCB DLBCL subtype has a GEP related to a GC cell of origin, with gene mutation instrumental for GC development, GC dark and light zone transitions and microenvironmental interactions, whereas the ABC subtype, which derives from cells of GC exit or post GC origin, with either germinal center-exit or early plasmablastic phenotype, and is characterized by being BCR and NF κ B dependent. In fact, although Hans algorithm may present concordance issues with GEP, it has being widely used in routine practice and is recommended by the World Health Organization on its last edition [2, 33].

Interestingly, when analyzing primary response to R-CHOP scheme, we found that miR-155b expression was associated to progressive disease in the nGC subgroup. This suggests that in this subgroup, miR-155b is more likely to indicate a poor prognostic feature. Although it was not statistically significant, its borderline *p*-value in multivariate analyses may be interpreted as a potential prognostic marker as result of our limited sample size. Quantifying miR-155b plasmatic levels by RT-PCR, Ahmadvand et al. found a negative impact of miR-155b high levels on DLBCL clinical outcomes, however they did not consider molecular subgroups on their analyses [8].

Similarly, Zhong et al. found that low miR-155b levels were related to complete response to R-CHOP therapy and, conversely, miR-155b was an independent prognostic factor in DLBCL [34]. Nevertheless, this observation is heterogeneous in literature. Contradictorily, Jung and Aguiar observed that high expression of miR-155 was associated to improved prognosis exclusively within the ABC subgroup [35]. More studies, with a large sample size, are needed to confirm the prognostic impact of miR-155b in DLBCL and in its subtypes.

On the other hand, also considering nGC cases, miR-125b tended to be an independent positive prognostic factor. miR-125b signalling has an important functional significance in the regulation of lymphoma tumor development and resistance of lymphoma cells to chemotherapy due its interactions with different pathways [36], but the literature is scarce when it comes to evaluate miR-125b as a potential prognostic marker in DLBCL [37, 38]. Yuan et al. found that high levels of miR-125b were associated to R-CHOP resistance and therefore to poor prognosis, as patients with higher miR-125b levels had a shorter overall survival [39]. Nevertheless, they analyzed serum miRNA rather than direct tumor miRNA and they did not stratify their cases in DLBCL subtypes, which may explain our conflicting results. As previously discussed, we found that miR-125b is associated to MYC only among GC-DLBCL, which suggests that miR-125b role in fact vary upon DLBCL subtype. It would be more accurate to evaluate miR-125b expression as a prognostic marker considering GC and nGC subgroups as different diseases rather than DLBCL as a single entity. Our analyses are limited due a small number of patients and further investigation is needed.

Also, in the nGC subgroup TP53 expression showed a trend of negative prognostic factor in the multivariate analyses. The TP53 protein is so-called the "genome guardian", a central tumor suppressor that mediates cell-cycle arrest, DNA repair, apoptosis and senescence. Alterations in the *TP53* gene may disrupt the protein functions and has been implicated on lymphomagenesis and disease progression [21, 22]. Although *TP53* genetic alterations have not been evaluated in our study, TP53 immunoexpression has been strongly correlated to *TP53* mutations in DLBCL [40, 41], as mutated *TP53* leads to protein stability in comparison to the shorter half-life wild type form. For instance, a classic study leaded by Xu-Monette et al. demonstrated that *TP53* mutations predict poor survival rates in all DLBCL patients. Albeit both molecular subtypes OS were affected by TP53 mutation, ABC-DLBCL patients treated with R-CHOP have significantly inferior OS and PFS com-

pared to GCB-DLBCL patients both in the entire DLBCL cohort [40]. As a matter of fact, the distinctive molecular programs harbored by GCB- and ABC-DLBCL might explain the differences in treatment response and in the impact of *TP53* mutations. Interestingly, Peroja et al found that overall *TP53* mutations was not of prognostic impact, but specific mutations involving the DNA-binding domain were associated to a shorter survival in DLBCL [41]. We have previously demonstrated that *TP53* overexpression may reflect specific *TP53* mutations in gliomas [42] and here we hypothesize that in the nGC-DLBCL the genetic alterations culminating in *TP53* immunoexpression are crucial to derange protein functions and to indicate worse survival.

In conclusion, the present study validates Hans algorithm as an alternative approach to subclassify DLBCL. We have demonstrated different pathways based on GC *versus* nGC classification and highlighted different clinical outcomes. In addition, miR-125b, miR-155b and *TP53* expression might represent potential prognostic factors in nGC-DLBCL. Further investigation, multi-institutional, with a larger number of subjects are needed to confirm this data and to achieve a better understanding of DLBCL molecular landscape.

ACKNOWLEDGEMENT

This work was supported by the Fundação Cearense de Apoio ao Desenvolvimento Científico e Tecnológico (FUNCAP) [PPSUS 2017].

COMPETITION INTERESTS

None.

ETHICS APPROVAL STATEMENT

The present study was approved by the hospital ethics committee of the Instituto do Câncer do Ceará under reference approval number 66308017.8.0000.5528.

CONTRIBUTORSHIP STATEMENT

All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by EHCNF, SBZ, PGBS, RMRB and SHBR. The first draft of the manuscript was written by EHCNF and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

REFERENCES

1. Ma XB, Zheng Y, Yuan HP, Jiang J, Wang YP. CD43 expression in diffuse large B-cell lymphoma, not otherwise specified: CD43 is a marker of adverse prognosis. *Human pathology*. 2015 Apr 1;46(4):593-9.
2. Alaggio R, Amador C, Anagnostopoulos I, Attygalle AD, Araujo IB, Berti E, Bhagat G, Borges AM, Boyer D, Calaminici M, Chadburn A. The 5th edition of the World Health Organization classification of haematolymphoid tumours: lymphoid neoplasms. *Leukemia*. 2022 Jul;36(7):1720-48.
3. Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delabie J, Ott G, Muller-Hermelink HK, Campo E, Braziel RM, Jaffe ES, Pan Z. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. *Blood*. 2004 Jan 1;103(1):275-82.
4. Neves Filho EH, Hirth CG, Frederico IA, Burbano RM, Carneiro T, Rabenhorst SH. EZH2 expression is dependent on MYC and TP53 regulation in diffuse large B - cell lymphoma. *Apmis*. 2020 Apr;128(4):308-15.
5. Peng B, Theng PY, Le MT. Essential functions of miR - 125b in cancer. *Cell proliferation*. 2021 Feb;54(2):e12913.
6. Larrabeiti-Etxebarria A, Lopez-Santillan M, Santos-Zorrozuza B, Lopez-Lopez E, Garcia-Orad A. Systematic review of the potential of microRNAs in diffuse large B cell lymphoma. *Cancers*. 2019 Jan 26;11(2):144.
7. Zheng Z, Qu JQ, Yi HM, Ye X, Huang W, Xiao T, Li JY, Wang YY, Feng J, Zhu JF, Lu SS. MiR-125b regulates proliferation and apoptosis of nasopharyngeal carcinoma by targeting A20/NF- κ B signaling pathway. *Cell death & disease*. 2017 Jun;8(6):e2855-.
8. Ahmadvand M, Eskandari M, Pashaiefar H, Yaghmaie M, Manoochehrabadi S, Khakpour G, Sheikhsaran F, Zohour MM. Over expression of circulating miR-155 predicts prognosis in diffuse large B-cell lymphoma. *Leukemia research*. 2018 Jul 1;70:45-8.
9. Bento L, Vögler O, Sas-Barbeito A, Muncunill J, Ros T, Martínez J, Quintero-Duarte A, Ramos R, Asensio VJ, Fernández-Rodríguez C, Salar A. Screening for Prognostic microRNAs Associated with Treatment Failure in Diffuse Large B Cell Lymphoma. *Cancers*. 2022 Feb 20;14(4):1065.
10. Shi Y, Ding D, Qu R, Tang Y, Hao S. Non-coding RNAs in diffuse large B-cell lymphoma. *OncoTargets and therapy*. 2020;13:12097.

11. Cheson BD. The International Harmonization Project for response criteria in lymphoma clinical trials. *Hematol Oncol Clin North Am.* 2007 Oct;21(5):841-54.
12. Fedchenko N, Reifenrath J. Different approaches for interpretation and reporting of immunohistochemistry analysis results in the bone tissue - a review. *Diagn Pathol.* 2014 Nov 29;9:221.
13. Horn H, Ziepert M, Becher C, et al. MYC status in concert with BCL2 and BCL6 expression predicts outcome in diffuse large B-cell lymphoma. *Blood.* 2013 Mar 21;121(12):2253-63.
14. Munch-Petersen HD, Ralfkiaer U, Sjö LD, Hother C, Asmar F, Nielsen BS, Brown P, Ralfkiaer E, Grønbæk K. Differential expression of miR-155 and miR-21 in tumor and stroma cells in diffuse large B-cell lymphoma. *Applied Immunohistochemistry & Molecular Morphology.* 2015 Mar 1;23(3):188-95.
15. Allred DC. Problems and solutions in the evaluation of hormone receptors in breast cancer. *Journal of Clinical Oncology.* 2008 May 20;26(15):2433-5.
16. Wight JC, Chong G, Grigg AP, Hawkes EA. Prognostication of diffuse large B-cell lymphoma in the molecular era: moving beyond the IPI. *Blood reviews.* 2018 Sep 1;32(5):400-15.
17. Xia Y, Zhang X. The spectrum of MYC alterations in diffuse large B-cell lymphoma. *Acta haematologica.* 2020;143(6):520-8.
18. Copie-Bergman C, Cuillière-Dartigues P, Baia M, Briere J, Delarue R, Canioni D, Salles G, Parrens M, Belhadj K, Fabiani B, Recher C. MYC-IG rearrangements are negative predictors of survival in DLBCL patients treated with immunochemotherapy: a GELA/LYSA study. *Blood, The Journal of the American Society of Hematology.* 2015 Nov 26;126(22):2466-74.
19. Wilda M, Busch K, Klose I, Keller T, Woessmann W, Kreuder J, Harbott J, Borkhardt A. Level of MYC overexpression in pediatric Burkitt's lymphoma is strongly dependent on genomic breakpoint location within the MYC locus. *Genes, Chromosomes and Cancer.* 2004 Oct;41(2):178-82.
20. Deng M, Xu-Monette ZY, Pham LV, Wang X, Tzankov A, Fang X, Zhu F, Visco C, Bhagat G, Dybkaer K, Chiu A. Aggressive B-cell lymphoma with MYC/TP53 dual alterations displays distinct clinicopathobiological features and response to novel targeted agents. *Molecular Cancer Research.* 2021 Feb;19(2):249-60.
21. Bouroumeau A, Bussot L, Bonnefoix T, Fournier C, Chapusot C, Casasnovas O, Martin L, McLeer A, Col E, David-Boudet L, Lefebvre C. c-MYC and p53 ex-

- pression highlight starry-sky pattern as a favourable prognostic feature in R-CHOP-treated diffuse large B-cell lymphoma. *The Journal of Pathology: Clinical Research*. 2021 Nov;7(6):604-15.
22. Wang XJ, Jeffrey Medeiros L, Bueso-Ramos CE, Tang G, Wang S, Oki Y, Desai P, Khoury JD, Miranda RN, Tang Z, Reddy N. P53 expression correlates with poorer survival and augments the negative prognostic effect of MYC rearrangement, expression or concurrent MYC/BCL2 expression in diffuse large B-cell lymphoma. *Modern Pathology*. 2017 Feb;30(2):194-203.
 23. Xie Y, Bulbul MA, Ji L, Inouye CM, Groshen SG, Tulpule A, O'Malley DP, Wang E, Siddiqi IN. p53 expression is a strong marker of inferior survival in de novo diffuse large B-cell lymphoma and may have enhanced negative effect with MYC coexpression: a single institutional clinicopathologic study. *American journal of clinical pathology*. 2014 Apr 1;141(4):593-604.
 24. Testoni M, Zucca E, Young KH, Bertoni F. Genetic lesions in diffuse large B-cell lymphomas. *Annals of Oncology*. 2015 Jun 1;26(6):1069-80.
 25. Iqbal J, Greiner TC, Patel K, Dave BJ, Smith L, Ji J, Wright G, Sanger WG, Pickering DL, Jain S, Horsman DE. Distinctive patterns of BCL6 molecular alterations and their functional consequences in different subgroups of diffuse large B-cell lymphoma. *Leukemia*. 2007 Nov;21(11):2332-43.
 26. Kim SW, Ramasamy K, Bouamar H, Lin AP, Jiang D, Aguiar RC. MicroRNAs miR-125a and miR-125b constitutively activate the NF- κ B pathway by targeting the tumor necrosis factor alpha-induced protein 3 (TNFAIP3, A20). *Proceedings of the National Academy of Sciences*. 2012 May 15;109(20):7865-70.
 27. Manfe V, Biskup E, Willumsgaard A, Skov AG, Palmieri D, Gasparini P, Lagana A, Woetmann A, Ødum N, Croce CM, Gniadecki R. cMyc/miR-125b-5p signalling determines sensitivity to bortezomib in preclinical model of cutaneous T-cell lymphomas. *PloS one*. 2013 Mar 19;8(3):e59390.
 28. Bedewy AM, Elmaghraby SM, Shehata AA, Kandil NS. Prognostic value of miRNA-155 expression in B-cell non-Hodgkin lymphoma. *Turkish Journal of Hematology*. 2017 Sep;34(3):207.
 29. He J, Xi Y, Gao N, Xu E, Chang J, Liu J. Identification of miRNA-34a and miRNA-155 as prognostic markers for mantle cell lymphoma. *Journal of International Medical Research*. 2021 May;49(5):03000605211016390.

30. Zhan C, Wang T, You H, Si C. Different expressions of miR-125b and SOX30 in malignant lymphomas and their significance. *J BUON*. 2018 Jul 1;23(4):1179-84.
31. Voropaeva EN, Pospelova TI, Voevoda MI, Maksimov VN, Orlov YL, Seregina OB. Clinical aspects of TP53 gene inactivation in diffuse large B-cell lymphoma. *BMC medical genomics*. 2019 Mar;12:35-44.
32. Iqbal J, Neppalli VT, Wright G, Dave BJ, Horsman DE, Rosenwald A, Lynch J, Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD. BCL2 expression is a prognostic marker for the activated B-cell-like type of diffuse large B-cell lymphoma. *Journal of Clinical Oncology*. 2006 Feb 20;24(6):961-8.
33. Skaftason A, Qu Y, Abdulla M, Nordlund J, Berglund M, Ednersson SB, Andersson PO, Enblad G, Amini RM, Rosenquist R, Mansouri L. Transcriptome sequencing of archived lymphoma specimens is feasible and clinically relevant using exome capture technology. *Genes, Chromosomes and Cancer*. 2022 Jan;61(1):27-36.
34. Zhong H, Xu L, Zhong JH, Xiao F, Liu Q, Huang HH, Chen FY. Clinical and prognostic significance of miR-155 and miR-146a expression levels in formalin-fixed/paraffin-embedded tissue of patients with diffuse large B-cell lymphoma. *Experimental and therapeutic medicine*. 2012 May 1;3(5):763-70.
35. Jung I, Aguiar RC. MicroRNA-155 expression and outcome in diffuse large B-cell lymphoma. *British journal of haematology*. 2009 Jan;144(1):138.
36. Bueno MJ, Gómez de Cedrón M, Gómez-López G, Pérez de Castro I, Di Lisio L, Montes-Moreno S, Martínez N, Guerrero M, Sánchez-Martínez R, Santos J, Pissano DG. Combinatorial effects of microRNAs to suppress the Myc oncogenic pathway. *Blood, The Journal of the American Society of Hematology*. 2011 Jun 9;117(23):6255-66.
37. Yuan WX, Gui YX, Na WN, Chao J, Yang X. Circulating microRNA-125b and microRNA-130a expression profiles predict chemoresistance to R-CHOP in diffuse large B-cell lymphoma patients. *Oncology letters*. 2016 Jan 1;11(1):423-32.
38. Feng Y, Zhong M, Zeng S, Wang L, Liu P, Xiao X, Liu Y. Exosome-derived miRNAs as predictive biomarkers for diffuse large B-cell lymphoma chemotherapy resistance. *Epigenomics*. 2019 Jan;11(1):35-51.
39. Yuan WX, Gui YX, Na WN, Chao J, Yang X. Circulating microRNA-125b and microRNA-130a expression profiles predict chemoresistance to R-CHOP in diffuse large B-cell lymphoma patients. *Oncology letters*. 2016 Jan 1;11(1):423-32.

40. Xu-Monette ZY, Wu L, Visco C, Tai YC, Tzankov A, Liu WM, Montes-Moreno S, Dybkær K, Chiu A, Orazi A, Zu Y. Mutational profile and prognostic significance of TP53 in diffuse large B-cell lymphoma patients treated with R-CHOP: report from an International DLBCL Rituximab-CHOP Consortium Program Study. *Blood, The Journal of the American Society of Hematology*. 2012 Nov 8;120(19):3986-96.
41. Peroja P, Pedersen M, Mantere T, Nørgaard P, Peltonen J, Haapasaari KM, Böhm J, Jantunen E, Turpeenniemi-Hujanen T, Rapakko K, Karihtala P. Mutation of TP53, translocation analysis and immunohistochemical expression of MYC, BCL-2 and BCL-6 in patients with DLBCL treated with R-CHOP. *Scientific Reports*. 2018 Oct 4;8(1):1-9.
42. Faria MH, Neves Filho EH, Alves MK, Burbano RM, de Moraes Filho MO, Rabenhorst SH. TP53 mutations in astrocytic gliomas: an association with histological grade, TP53 codon 72 polymorphism and p53 expression. *Apmis*. 2012 Nov;120(11):882-9.

Supplementary table - biomarkers distribution and clinical features according to miR-125b and miR-155b positivity.

	GC-DLBCL						nGC-DLBCL					
	miR-125b+	miR-125b -	<i>p</i>	miR-155b+	miR-155b -	<i>p</i>	miR-125b+	miR-125b -	<i>p</i>	miR-155b+	miR-155b -	<i>p</i>
Age >60	17	15	0.85	6	27	0.73	23	13	1	9	28	0.78
Age <60	21	15		9	28		16	10		8	18	
Stage I-II	23	16	0.88	10	31	0.18	20	14	0.50	8	26	0.59
Stage III-IV	16	9		3	22		17	7		8	16	
>1 Extranodal disease	3	4	0.69	2	5	0.63	5	2	0.70	3	4	0.38
≤1 Extranodal disease	36	26		13	52		33	21		14	40	
Elevated LDH levels	20	17	0.71	9	28	0.53	23	10	0.39	10	23	1
Normal LDH levels	11	13		4	20		12	9		6	15	
B symptoms +	22	19	0.94	4	36	0,006	20	10	0.84	6	24	0.29

B symptoms -	15	11		11	16		17	11		10	18	
ECOG PS ≥ 2	14	7	0.16	7	21	0.13	14	10	0.54	4	20	0.54
ECOG PS <2	8	12		1	16		15	6		6	15	
MYC +	3	0	0.24	0	3	1	8	2	0.30	2	8	0.71
MYC -	36	33		15	54		32	21		15	38	
BCL2 +	26	23	0.98	11	38	0.76	25	17	0.51	11	31	1
BCL2 -	13	10		4	19		15	6		6	15	
TP53 +	12	9	0.94	4	17	1	15	7	0.77	6	16	1
TP53 -	27	24		11	40		25	16		11	30	
MYC/BCL2 +	2	0	0.49	0	2	1	6	1	0.40	2	4	0.65
MYC/BCL2 -	37	33		15	55		35	22		15	42	
MYC/TP53 +	1	0	1	0	1	1	5	2	1	2	5	1
MYC/TP53 -	38	33		15	56		35	21		15	41	

MYCt +	10	8	1	2	16	0.32	10	2	0.18	3	9	1
MYCt -	29	25		13	41		30	21		14	37	
BCL2t +	6	4	0.74	1	9	0.67	9	5	1	2	11	0.48
BCL2t -	33	29		14	48		32	18		15	35	
BCL6t +	4	6	0.49	1	8	0.67	1	1	1	0	1	1
BCL6t -	35	28		14	49		39	23		17	45	
Double-hit +	4	4	1	1	7	1	4	2	1	0	6	0.17
Double-hit -	35	29		14	50		36	21		17	40	
Ki-67 $\geq$ 70%	28	16	0.04	11	32	0.25	32	15	0.31	12	35	0.90
Ki-67 < 70%	11	17		4	25		8	8		5	11	

Note: chi-square or Fisher exact test was used to compare miRNA positive and negative groups.

CAPÍTULO 3

Artigo: Letter to the Editor - Molecular profile of EBV-DLBCL in a Brazilian population.

Autores: Eduardo Henrique Neves Filho, Silvia Helena Rabenhorst

Ano de confecção: 2023.

LETTER TO THE EDITOR

MOLECULAR PROFILE OF EBV-DLBCL IN A BRAZILIAN POPULATION.

EDUARDO HENRIQUE CUNHA NEVES FILHO and SILVIA HELENA BAREM
RABENHORST

LABGEM - Departamento de Patologia e Medicina Legal - Universidade Federal Do
Ceará - Fortaleza/Brazil.

Corresponding author:

Dr Eduardo H C Neves Filho, MD
Laboratório de Genética Molecular (LABGEM).
Departamento de Patologia e Medicina Legal - Faculdade de Medicina
Universidade Federal do Ceará
Rua Cel. Nunes de Melo, 1315
Campus do Porangabussu CEP 60.183-630
Fortaleza/CE
Brasil
edu0689@yahoo.com.br
Phone: +55 85 981810904

INTRODUCTION

Epstein-Barr virus (EBV)+ diffuse large B-cell lymphoma not otherwise specified (EBV-DLBCL) is a rare entity, especially in Western countries, with only few studies considering the Brazilian population [1, 2]. EBV-DLBCL consists of a group of highly heterogeneous tumors with different clinicopathological characteristics, genetic alterations, responses to therapy and prognosis, in which lack of uniform criteria and heterogeneous geographic distribution may account for differences reported in molecular features, especially variations in microRNAs expression among lymphoma patients due to EBV infection [3, 4]. So that, the aim of this study was to analyze the presentation profile of commonly studied biomarkers in DLBCL, such as *MYC*, *BCL2*, *BCL6* chromosomic rearrangements and *MYC*, *BCL2*, *EZH2* and *TP53* expression in a series of EBV-DLBCL. In addition, we also considered miR-125b and miR-155b detection in these cases.

MATERIALS AND METHODS

Samples of 139 DLBCL cases were obtained from the files of the Cancer Institute of Ceará, Fortaleza-Brazil, from 2012-2017, with the institutional internal review board's approval. The pathological diagnoses were established according to the criteria of the 2016 World Health Organization classification, and determination of the germinal center (GC) or non-GC phenotype was based on the Hans algorithm. EBV detection was analyzed by RNA *in situ* hybridization (ISH) using complementary probes to the transcript EBER1 as previously described and EBV positivity was considered when at least 80% of neoplastic nuclei were stained [3, 5]. The representative tumor areas in each case were selected by microscopic examination to make a tissue microarray (TMA) with 2.0 mm punch using Manual Tissue Microarrayer QuickRay™ system (UNITMA Co., Bokjeong-ro, Korea), in which immunohistochemistry was performed on 2-µm tissue microarray sections using *EZH2* (Clone 11, dilution 1:50, Cell Marque™, Darmstadt, Germany), *MYC* (Clone Y69, dilution 1:50, Abcam, Shanghai, China), *TP53* (DO-7, ready-for-use, DAKO™, Glostrup, Denmark), *BCL2* (Clone 124, ready-for-use, DAKO™) and Ki-67 (Clone MIB-1, ready-for-use, DAKO™) as previously described. FISH dual-color break-apart probes (*MYC*/8q24 and *BCL6*/3q27) and dual-color fusion translocation probes (*BCL2*/18q21) (Abbott

Laboratories, Des Plaines, IL) were used for translocation detection as previously describe [6]. For miR-125b and miR-155b we performed ISH using specific LNA probes (hsa-miR-125b-5p miRCURY and hsa-miR-155b-5p miRCURY LNA miRNA detection probe, concentration at 40nM for both probes, Exiqon™, Vedbæk, Denmark) as previously described [7]. As we had a very limited sample (see results section), all our findings were presented descriptively, with no statistic analyses.

RESULTS

EBV was positive in 7.9% (11/139) in the series of selected cases. Although the number of EBV-positive cases is not high, this series of DLBCL, composed of 63.6% nGC subtype, showed peculiar aspects related to the studied markers (Table 1). Interestingly, the majority of nGC patients presented extranodal disease (5/7) and, conversely, only one CG case was extranodal (1/4).

Most of the tumors (72.7%; 9/11) had a high proliferative index (Ki-67>70%) independently on being GC or nGC subtype. miR-125b was detected in almost all of these cases, and conversely, only two cases were miR155b positive. It is noteworthy that BCL2 expression was observed in most cases (81.8%; 9/11), although *BCL2* translocation was a rare event (18.2%; 2/11). Similarly, MYC expression and *MYC* translocation was observed in only one and two patients, respectively. TP53, on the other hand, was commonly expressed (45.4%; 5/11) regardless the molecular subtype.

DISCUSSION

EBV-related DLBCL is rare in immunocompetent patients and its frequency varies geographically, comprising as low as 2% of cases in Europe *versus* approximately 9% in Latin American or Asian studies [3, 8]. The prevalence of EBV in this study was high considering western population but is similar to another Brazilian study (8.5%) [1, 9]. To our knowledge, this is the first description of a Northeastern Brazil series of immunocompetent EBV-DLBCL with biomarker expression profile.

Dojcinov et al [10] and Zhou et al [8] pointed EBV-DLBCL as a mainly extran-

odal disease. We found similar results considering the nGC cases, although in the general sample no difference was observed (6 extranodal x 5 nodal). However, our series presented more CG cases and these authors did not consider the molecular subtypes on their analyses, which may suggest that extranodal disease is a nGC EBV-DLBCL feature. As demonstrated by Bourbon et al [3], *BCL2* was widely expressed. Interestingly, this does not seem to happen due *BCL2* translocation, as it is a rare event in our series, but probably due a virus intrinsic mechanism. For instance, as discussed by Fu et al (2013), EBV nuclear and membrane proteins, specially LMP1, may induce *BCL2* overexpression by *BCL2* upregulation as the main immortalization effect on B-lymphomas [11].

Draws attention that miR-125b was detected in almost all high proliferative cases independently on CG versus nCG classification. Several studies have reported that miR-125b activates NF- κ B pathway in lymphoma [12, 13]. In addition, EBV might also mediate NF- κ B pathway through LMP1, resulting in high proliferative B-cell disorders [3, 10]. Interestingly, in our series, only two cases showed miR-155b expression. The literature is contradictory in EBV-induced changes of miR-155b expression in lymphoma. Although some authors points miR-155b as one of the most frequently expressed microRNA in DLBCL [14, 15], Imig et al. (2011), in accordance to our findings, showed that miR-155b was down-regulated by more than two folds in EBV-DLBCL compared to EBV-negative DLBCL patients [16]. It is noteworthy that these studies were performed on *in vitro* samples, in different populations and mostly applied gene sequencing and RT-PCR on their analyses, which albeit are more sensitive techniques, are less specific than ISH, what may explain our diverging results. As demonstrated in a previous study, EZH2 expression is consistently associated with high proliferative index in DLBCL [6]. Although only four cases were EZH2-positive in these selected EBV patients, it is interesting to note that in miR-125b negative cases with high Ki-67 index, EZH2 was expressed and, conversely, in miR-125b negative cases with low proliferative index, EZH2 is either negative. A few studies have shown that chronic activation of NF- κ B can also induce EZH2 expression in B-cell malignancies [17, 18]. This pathway might provide a mechanism through which tumor induced NF- κ B can inhibit tumor suppressor functions and promote tumorigenesis. Furthermore, the association miR-125b/EZH2 positive could be an additive factor.

This study is limited to a small sample size due the rarity of EBV-DLBCL

among the Brazilian population; nevertheless, it is representative of the EBV-DLGBL cases frequency treated at the Ceará Cancer Institute in a relatively short period of time (2012-2017). Others studies, multi-institutional, with a larger number of subjects are needed to a better understanding of EBV-DLBCL molecular landscape.

CONFLICTS OF INTEREST

The authors have no conflict of interest to declare.

REFERENCES

1. Poles WA, Nishi EE, de Oliveira MB, Eugênio AI, de Andrade TA, Campos AH, de Campos RR, Vassallo J, Alves AC, Scapulatempo Neto C, Paes RA. Targeting the polarization of tumor-associated macrophages and modulating mir-155 expression might be a new approach to treat diffuse large B-cell lymphoma of the elderly. *Cancer Immunology, Immunotherapy*. 2019 Feb;68(2):269-82.
2. Oliveira CC, Maciel-Guerra H, Kucko L, Hiramã EJ, Brilhante AD, Quevedo FC, da Cunha IW, Soares FA, Niero-Melo L, Dos Reis PP, Domingues MA. Double-hit lymphomas: clinical, morphological, immunohistochemical and cytogenetic study in a series of Brazilian patients with high-grade non-Hodgkin lymphoma. *Diagnostic pathology*. 2017 Dec;12(1):1-9.
3. Bourbon E, Maucourt-Boulch D, Fontaine J, Mauduit C, Sesques P, Safar V, Ferrant E, Golfier C, Ghergus D, Karlin L, Lazareth A. Clinicopathological features and survival in EBV-positive diffuse large B-cell lymphoma not otherwise specified. *Blood advances*. 2021 Aug 24;5(16):3227-39.
4. Soltani S, Zakeri A, Tabibzadeh A, Zakeri AM, Zandi M, Siavoshi S, Seifpour S, Farahani A. A review on EBV encoded and EBV-induced host microRNAs expression profile in different lymphoma types. *Molecular Biology Reports*. 2021 Feb;48(2):1801-17.
5. Ferrasi AC, Pinheiro NA, Rabenhorst SH, Caballero OL, Rodrigues MA, Carvalho F, Leite CV, Ferreira MV, Barros MA, Pardini MI. *Helicobacter pylori* and EBV in gastric carcinomas: methylation status and microsatellite instability. *World Journal of Gastroenterology: WJG*. 2010 Jan 1;16(3):312.
6. Neves Filho EH, Hirth CG, Frederico IA, Burbano RM, Carneiro T, Rabenhorst SH. EZH2 expression is dependent on MYC and TP53 regulation in diffuse large B-cell lymphoma. *Apmis*. 2020 Apr;128(4):308-15.
7. Munch-Petersen HD, Ralfkiaer U, Sjö LD, Hother C, Asmar F, Nielsen BS, Brown P, Ralfkiaer E, Grønbæk K. Differential expression of miR-155 and miR-21 in tumor and stroma cells in diffuse large B-cell lymphoma. *Applied immunohistochemistry & molecular morphology*. 2015 Mar 1;23(3):188-95.

8. Zhou Y, Xu Z, Lin W, Duan Y, Lu C, Liu W, Su W, Yan Y, Liu H, Liu L, Zhong M. Comprehensive genomic profiling of EBV-positive diffuse large B-cell lymphoma and the expression and clinicopathological correlations of some related genes. *Frontiers in oncology*. 2019 Jul 25;9:683.
9. Hofscheier A, Ponciano A, Bonzheim I, Adam P, Lome-Maldonado C, Vela T, Cortes E, Ortiz-Hidalgo C, Fend F, Quintanilla-Martinez L. Geographic variation in the prevalence of Epstein–Barr virus-positive diffuse large B-cell lymphoma of the elderly: a comparative analysis of a Mexican and a German population. *Modern Pathology*. 2011 Aug;24(8):1046-54.
10. Dojcinov SD, Fend F, Quintanilla-Martinez L. EBV-positive lymphoproliferations of B-T-and NK-cell derivation in non-immunocompromised hosts. *Pathogens*. 2018 Mar 7;7(1):28.
11. Fu Q, He C, Mao ZR. Epstein-Barr virus interactions with the Bcl-2 protein family and apoptosis in human tumor cells. *Journal of Zhejiang University SCIENCE B*. 2013 Jan;14(1):8-24.
12. Zheng Z, Qu JQ, Yi HM, Ye X, Huang W, Xiao T, Li JY, Wang YY, Feng J, Zhu JF, Lu SS. MiR-125b regulates proliferation and apoptosis of nasopharyngeal carcinoma by targeting A20/NF- κ B signaling pathway. *Cell death & disease*. 2017 Jun;8(6):e2855-.
13. Kim SW, Ramasamy K, Bouamar H, Lin AP, Jiang D, Aguiar RC. MicroRNAs miR-125a and miR-125b constitutively activate the NF- κ B pathway by targeting the tumor necrosis factor alpha-induced protein 3 (TNFAIP3, A20). *Proceedings of the National Academy of Sciences*. 2012 May 15;109(20):7865-70.
14. Linnstaedt SD, Gottwein E, Skalsky RL, Luftig MA, Cullen BR. Virally induced cellular microRNA miR-155 plays a key role in B-cell immortalization by Epstein-Barr virus. *Journal of virology*. 2010 Nov 15;84(22):11670-8.
15. Cao P, Zhang M, Wang L, Sai B, Tang J, Luo Z, Shuai C, Zhang L, Li Z, Wang Y, Li G. miR-18a reactivates the Epstein-Barr virus through defective DNA damage response and promotes genomic instability in EBV-associated lymphomas. *BMC cancer*. 2018 Dec;18(1):1-5.
16. Imig J, Motsch N, Zhu JY, Barth S, Okoniewski M, Reineke T, Tinguely M, Faggioni A, Trivedi P, Meister G, Renner C. microRNA profiling in Epstein–Barr virus-associated B-cell lymphoma. *Nucleic acids research*. 2011 Mar 1;39(5):1880-93.

17. Iannetti A, Ledoux AC, Tudhope SJ, Sellier H, Zhao B, Mowla S, Moore A, Hummerich H, Gewurz BE, Cockell SJ, Jat PS. Regulation of p53 and Rb links the alternative NF- κ B pathway to EZH2 expression and cell senescence. *PLoS genetics*. 2014 Sep 25;10(9):e1004642.
18. Di Pietro A. Epstein–Barr Virus Promotes B Cell Lymphomas by Manipulating the Host Epigenetic Machinery. *Cancers*. 2020 Oct 19;12(10):3037.

TABLE 1 - Distribution of the studied biomarkers expression profile over the 11 EBV-DLBCL cases

Patient	Subtype	MYC	BCL2	TP53	<i>MYC</i> t	<i>BCL2</i> t	<i>BCL6</i> t	EZH2	miR125b	miR155b	Ki-67
1	CG	Negative	Positive	Negative	Negative	Negative	Negative	Negative	Positive	Positive	70%
2	CG	Negative	Positive	Negative	Negative	Negative	Negative	Positive	Negative	Negative	70%
3	CG	Negative	Positive	Positive	Negative	Positive	Negative	Negative	Positive	Negative	85%
4	CG	Negative	Positive	Positive	Positive	Negative	Negative	Negative	Positive	Negative	70%
5	nCG	Negative	Positive	Positive	Negative	Negative	Negative	Positive	Positive	Negative	80%
6	nCG	Negative	Positive	Negative	Negative	Negative	Negative	Negative	Negative	Negative	30%
7	nCG	Negative	Positive	Positive	Negative	Negative	Negative	Negative	Positive	Negative	60%
8	nCG	Negative	Positive	Negative	negativo	Positive	Negative	Positive	Positive	Positive	85%

9	nCG	Positive	Negative	Positive	Negative	Negative	Negative	Positive	Negative	Negative	70%
10	nCG	Negative	Positive	Negative	Negative	Negative	Negative	Negative	Negative	Negative	20%
11	nCG	Negative	Negative	Negative	Positive	Negative	Negative	Negative	Positive	Negative	90%

CG - germinal center; nCG - non-germinal center; *MYCt* - *MYC* translocation; *BCL2t* - *BCL2* translocation; *BCL6t* - *BCL6* translocation

10 CONCLUSÕES

Objetivo 1- Associar a expressão proteica de MYC, BCL2, TP53 e EZH2 nos casos de LDGCB de acordo com a célula de origem e com a presença das translocações envolvendo MYC, BCL2 e BCL6.

Conclusão:

- A) a expressão de MYC foi associada ao subtipo nCG. Neste subgrupo, a expressão de EZH2, frequentemente observada nos casos de LDGCB avaliados, foi associada à expressão de MYC e TP53 e a um alto índice. Adicionalmente, a translocação de BCL2 foi associada à expressão de BCL2 somente no subtipo nCG.
- B) nos LDGCB tipo CG, a translocação de BCL6 parece ser uma característica marcante. Neste subgrupo, entretanto, a expressão de EZH2 está associada apenas à expressão de TP53;
- C) a translocação de MYC está associada à expressão de MYC em ambos os tipos moleculares e está envolvida na linfomagenese independente das células de origem.
- D) os diferentes padrões de expressão dependendo da classificação pela célula de origem, assim, descrevem uma via molecular envolvendo EZH2/MYC/TP53 característica do subgrupo nCG e, por outro lado, uma via EZH2/TP53 no subtipo CG.

Objetivo 2- Traçar o perfil de expressão de miR-125b e miR-155 nos casos de LDGCB.

Conclusão:

- A) nos LDGCB, a expressão de miR-125b foi frequente, sendo associada à elevada proliferação celular independentemente do subtipo molecular analisado. No subgrupo CG, entretanto, sua expressão foi associada à expressão de MYC;
- B) a expressão de miR-155b foi baixa e não apresentou nenhuma associação com os demais biomarcadores avaliados ou com o subtipo molecular.

Objetivo 3- Determinar a frequência da infecção por EBV nos casos de LDGCB, correlacionando os dados clínico-patológicos com os dados encontrados.

Conclusão:

- A) o EBV foi raramente expresso em LDGCB na nossa população;
- B) os casos de LDGCB associados ao EBV normalmente apresentam alto índice proliferativo e alta frequência da expressão de miR-125b.

Objetivo 4- Avaliar a importância prognóstica e o potencial terapêutico dos dados encontrados.

Conclusão:

- A) o fenótipo CG demonstrou ser fator prognóstico positivo independente, estando associado a maior sobrevida global;
- B) EZH2 não apresentou associações com a resposta primária ao R-CHOP ou com a sobrevida global dos casos de LDGCB avaliados;
- C) A expressão de miR-155b foi associada à progressão de doença em pacientes tratados com R-CHOP no subgrupo nCG e a expressão de BCL2 à ausência de resposta nesses pacientes;
- D) A expressão de TP53, miR-125b e miR-155b demonstraram tendência estatística como potenciais biomarcadores prognósticos no subgrupo nCG.

REFERÊNCIAS

ALIZADEH, A. A. *et al.* Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. **Nature**, v. 403, p. 503-511, 2000.

AMERICAN JOINT COMMITTEE ON CANCER (AJCC). **AJCC Cancer Staging Manual**. 8.ed. Chicago-IL: Springer, 2018.

CAO, Y. *et al.* Mutations or copy number losses of CD58 and TP53 genes in diffuse large B cell lymphoma are independent unfavorable prognostic factors. **Oncotarget**, v. 7, n. 50, p. 83294-83307, 13 dez. 2016. doi: 10.18632/oncotarget.13065.

CHESON, B. D. The International Harmonization Project for response criteria in lymphoma clinical trials. **Hematol Oncol Clin North Am**, v. 21, p. 841–54, 2007.

COURVILLE, E. L. *et al.* EBV-negative monomorphic B-cell post-transplant lymphoproliferative disorders are pathologically distinct from EBV-positive cases and frequently contain TP53 mutations. **Mod Pathol**, v. 29, n. 10, p. 1200-1211, 2016. doi: 10.1038/modpathol.2016.130.

ENOMOTO, Y. *et al.* Emu/miR-125b transgenic mice develop lethal B-cell malignancies. **Leukemia**, v. 25, n. 12, p. 1849-1856, dez. 2011.

FAN, Jian-Bing. Next-Generation MicroRNA Expression Profiling Technology. **Methods Mol Biol**. v. 822, p. 67-84, 2012. doi:10.1007/978-1-61779-427-8 (Chapter 5), p.67-84. doi:10.1007/978-1-61779-427-8_5.

FEDCHENKO, N.; REIFENRATH, J. Different approaches for interpretation and reporting of immunohistochemistry analysis results in the bone tissue - a review. **Diagn Pathol**, v. 9, p.221, 2014.

FERRASI, A. C. *et al.* Helicobacter pylori and EBV in gastric carcinomas: methylation status and microsatellite instability. **World J Gastroenterol**, v. 16, n. 3, p. 312-319, 21 jan. 2010.

FREEDMAN, A. S.; ASTER, J. C. **Epidemiology, clinical manifestations, pathologic features, and diagnosis of diffuse large B cell lymphoma**. 2019. Disponível em: https://www.uptodate.com/contents/epidemiology-clinical-manifestations-pathologic-features-and-diagnosis-of-diffuse-large-b-cell-lymphoma?source=related_link. Acesso em: 25 maio 2022.

FREEDMAN, A. S.; ASTER, J. C. **Prognosis of diffuse large B cell lymphoma**. 2020. Disponível em: <https://www.uptodate.com/contents/prognosis-of-diffuse-large-b-cell-lymphoma>. Acesso em 25 maio 2022.

FREEDMAN, A. S.; FREIDBERG, J. W.; ASTER, J. C. **Clinical presentation and initial evaluation of non-Hodgkin lymphoma**. Disponível em: https://www.uptodate.com/contents/clinical-presentation-and-initial-evaluation-of-non-hodgkin-lymphoma?search=Clinical%20presentation%20and%20initial%20evaluation%20of%20non-Hodgkin%20lymphoma&source=search_result&selectedTitle=1~150&usage_type=default&display_rank=1. 2022. Acesso em 25 maio 2022.

GLOBOCAN. **Non-Hodgkin Lymphoma Fact Sheet**. 2020. Disponível em: <https://gco.iarc.fr/today/data/factsheets/cancers/34-Non-hodgkin-lymphoma-fact-sheet.pdf>. Acesso em: 9 jun 2022.

HOELLER, S. *et al.* Epstein-Barr virus-positive diffuse large B-cell lymphoma in elderly patients is rare in Western populations. **Hum Pathol**, v. 41, n. 3, p. 352-357, mar. 2010. doi: 10.1016/j.humpath.2009.07.024.

HOFSCHEIER, A. *et al.* Geographic variation in the prevalence of Epstein-Barr virus-positive diffuse large B-cell lymphoma of the elderly: a comparative analysis of a Mexican and a German population. **Mod Pathol**, v. 24, n. 8, p. 1046-1054, ago. 2011. doi: 10.1038/modpathol.2011.62.

HORN, H. *et al.* MYC status in concert with BCL2 and BCL6 expression predicts outcome in diffuse large B-cell lymphoma. **Blood**, v. 121, p. 2253-2263, 2013.

INSTITUTO NACIONAL DE CÂNCER (INCA). **Estimativas 2020**. Disponível em: <http://www.inca.gov.br/estimativa/2020>. Acesso em: 9 jun. 2022.

JAFFE, E. S. *et al.* **Hematopathology**. 2.ed. Philadelphia: Elsevier, 2017.

JOHNSON, N. A.; SLACK, G. W.; SAVAGE, K. J. *et al.* Concurrent expression of MYC and BCL2 in diffuse large B-cell lymphoma treated with rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone. **J Clin Oncol**, v. 30, n. 28, p. 3452-3459, 2012.

JØRGENSEN, L. K. *et al.* MicroRNAs as novel biomarkers in diffuse large B-cell lymphoma--a systematic review. **Dan Med J**, v. 62, n. 5, 2015. PIMD: 26050834

KUMAR, M. *et al.* Negative regulation of the tumor suppressor p53 gene by microRNAs. **Oncogene**, v. 30, p. 843-853, 2011.

LARRABEITI-ETXEBARRIA, A. *et al.* Systematic Review of the Potential of MicroRNAs in Diffuse Large B Cell Lymphoma. **Cancers**, v. 11, n. 2, p. 144, 2019. doi:10.3390/cancers11020144.

LAWRIE, C. H. *et al.* Expression of microRNAs in diffuse large B cell lymphoma is associated with immunophenotype, survival and transformation from follicular lymphoma. **J. Cell. Mol. Med.**, v. 13, n. 7, p. 1248–1260, 2009. doi:10.1111/j.1582-4934.2008.00628.x.

LAWRIE, C. H. MicroRNAs and lymphomagenesis: A functional review. **Br J Haematol**, v. 160, p. 571-581, 2013.

LENZ, G. *et al.* Stromal gene signatures in large-B-cell lymphomas. **N Engl J Med**, v. 359, p. 2313-2323, 2008.

LI, C. *et al.* Copy number abnormalities, MYC activity and the genetic fingerprint of normal B cells mechanistically define the microRNA profile of diffuse large B-cell lymphoma. **Blood**, v. 113, p. 6681-6690, 2009.

LIU, Y. *et al.* EZH2 overexpression in primary gastrointestinal diffuse large B-cell lymphoma and its association with the clinicopathological features. **Hum Pathol**, v. 64, p. 213-221, 2017.

MUNCH-PETERSEN, H. D. *et al.* Differential Expression of miR-155 and miR-21 in Tumor and Stroma Cells in Diffuse Large B-Cell Lymphoma Cell Lymphoma. **Appl Immunohistochem Mol Morphol**, v. 23, n. 3, p. 188-195, mar. 2015.

NI, H. *et al.* MicroRNAs in diffuse large B-cell lymphoma. **Oncol Lett**, v. 11, n. 2, p. 1271-1280, fev. 2016.

NOGAI, H.; DÖRKEN, B.; LENZ, G. Pathogenesis of non-Hodgkin's lymphoma. **J Clin Oncol**, v. 29, n. 14, p. 1083-1811, maio 2011. doi: 10.1200/JCO.2010.33.3252

OKEN, M.M. *et al.* Toxicity and response criteria of the Eastern Cooperative Oncology Group. **Am J Clin Oncol**. v. 5, n. 6, p. 649-655, 1982.

PARK, S. *et al.* The impact of Epstein-Barr virus status on clinical outcome in diffuse large B-cell lymphoma. **Blood**, v. 110, n. 3, p. 972-978, 1 ago. 2007.

PASQUALUCCI, L.; DALLA-FAVERA, R. The genetic landscape of diffuse large B-cell lymphoma. **Semin Hematol**, v. 52, n. 2, p. 67-76, 2015.

SARIKOC, F. *et al.* (). An automated prognosis system for estrogen hormone status assessment in breast cancer tissue samples. **Turkish Journal of Electrical Engineering and Computer Sciences**, v. 21, n. 4, p. 1199-1121, 2013. doi: 10.3906/elk-1111-10.

SWERDLOW, S. H. *et al.* The 2016 revision of the World Health Organization classification of lymphoid neoplasms. **Blood**, v. 127, n. 20, p. 2375-2390, 19 maio 2016. doi: 10.1182/blood-2016-01-643569.

SWERDLOW, S. H. Diagnosis of 'double hit' diffuse large B-cell lymphoma and B-cell lymphoma, unclassifiable, with features intermediate between DLBCL and Burkitt lymphoma: when and how, FISH versus IHC. **Hematology Am Soc Hematol Educ Program**, n. 1, p. 90-99, 2014.

THE NON-HODGKIN'S LYMPHOMA CLASSIFICATION PROJECT. A clinical evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. **Blood**, v. 89, n. 11, p. 3909-3918, 1997.

TIAN, X. *et al.* Differential expression of enhancer of zeste homolog 2 (EZH2) protein in small cell and aggressive B-cell non-Hodgkin lymphomas and differential regulation of EZH2 expression by p-ERK1/2 and MYC in aggressive B-cell lymphomas. **Mod Pathol**, v. 29, p. 1050-1057, 2016.

TIAN, X.; XU, J.; DORFMAN, D. M. Utility of combined EZH2, p-ERK1/2, p-STAT, and MYC expression in the differential diagnosis of EZH2-positive Hodgkin Lymphomas and related large B-cell lymphomas. **Am J Surg Pathol**, v. 43, p. 102-109, 2019.

TOLEDO, F.; WAHL, G. M. Regulating the p53 pathway: in vitro hypotheses, in vivo veritas. **Nat. Rev. Cancer**, v. 6, p. 909-923, 2006.

UNER, A. *et al.* The presence of Epstein-Barr virus (EBV) in diffuse large B-cell lymphomas (DLBCLs) in Turkey: special emphasis on 'EBV-positive DLBCL of the elderly'. **APMIS**, v. 119, n. 4-5, p. 309-316, abr. 2011. doi: 10.1111/j.1600-0463.2011.02736.x.

VENTURA, A. *et al.* Restoration of p53 function leads to tumour regression in vivo. **Nature**, v. 445, p. 661-665, 2007.

XUE, W. *et al.* Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. **Nature**, v. 445, p. 656-660, 2007.

XU-MONETTE *et al.* Mutational profile and prognostic significance of TP53 in diffuse large B-cell lymphoma patients treated with R-CHOP: report from an International

DLBCL Rituximab-CHOP Consortium Program Study. **Blood**, v. 120, n. 19, p. 3986-3996, 2012.